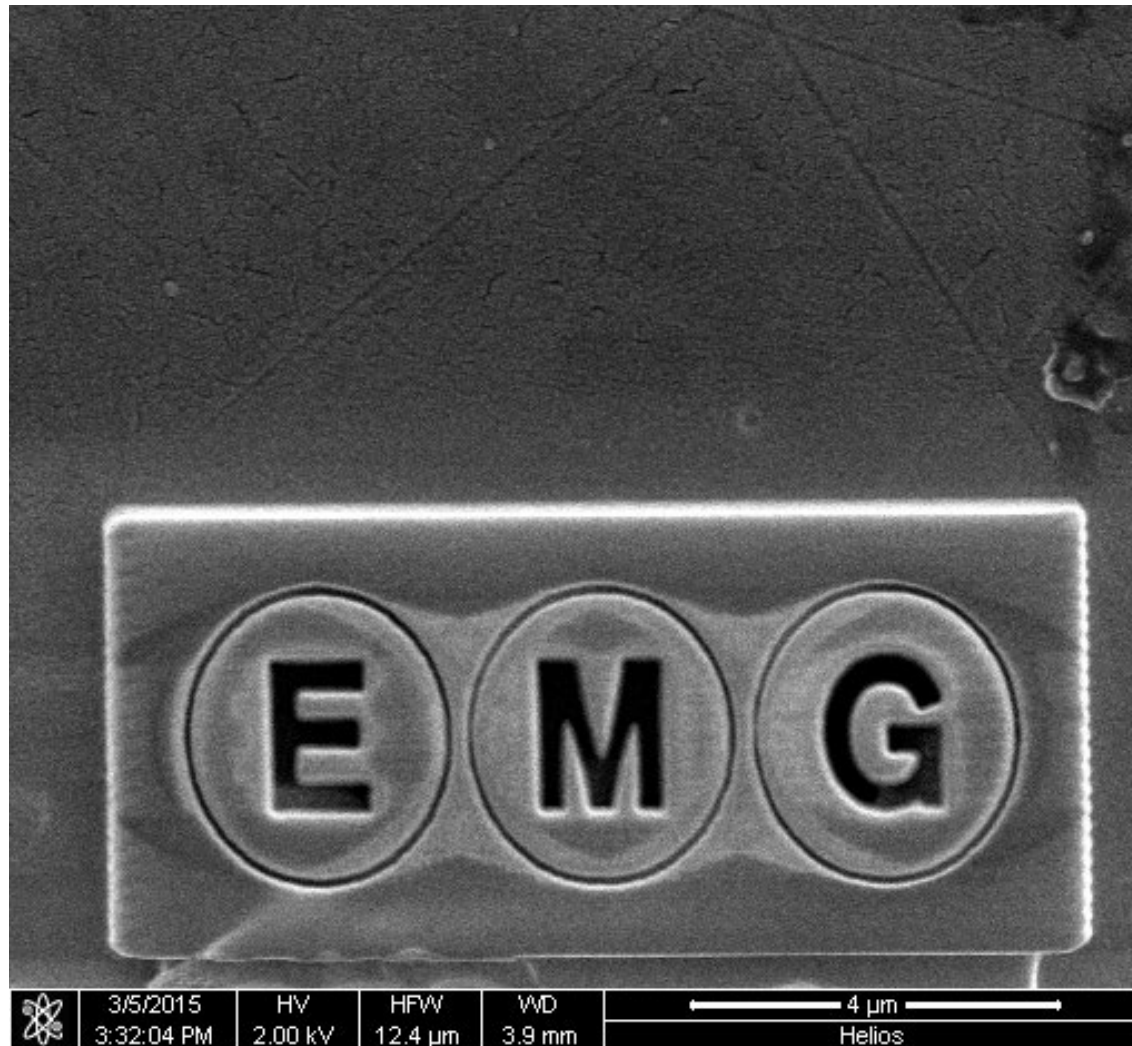


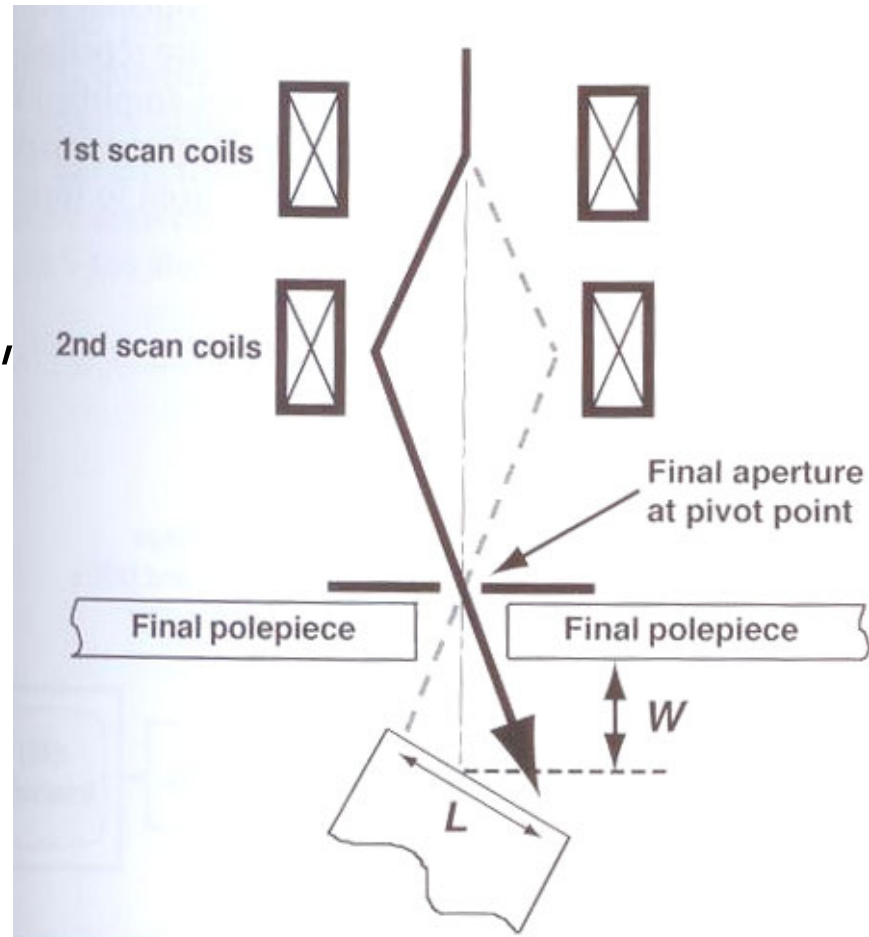
FIB/SEM

Feb. 29, 2016



SEM Basics

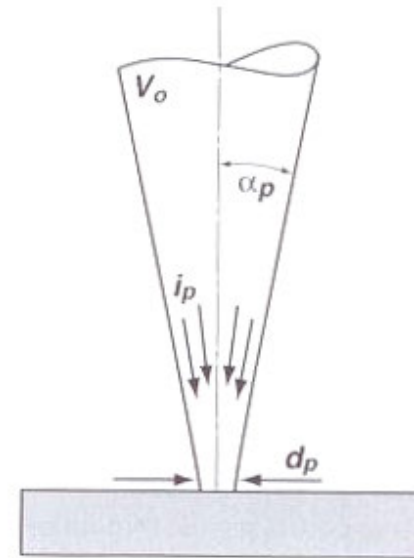
- ◆ Electron probe is focused to a sharp point
- ◆ The probe is scanned across the specimen point by point, with each point producing signal
- ◆ Scan coils deflect beam to move across sample



