

Welcome to electron microscopy at SEMC

- 1. Welcome new students
- 2. Course logistics
- 3. Introduction to EM and Roundtable
- 4. Tour of the facility

















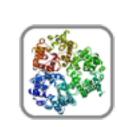












Protein Production COMPPÅ























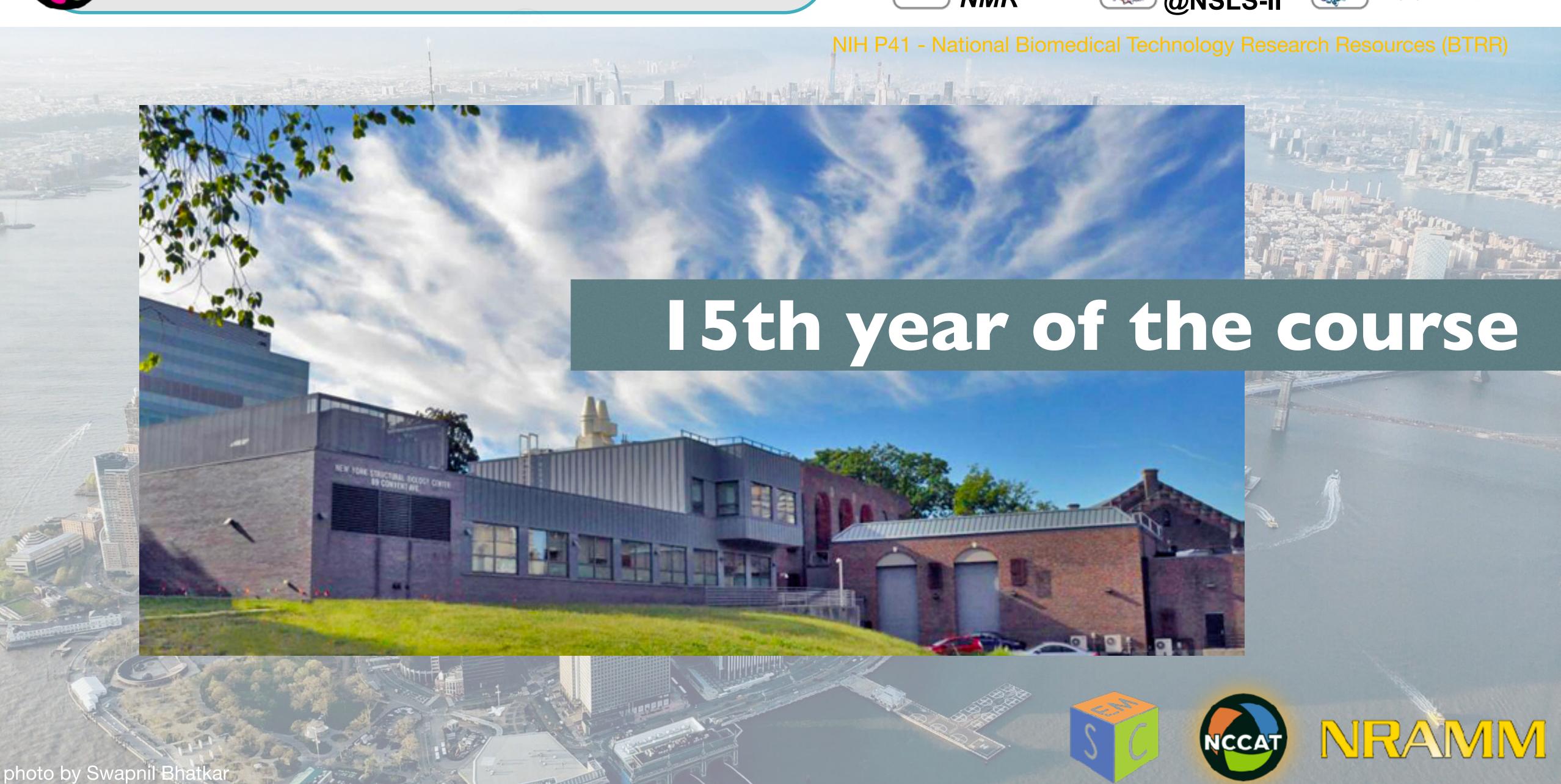


NMR



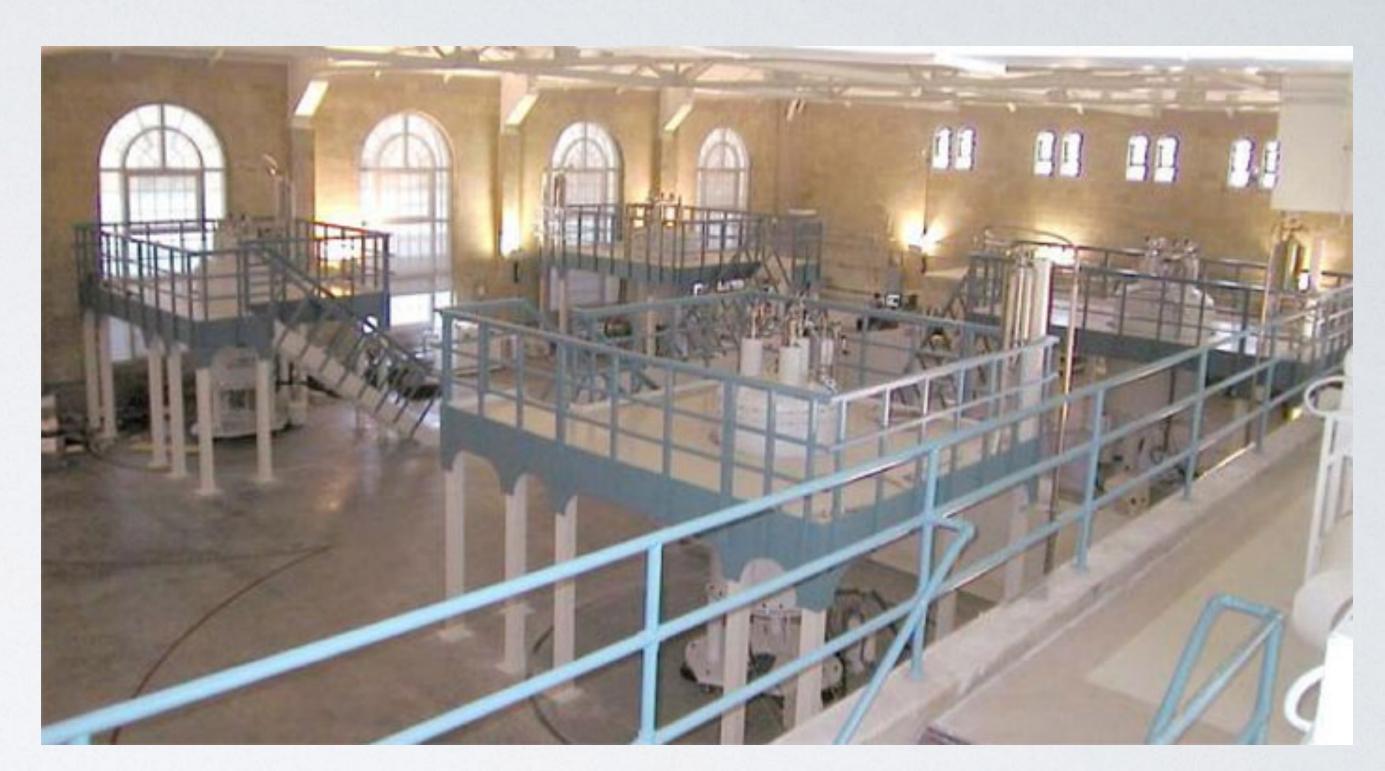


Protein Production COMPPÅ



Course logistics

- Questionnaire
 - email list
 - SEMC Winter-Spring EM Course Handbook
- Active research facility
 - public areas



Course logistics: main website

semc.nysbc.org/the-winter-spring-2020-em-course/



SIMONS ELECTRON MICROSCOPY CENTER



USER RESOURCES

PUBLICATIONS INSTRUMENTATION **NEWS & EVENTS**

Q

EVENTS

Upcoming Events

News and Past Events

Forums

Workshops and Courses

Workshops and Symposia at **Outside Organizations**

The Winter-Spring 2020 EM Course

General registration for the 2020 winter course is closed. If you registered for this course through your institution's registrar (with a cutoff in January), then please let us know as we have separate reserved spots for students taking the course for academic credit.

About the course