Goldstein et al, 2003

SEM Beam: probe size

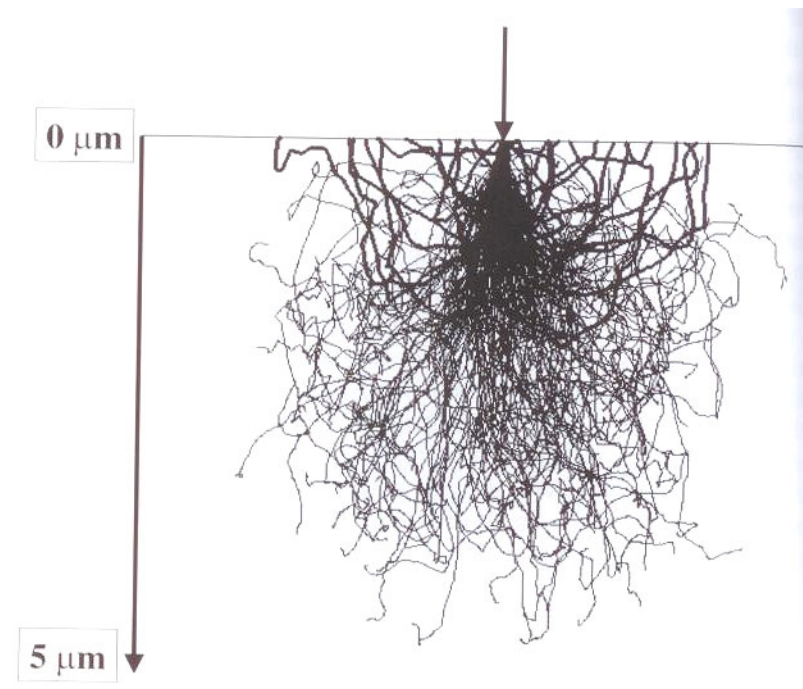
- ◆ Ideally want as small a probe as possible, relative to pixel size
- ◆ Probe size is determined by voltage, current, divergence angle
- ◆ Lens distortions
 - ◆ Spherical aberration (focus different at center and edge of lens) – instrument dependent
 - ◆ Aperture diffraction
 - ◆ Astigmatism (user correctable)
 - ◆ Chromatic aberration – voltage dependent



Goldstein et al, 2003

Beam-Specimen Interaction

- Monte Carlo simulation of a 20 keV beam in Si
 - Dark traces: electrons which left the sample (BSE's)
- Electrons may be scattered elastically or inelastically
- Probability of elastic scattering $\sim Z^2$
- Inelastic scattering:
 - Secondary electrons
 - X-rays

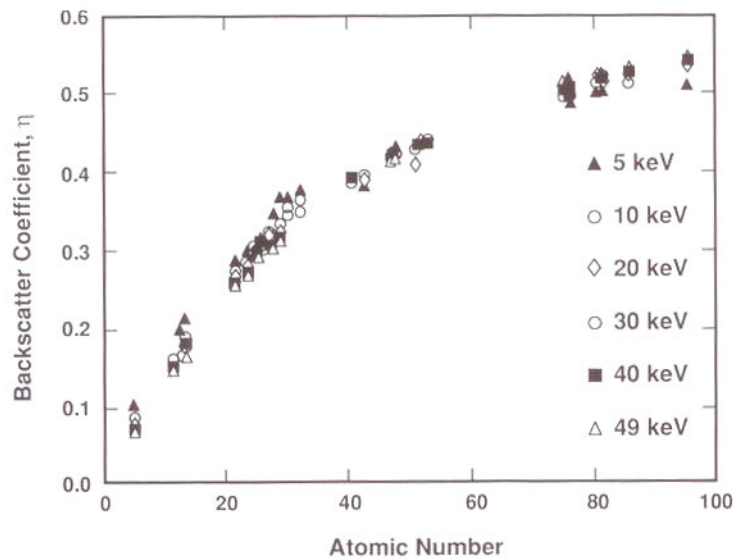


Goldstein et al, 2003

BSE's and SE's

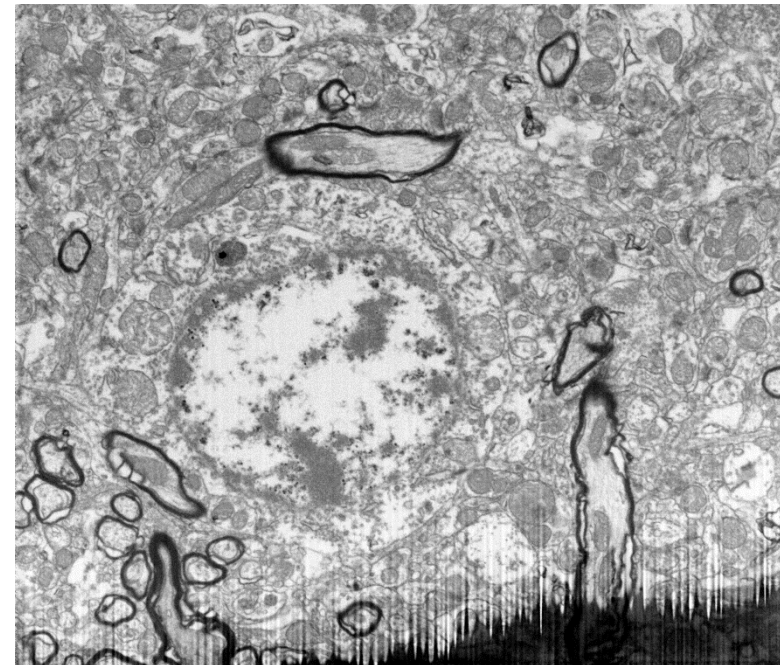
BSE efficiency is material dependent, voltage independent

Fraction of e's that backscatter

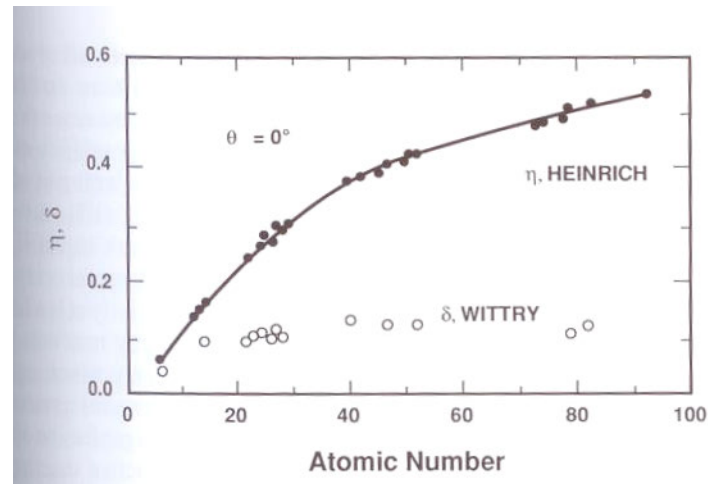


Goldstein et al, 2003

BSE's give contrast between light and heavy elements



Specimen Dependence of BSE, SE

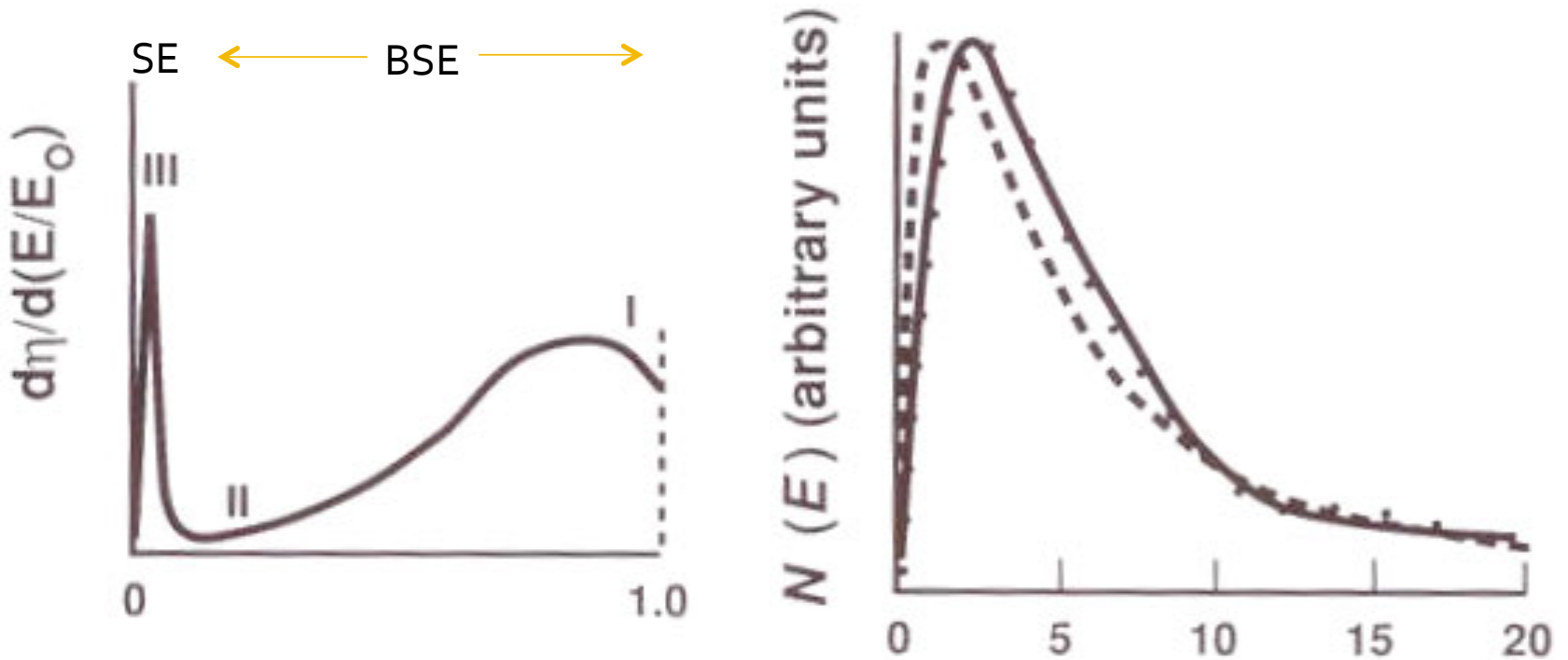


30 keV

SE's are less sensitive to atomic number than BSE's
(may be more sensitive at lower beam energies)

Goldstein et al, 2003

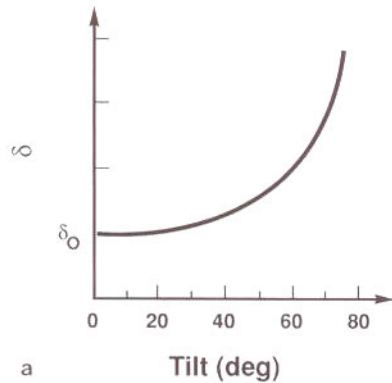
Energies of SE and BSE



SE's have much lower energy (majority < 20 eV)