Electron microscopy in combination with image analysis is increasingly powerful in producing 3D structures of individual molecules and large macromolecular complexes that are unapproachable by other methods. This course is focused on the concepts and theories behind electron microscopy and will be taught in a reverse classroom format based on Grant Jensen's online course (Getting Started in Cryo-EM from Caltech). Students will be responsible for watching these online lectures prior to class. Each week guest lecturers and SEMC staff lead discussions on the practice of solving molecular structures by electron microscopy.

The course will be held at the New York Structural Biology Center at 89 Convent Ave (133rd St).

Course Schedule

Classes in SEMC or NYSBC seminar rooms (Winter course: Mondays 3:30-5pm and Wednesdays 3:30-5pm | Short course weeks: 9:30am-11am)

EM fundamentals section (month of January)

Jan 6: Lecture - Introduction & SEMC tour (Ed Eng - NYSBC)

Jan 8: Practical - Sample Preparation & Support films (Micah Rapp/Ed Eng - COLU/NYSBC)

Jan 13: Lecture – Basic anatomy of the electron microscope (Micah Rapp – COLU)

Jan 15: Practical – TEM use

Jan 20: MLK Jr holiday - No class

Course Administrator:

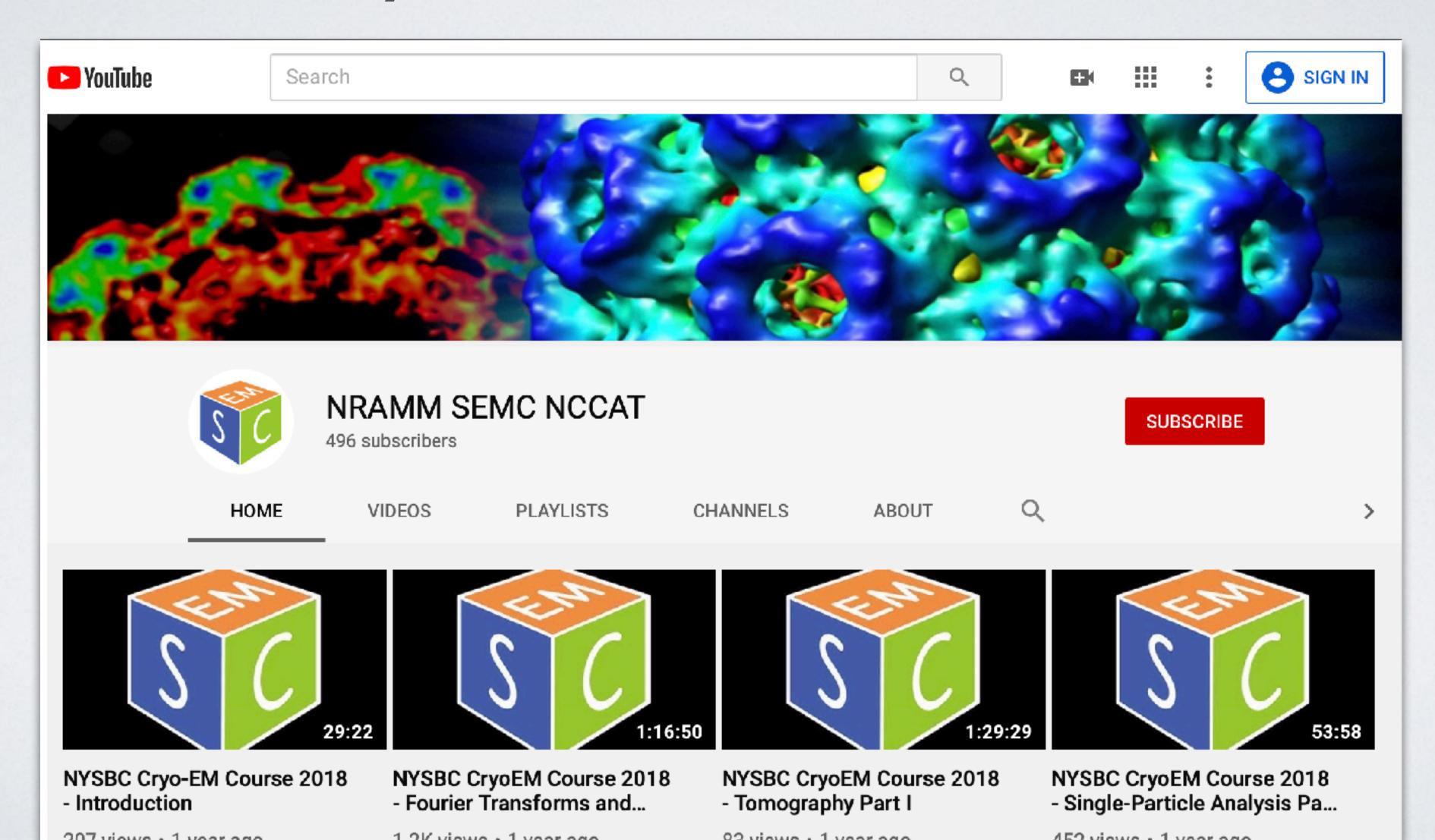
Ed Eng (eeng@nysbc.org)