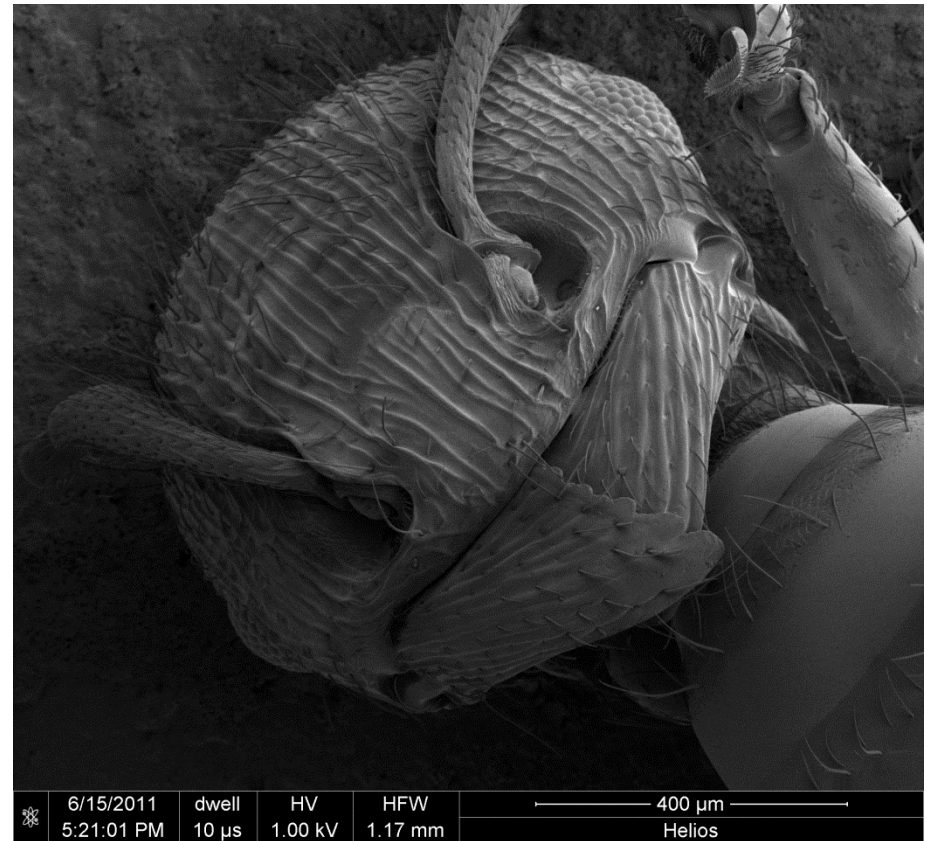
Goldstein et al, 2003

Angular Dependence of SE's



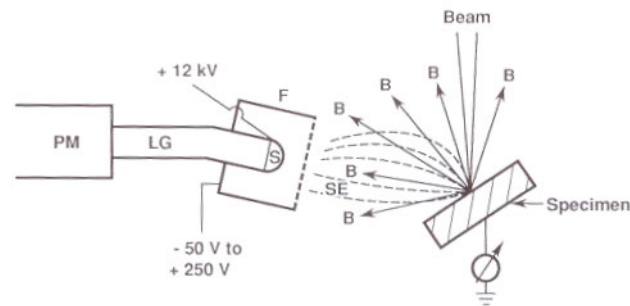
Goldstein et al, 2003

SE's also give topographic information



Detection of BSE's, SE's

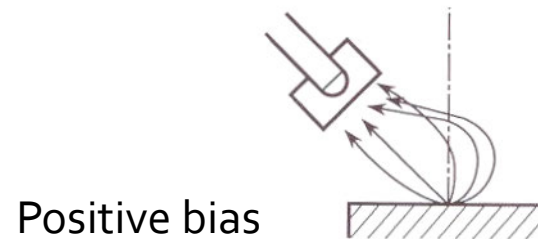
Everhart-Thornley (ET) Detector



Electrons strike scintillator, releasing photons
Photons travel to photomultiplier tube
Eventually converted to electric signal, storing intensity values
Combined SE/BSE detector

Apply bias

- ◆ Bias can be applied to detector, directing electrons toward or away from it
 - ◆ Negative bias (< -50 V): detect only BSE's
 - ◆ Positive bias: collect more SE's, indirect BSE's : greater total signal



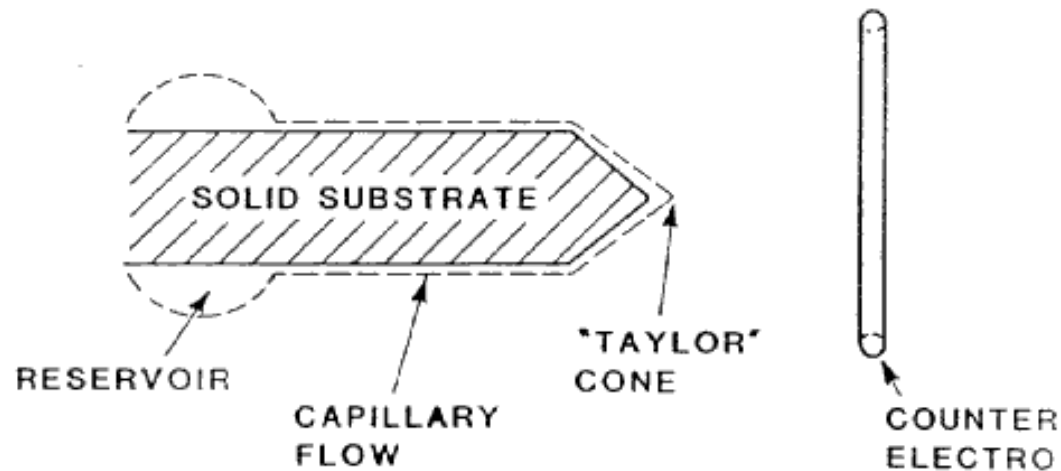
SEM summary

- Images formed by scanning points across sample
- For higher resolution, want to minimize both probe size and interaction volume
 - Low voltage operation
 - But still need enough signal for detection
- Use Through-lens detector for higher resolution imaging
- Apply negative bias to detect mainly BSE's

FIB Operation

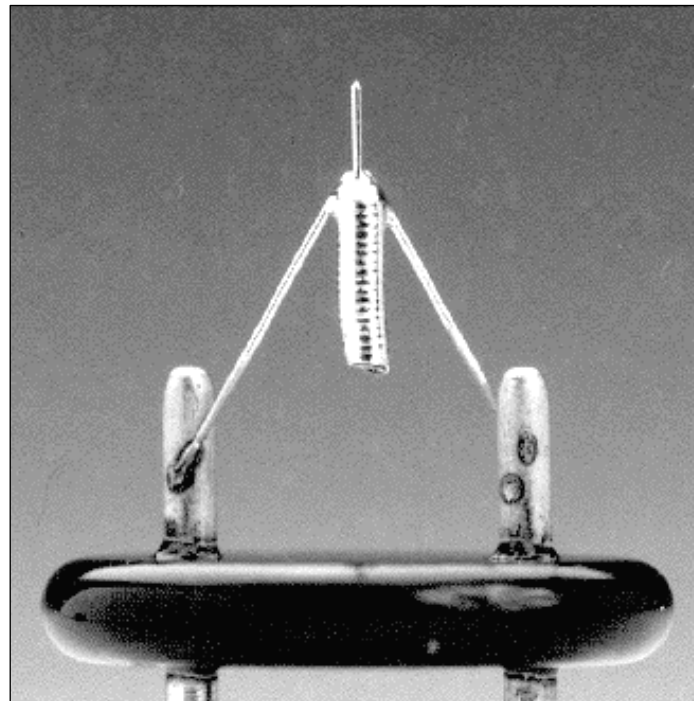
Basic Mechanism

- ◆ Liquid Flow from Reservoir
- ◆ Ion Formation
- ◆ External Beam Interactions



Gallium is the Most Popular LMIS

- ◆ A liquid metal
- ◆ Room temperature operation
- ◆ Long lived (500-1500 hr sources)
- ◆ High vacuum compatible
- ◆ Large ion for sputtering

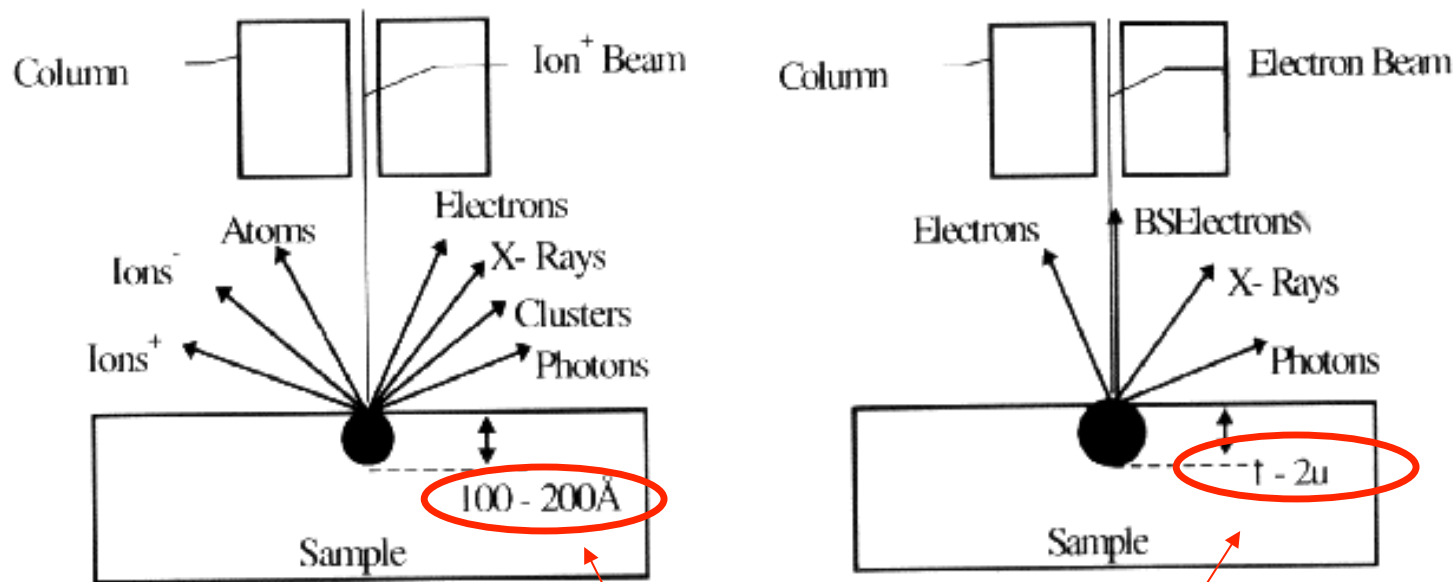


Ion Column

- ◆ Source - LMIS at top
- ◆ Focusing Optics
 - Use Electrostatic lenses since ions are heavier than electrons.
- ◆ Deflection Electronics/Pattern Board
- ◆ High-speed Blanking
 - Need to prevent milling while blanking