Teaching Assistant:

Micah Rapp (mar2294@columbia.edu)

Course logistics: live stream

youtube.com/nrammsemc



Course logistics

Section la : EM fundamentals section

b:2D EM section

c: SEMC Appion workshops

d: SEMC journal clubs

Section 2 : Single-particle short-course

Section 3: Tomography short-course



Course logistics

Mondays

3:30-5pm - A-11 seminar room / SEMC conference room

Lecture schedule

Jan 6: Introduction & SEMC tour

Jan 13: Basic anatomy of the electron microscope

Jan 20: MLK Jr holiday — No class

Jan 27: Fourier transforms and Image Formation

Feb 3 : MicroED (Bill Rice – NYU)

Feb 10: Helical reconstruction (Hernando Sosa – Einstein)

Feb 17 : President's day holiday — No class

Feb 24 : Q&A – open forum & primer to SPA

Wednesdays

Starts at 3:30 - SEMC conference room

Recitation schedule

Jan 8 : Sample Preparation &

Support films

Jan 15:TEM use

Jan 22 : Journal club

Jan 29: Image pre-processing

Feb 5 : Journal club

Feb 12 : Journal club

Feb 19: Journal club

Course logistics: class for credit

Component

Percentage

Recitation

50%

- JC/HW/questions

Practical Worksheet

10% x 4

Attendance

10%

Wednesdays

Starts at 3:30 - SEMC conference room

Recitation schedule

Jan 8 : Sample Preparation & Support films

Jan 15:TEM use

Jan 22 : Journal club

Jan 29: Image pre-processing

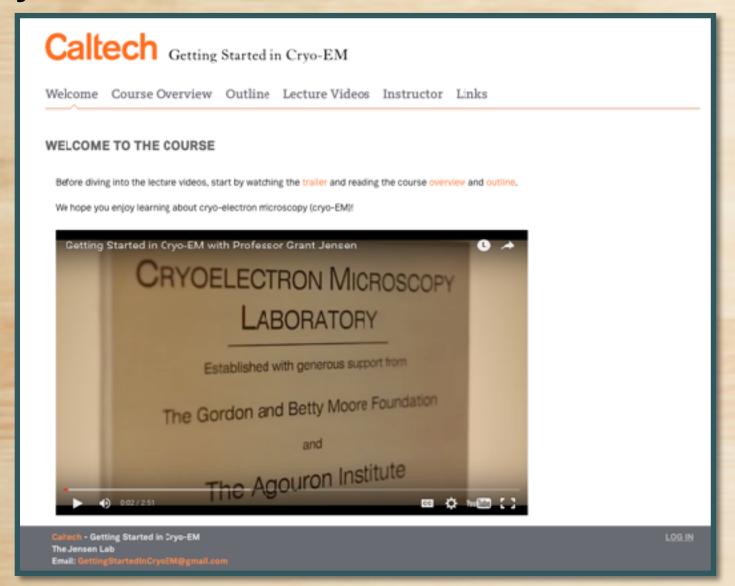
Feb 5 : Journal club

Feb 12: Journal club

Feb 19: Journal club

Course logistics: recitations

cryo-em-course.caltech.edu/videos



Part 4: Fundamental Challenges in Biological TEM & Sample Prep

Unit 2: Sample Preparation youtube.com/playlist?
list=PL8_xPU5epJdfd5fM2CjQlt
R-iRIIEIJk8