Using the System

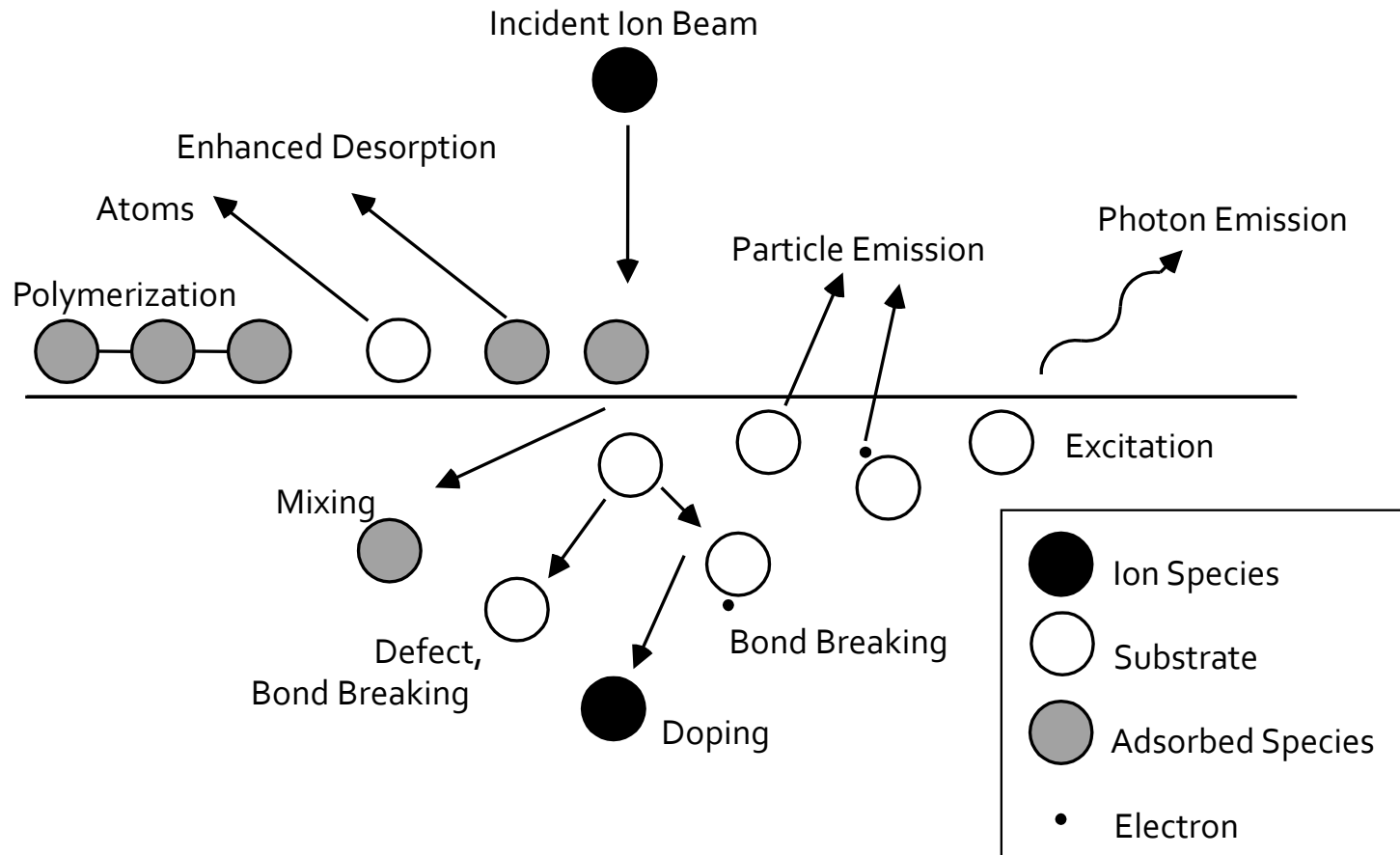
◆ Beam Interactions



Note difference in interaction volume

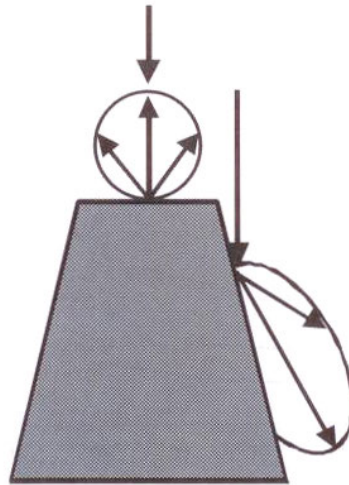


Ion Beam to Sample Interactions

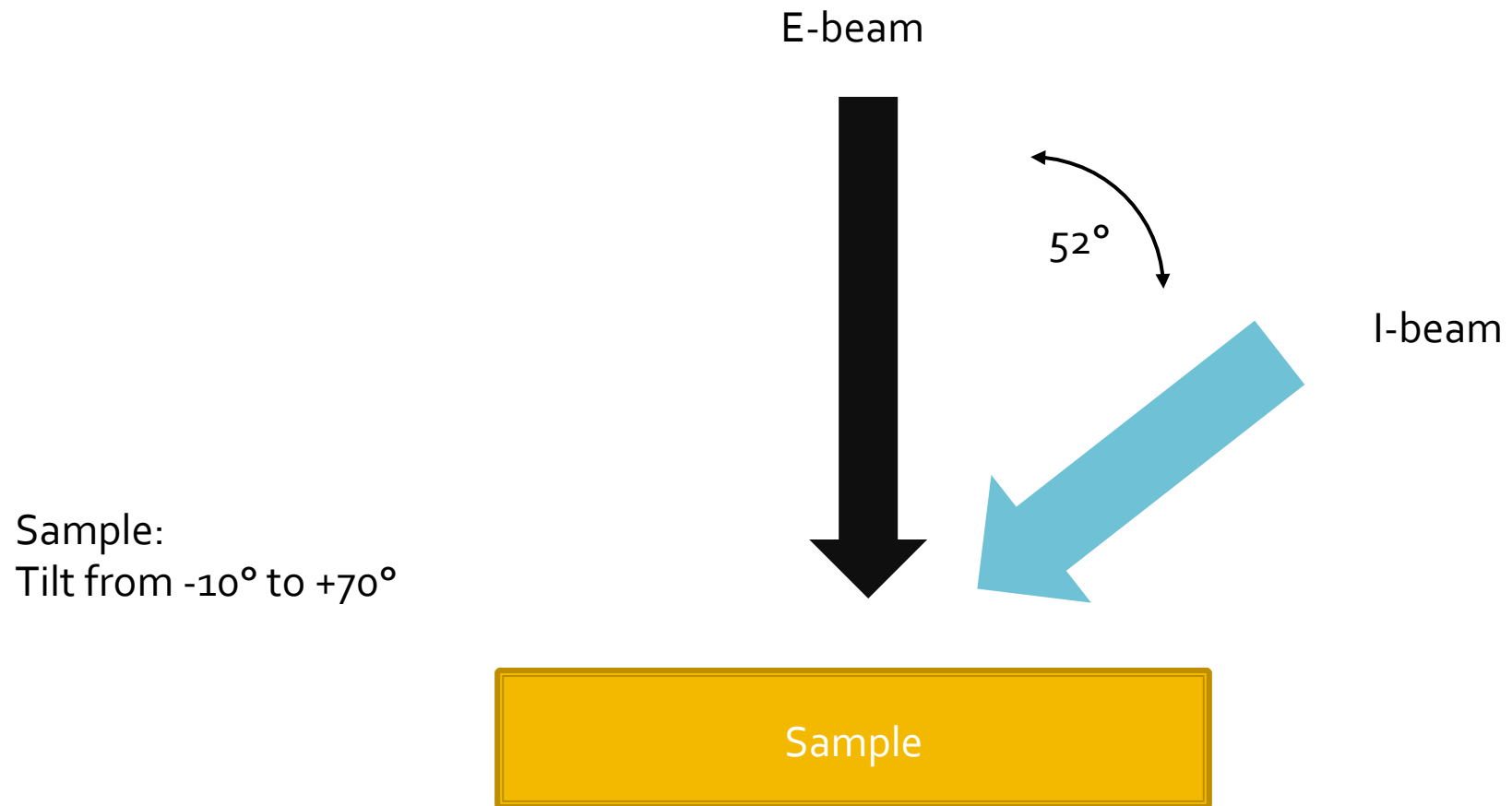


Sputtered Particles

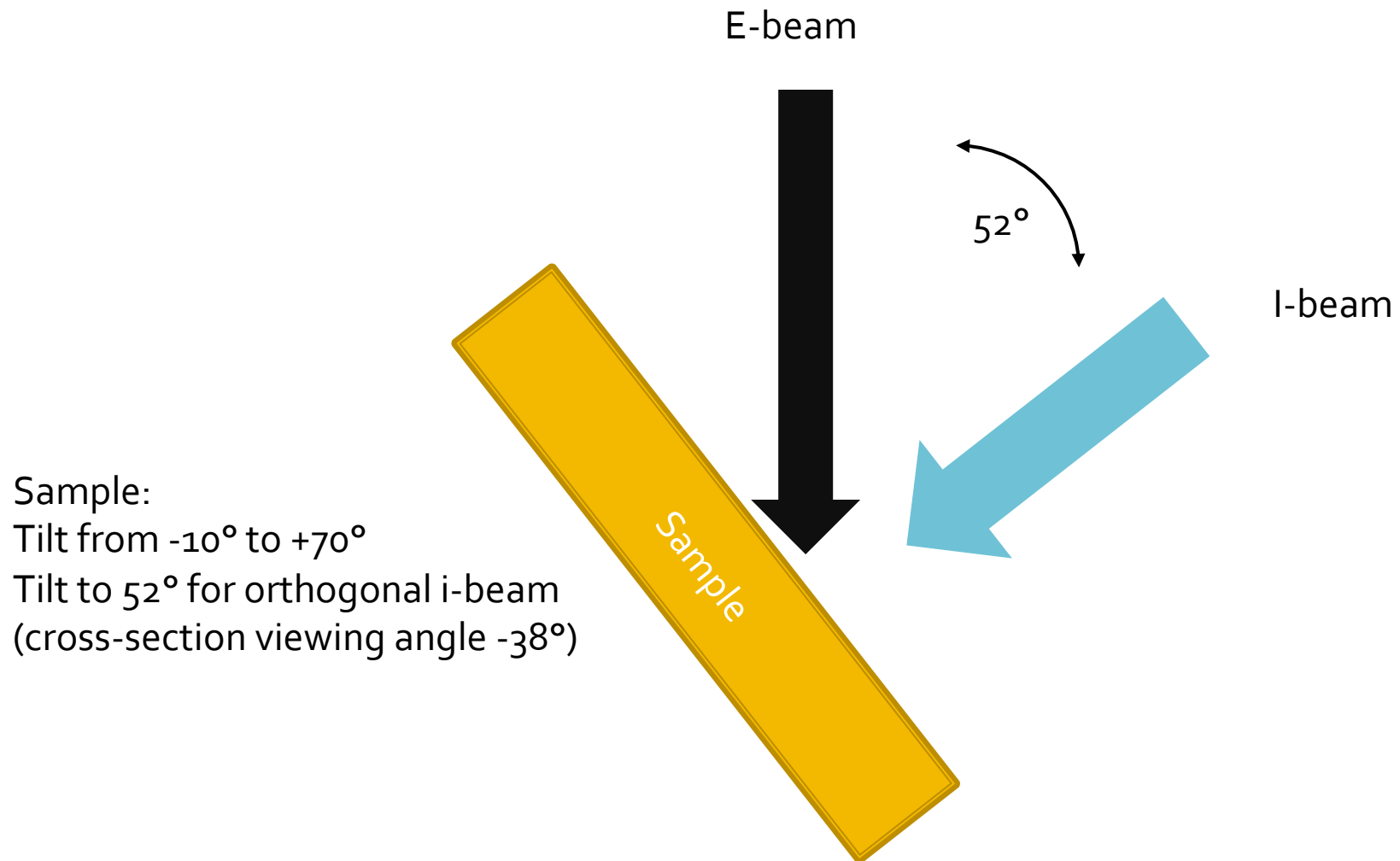
Sputtered Particle Ejection Behavior



Geometry



Geometry



Applications to Resin- embedded tissue

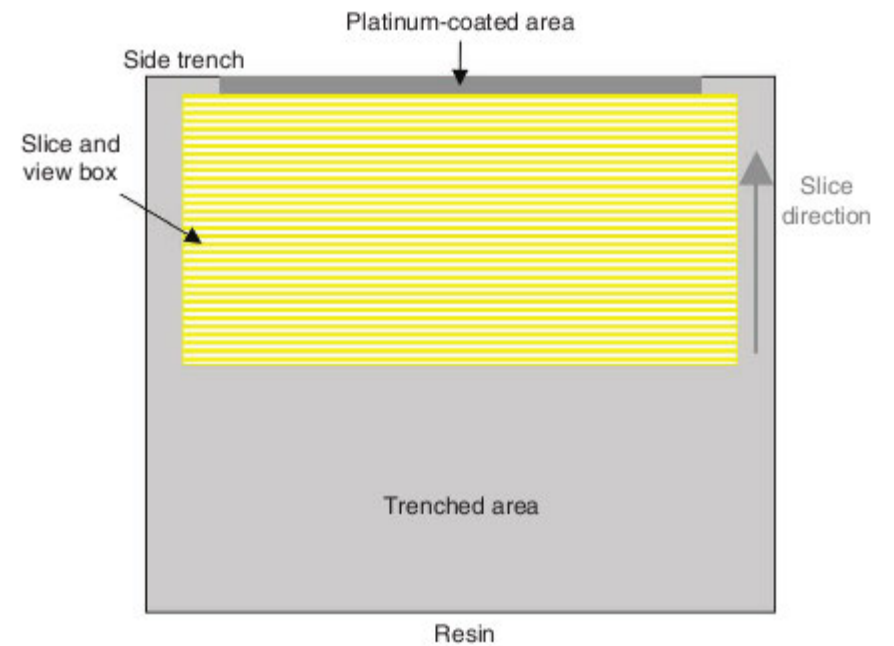
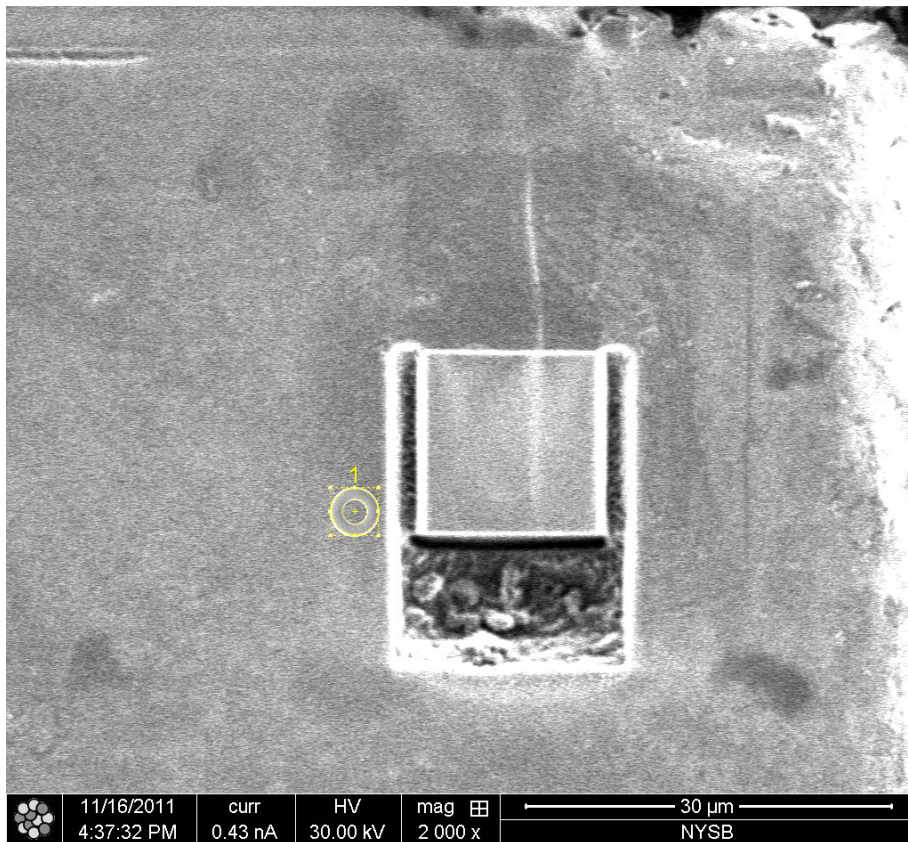
Tissue or Cells

- Sample Prep
 - High Pressure frozen (optional)
 - Chemically fixed, freeze substituted
 - Resin embedded
 - En bloc staining
 - OsO_4 , U Ac, Pb citrate
 - Osmium impregnation (OTO)
 - Want to make samples more conductive, more heavily stained
 - Thin conductive layer (C, Pt, Au-Pd) coated just before insertion

Imaging conditions

- Low voltage (2 keV or less)
 - Want to image only the surface
 - Minimal depth penetration (slice as thin as 10 nm)
 - No topographic information
 - Elemental contrast (C vs Os)
 - Through-lens detector for highest resolution
 - BSE mode (positive bias)
 - Stained parts will show up as bright on dark

Milling Samples



Bushby et al, 2011

Set up for Slice and View

The screenshot displays the xT microscope Control v4.5.3 build 2378 - supervisor (Supervisor) interface. The main window is divided into four quadrants, each showing a different view of the specimen. The top-left view shows a cross-section of a specimen with a yellow crosshair. The top-right view shows a similar cross-section from a different angle. The bottom-left view shows a top-down view of the specimen with a yellow crosshair. The bottom-right view shows a top-down view of the specimen with a yellow crosshair and a green arrow pointing to a feature labeled "4 mm Dynamic Focus: -38°".