cryoem101.org



Chapter I: Sample Purification

Chapter 2: Cryo-EM
Grid Preparation

Wednesdays

Starts at 3:30 - SEMC conference room

Recitation schedule

Jan 8 : Sample Preparation & Support films

Jan 15:TEM use

Jan 22 : Journal club

Jan 29 : Image pre-processing

Feb 5 : Journal club

Feb 12: Journal club

Feb 19 : Journal club

Course logistics: recitations

SEMC conference room



Wednesdays

Starts at 3:30 - SEMC conference room

Recitation schedule

Jan 8 : Sample Preparation & Support films

Jan 15:TEM use

Jan 22 : Journal club

Jan 29: Image pre-processing

Feb 5 : Journal club

Feb 12: Journal club

Feb 19: Journal club

Course logistics

Section la : EM fundamentals section

b: 2D EM section

c: SEMC Appion workshops - Jan 30

https://www.surveymonkey.com/r/BHVHYK3

Section 2 : Single-particle short-course - March 2

d: Additional journal clubs

Section 3: Tomography short-course - April 13

le: Course wrap up - TBD

Recitation schedule

Jan 8 : Sample Preparation & Support films

Jan 15:TEM use

Jan 22 : Journal club

Jan 29: Image pre-processing

Feb 5 : Journal club

Feb 12: Journal club

Feb 19: Journal club



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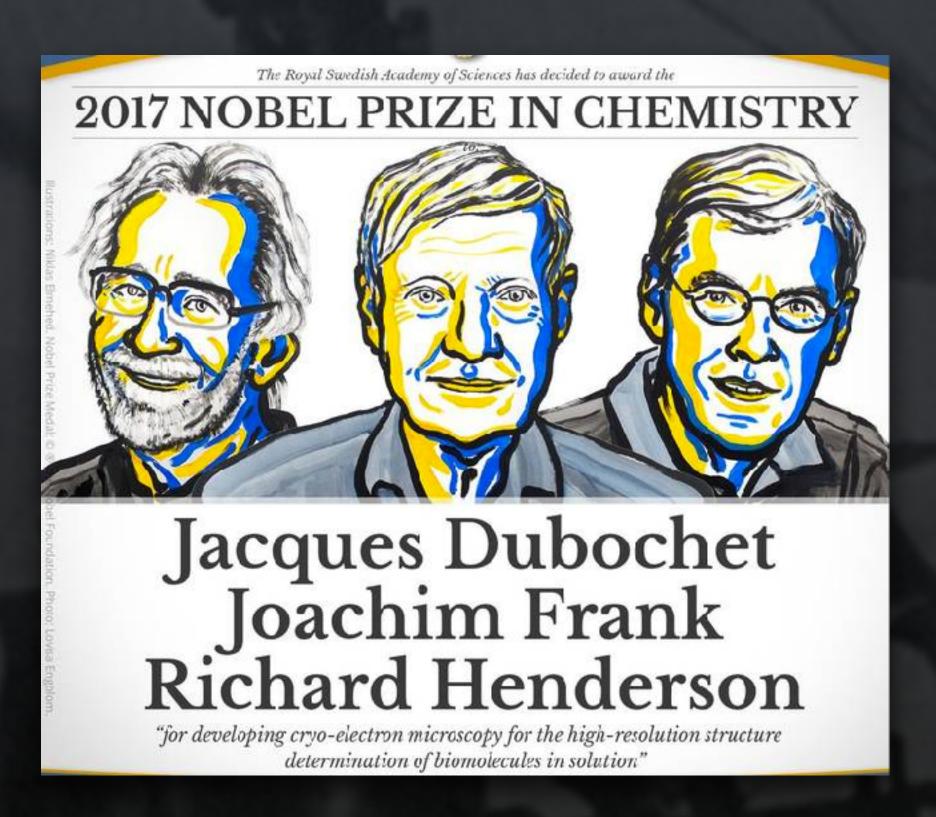


CRYOEM: TECHNOLOGY ON THE RISE

Single-particle cryo-electron microscopy (cryoEM) is the Method of the Year 2015

nature methods Review on CRISPR-Cas9 specificity Reconstruction of dense neural populations Photoswitchable probe for photoacoustic imaging A refined force field for DNA simulations ■ METHOD OF THE YEAR 2015

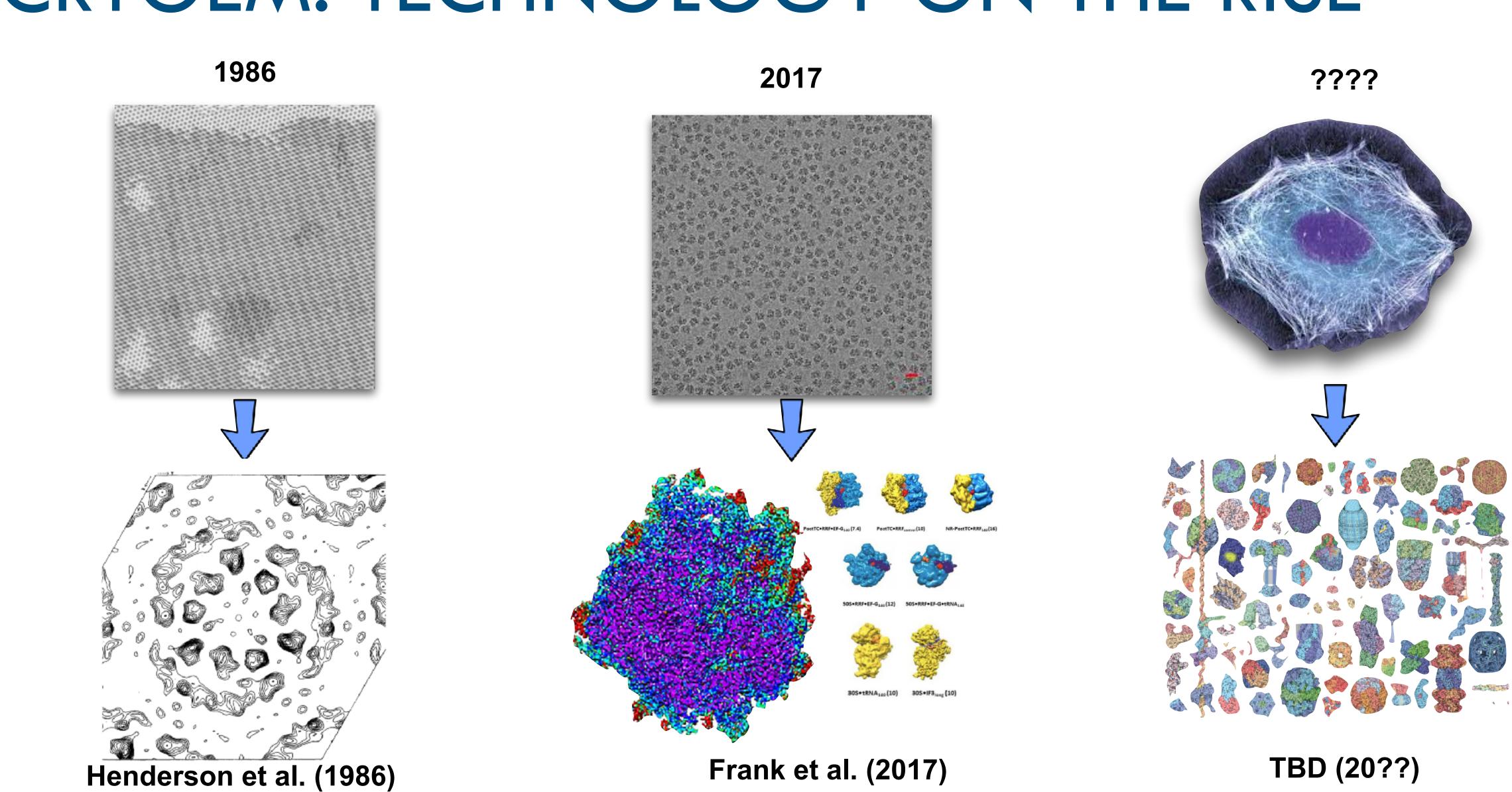
Chemistry Nobel prize 2017



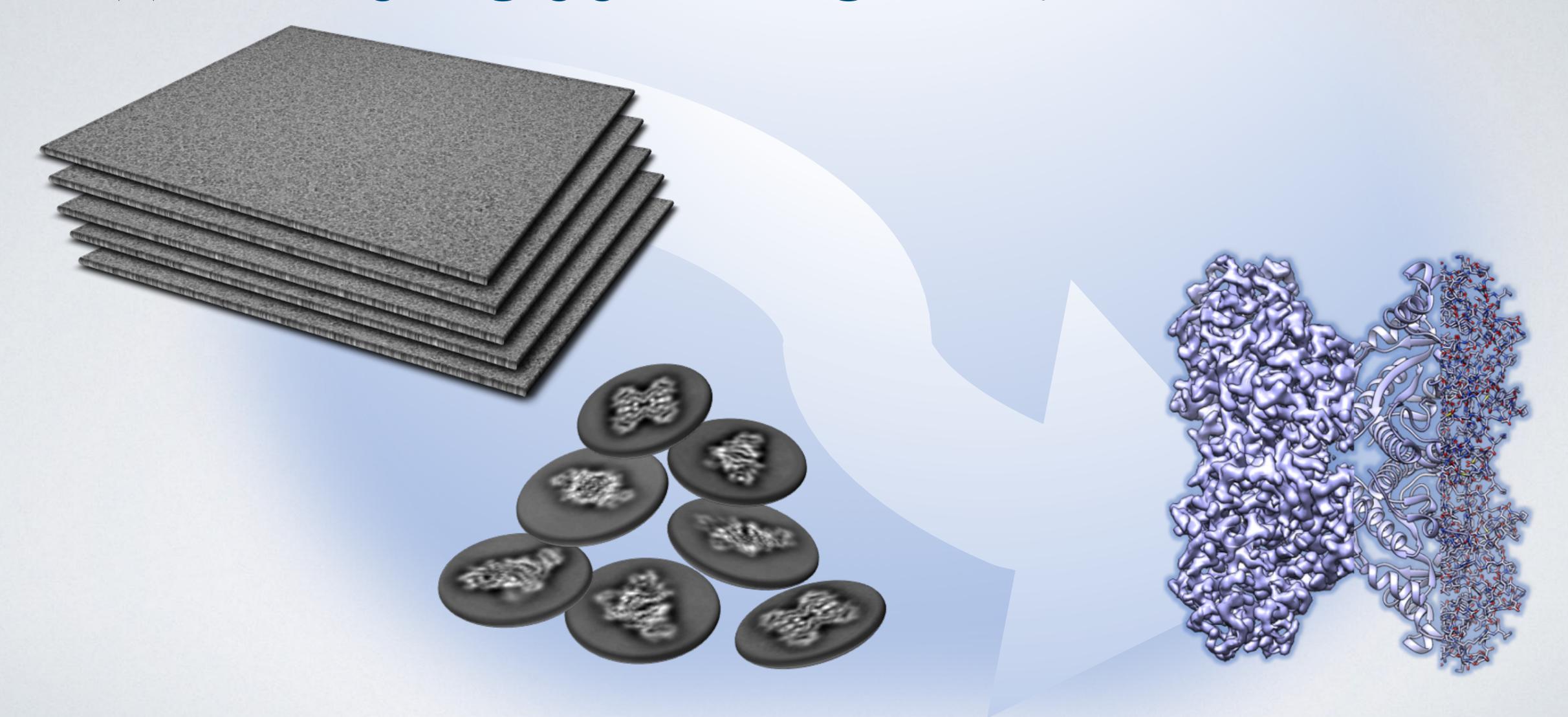
microED
Science breakthrough of the year runner-up 2018



CRYOEM: TECHNOLOGY ON THE RISE



WHAT IS POSSIBLE TODAY?



WHAT BROUGHT ABOUT THE RESOLUTION REVOLUTION? (~2012-2014)

Microscopes



17,000 images motion correction 2,675,742 particles

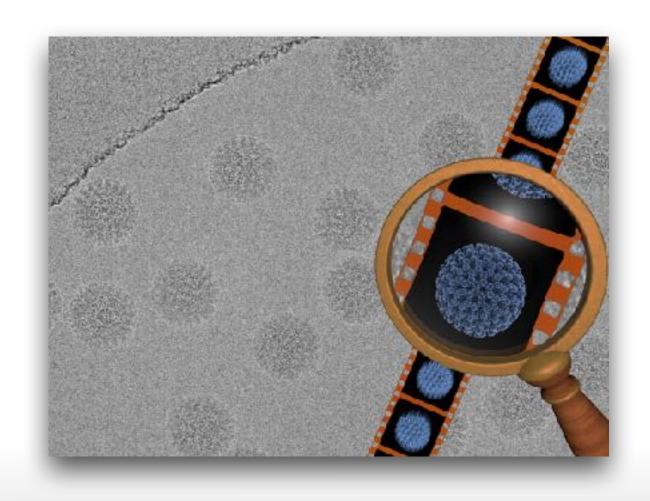
efinement tocused on complex minus GasAH