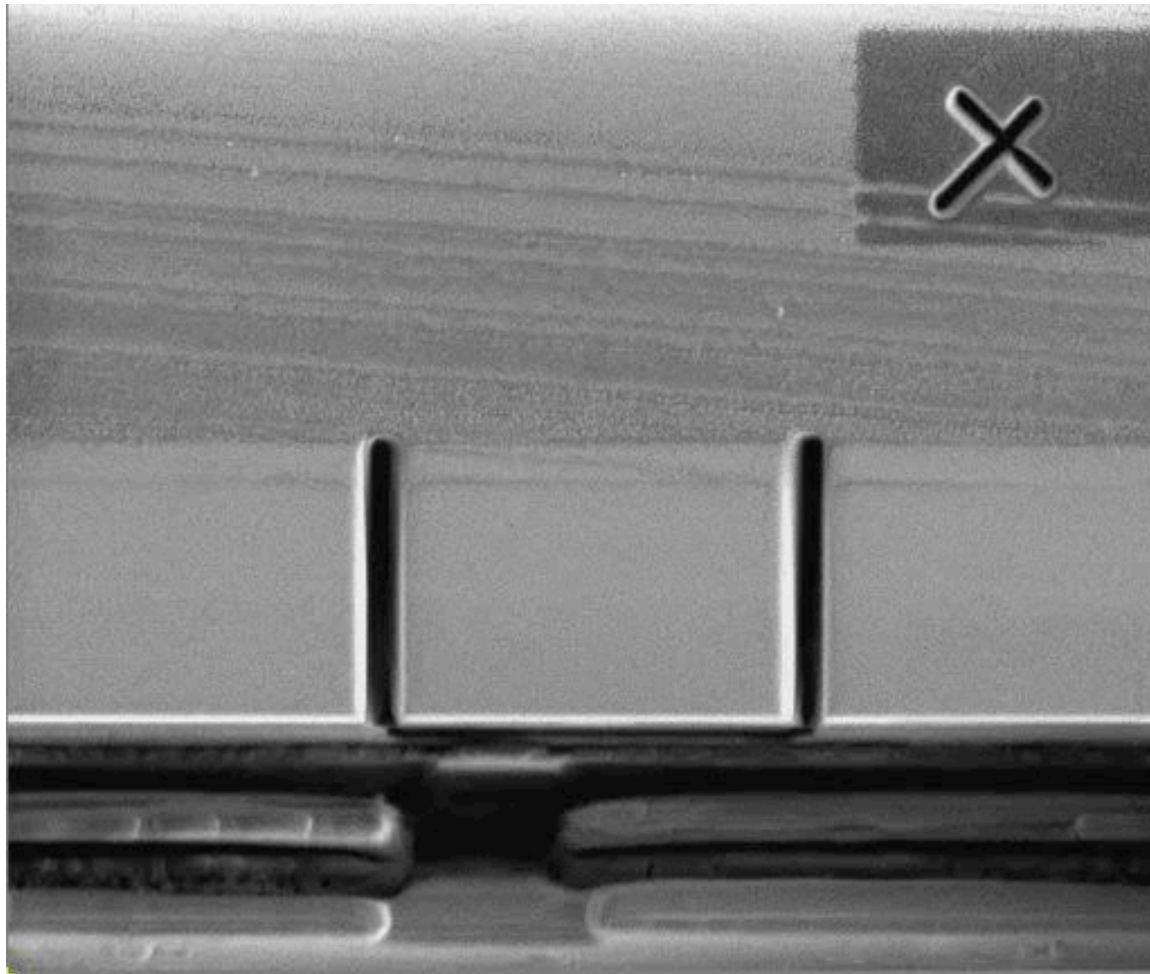
The interface includes a menu bar (File, Edit, Detectors, Scan, Beam, Patterning, Stage, Tools, Window, Help) and a toolbar with various icons. The right-hand side of the interface contains a control panel with the following sections:

- Vacuum:** Pump, Vent
- System:** Wake Up, Sleep
- Column:** Beam Current (Beam On, 80 pA), High Voltage (30.00 kV)
- Magnification:** Magnification (3500 x)
- Beam:** Stigmator, Beam Shift
- Rotation:** Scan Rotation (0.0°)
- Beam Deceleration:** Stage Bias (1000 V)
- Detectors:** Contrast (59.2), Brightness (43.9)
- Status:** Specimen Current (0.86 pA), Ion Beam Current (1.37 nA), Chamber Pressure (1.18e-5 mbar)

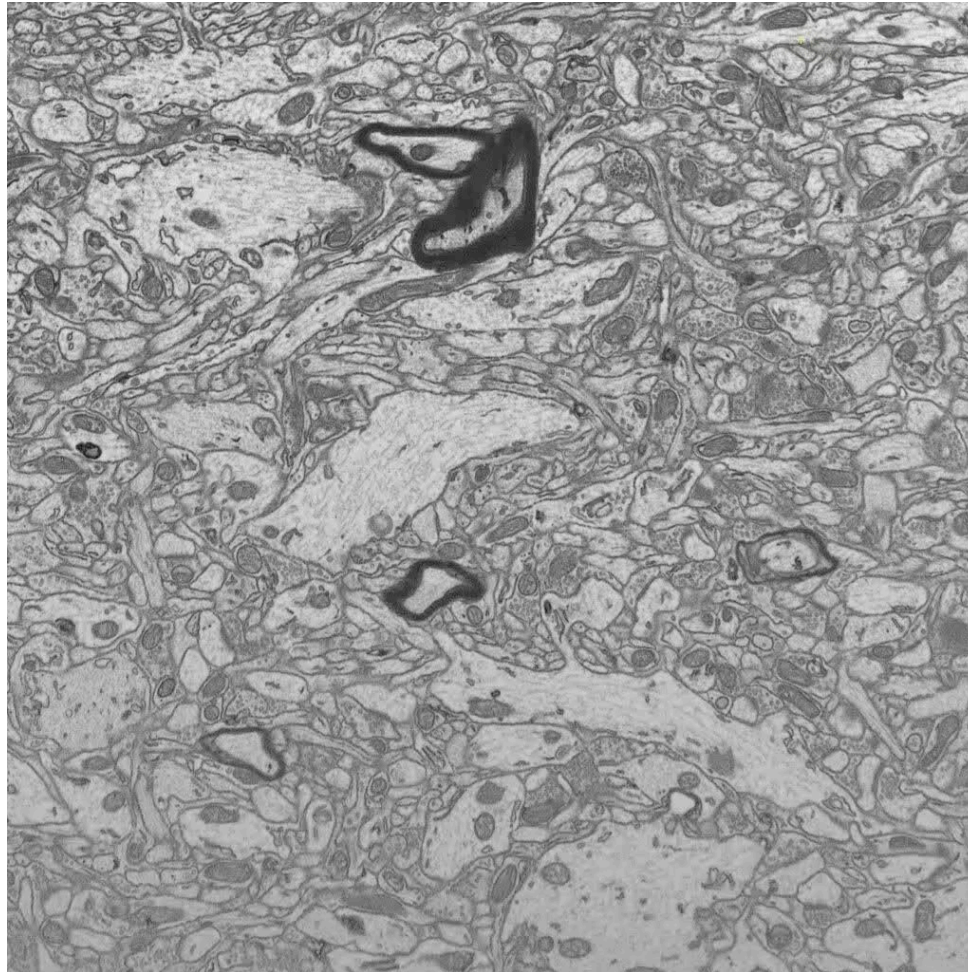
The bottom status bar displays the following information:

Date/Time	HV	HFWD	WD	Scale	Date/Time	det	x	y	tilt
2/22/2012 12:45:36 PM	2.00 kV	29.8 μm	4.0 mm	10 μm	2/22/2012 12:34:24 PM	CCD	0.7432 mm	-0.9972 mm	52°
2/15/2012 4:14:39 PM	25.00 kV	2.52 mm	4.0 mm	500 μm					

Milling: i-beam view



Example Movie: Neural Tissue



Ideal workflow

- Samples stained and embedded
- Thin slice for overall map – make easier to find features (LM or ultrathin EM)
- Face of block polished
- Set up for slice and view (1 day)
- Collect slices (1-5 days)
- Align, process (IMOD, Amira) (1 day)
- Segmentation (IMOD, Amira) (weeks-months)