Subtraction of micelle and GasAl-

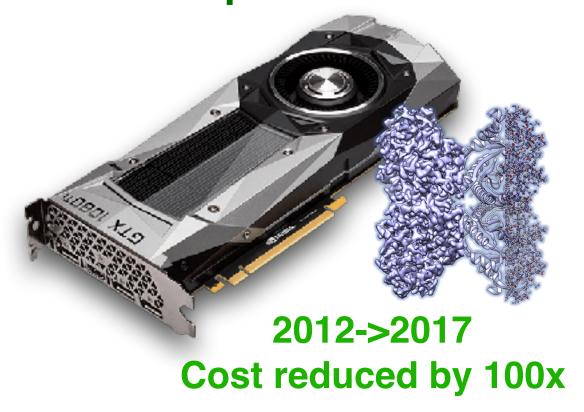
5% used in map! (3.9 Å in core region)

139,299 particles (23%)

Direct Detectors



Computers



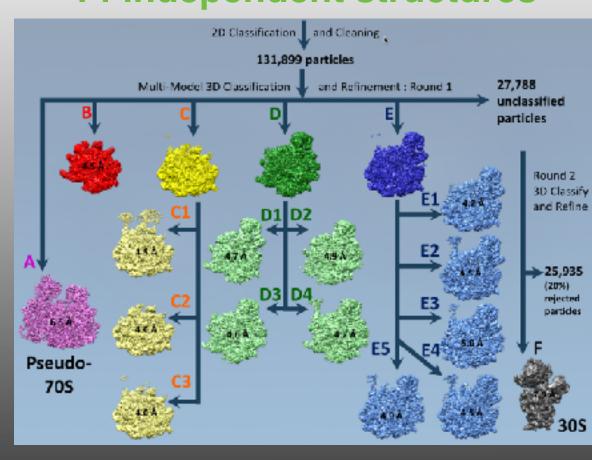
Leginon / SerialEM / EPU, ...

MotionCorr2, Unblur, ...

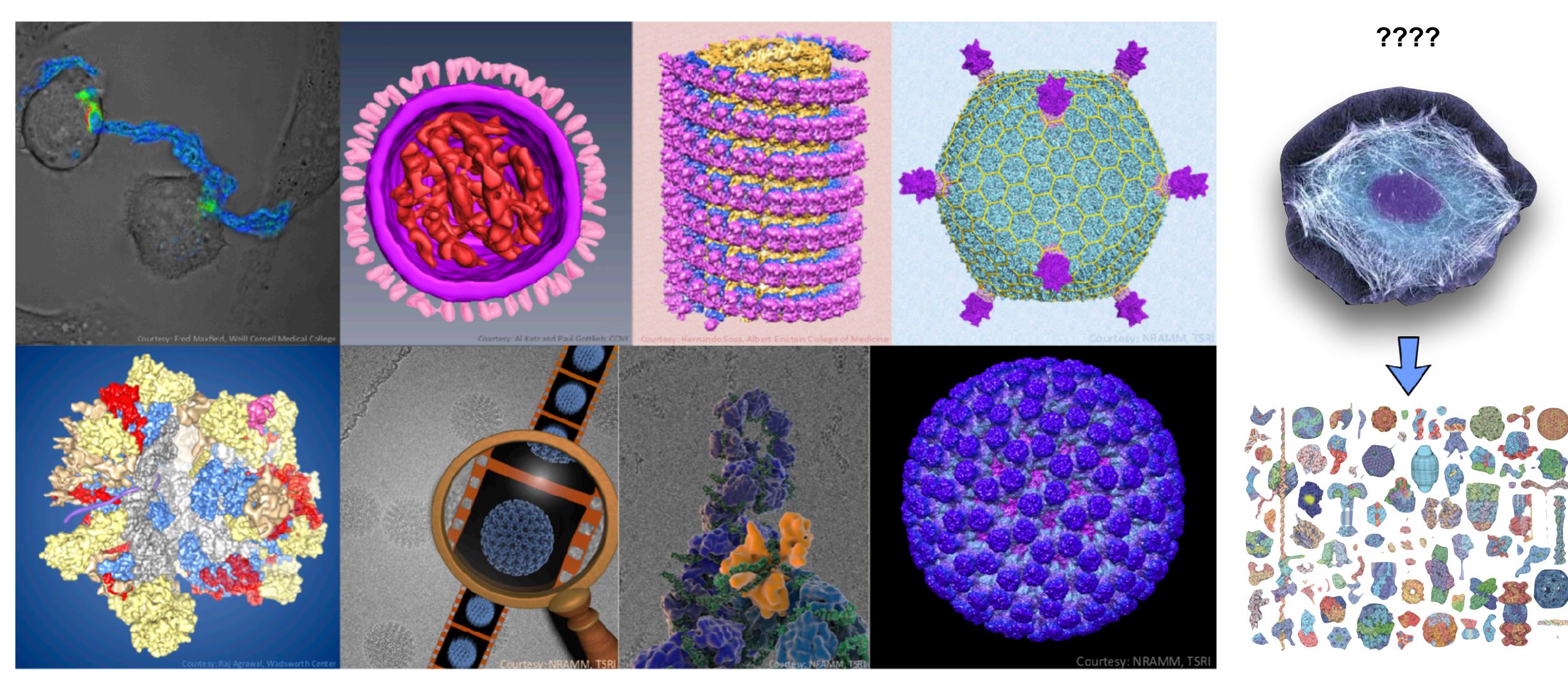
EMAN, Sparx, SPHIRE, XMIPP, ...

RELION, FREALIGN/cisTEM, cryoSPARC

14 independent structures

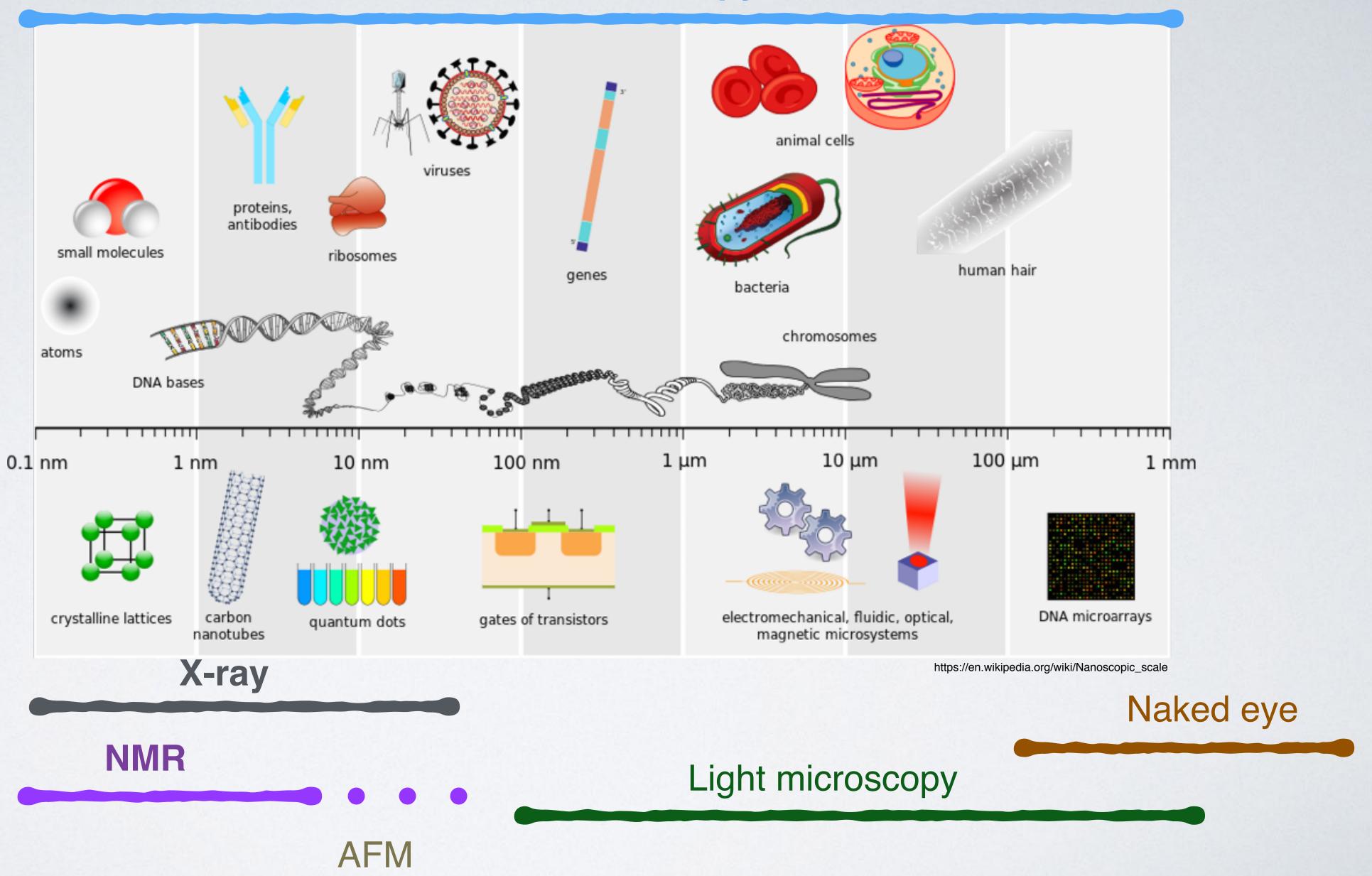


CRYOEM: TECHNOLOGY ON THE RISE



Electron Microscopy

CRYOEM: SCALE WITHIN BIOLOGY



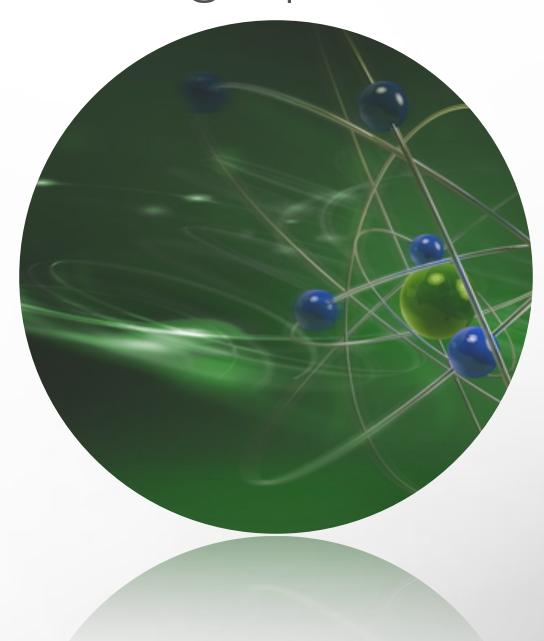
CRYOEM:

MODALITIES TOOLS

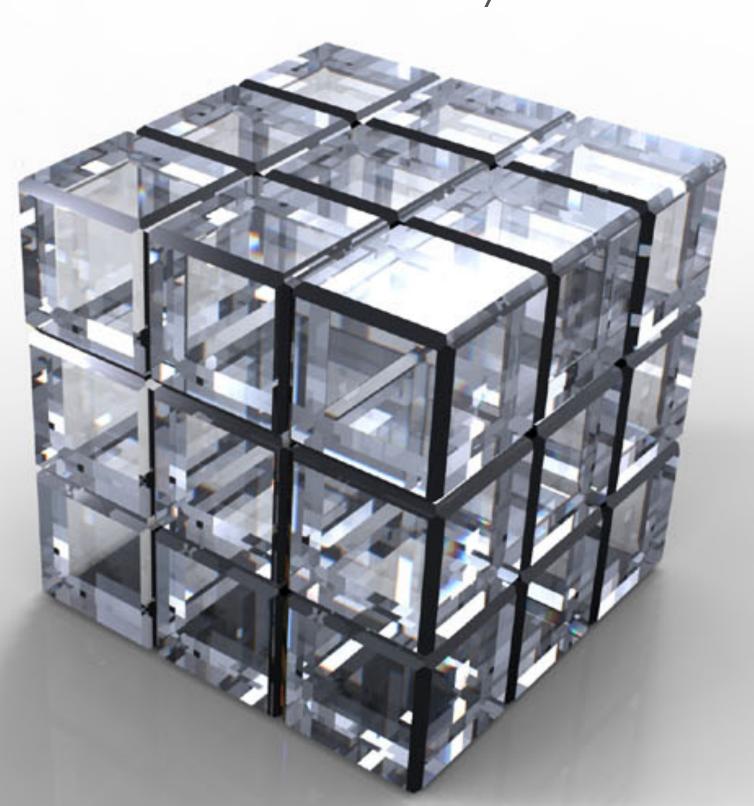
Tomography



Single-particle



2D arrays



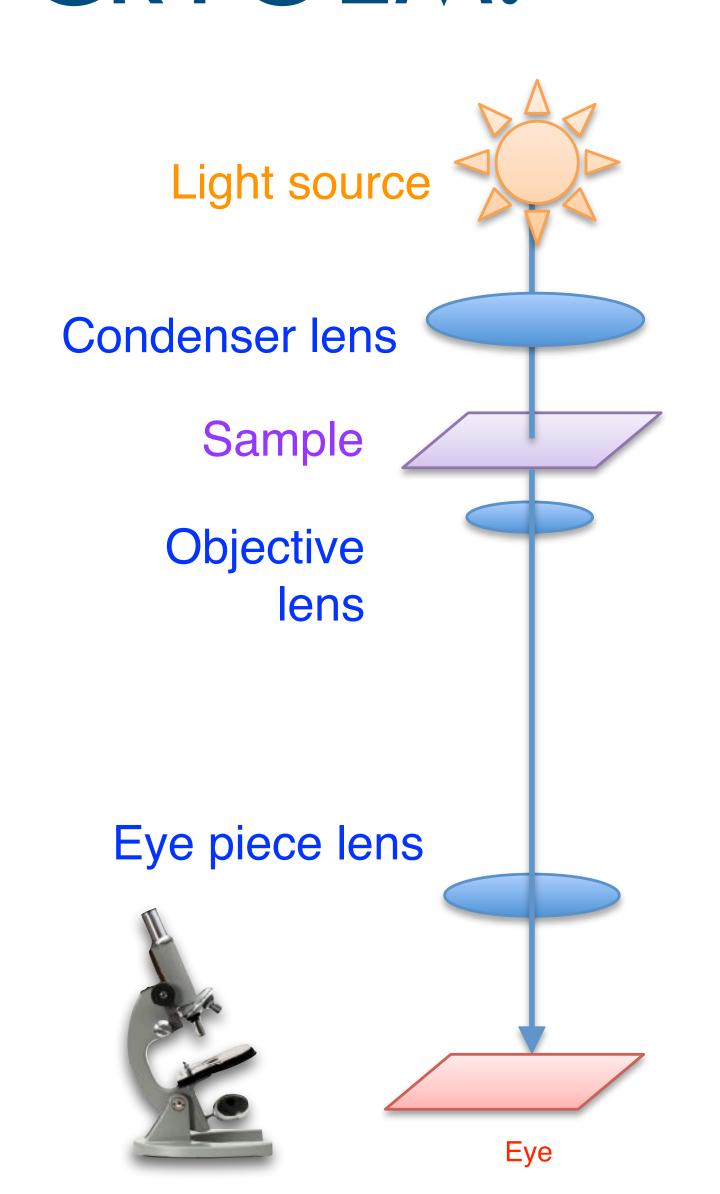


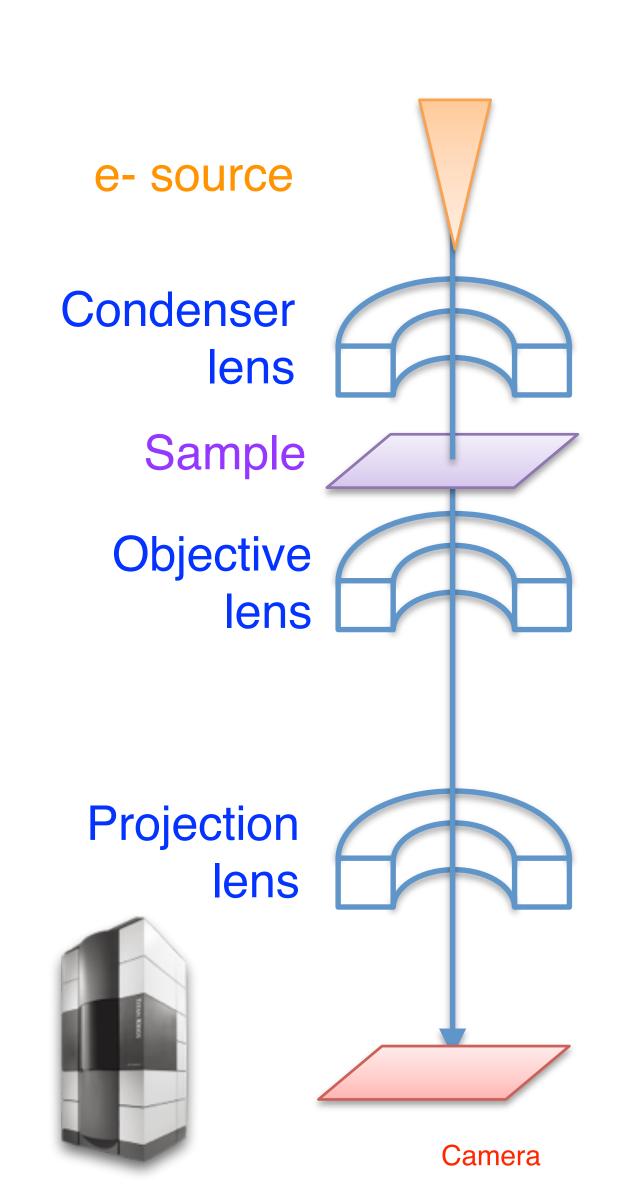


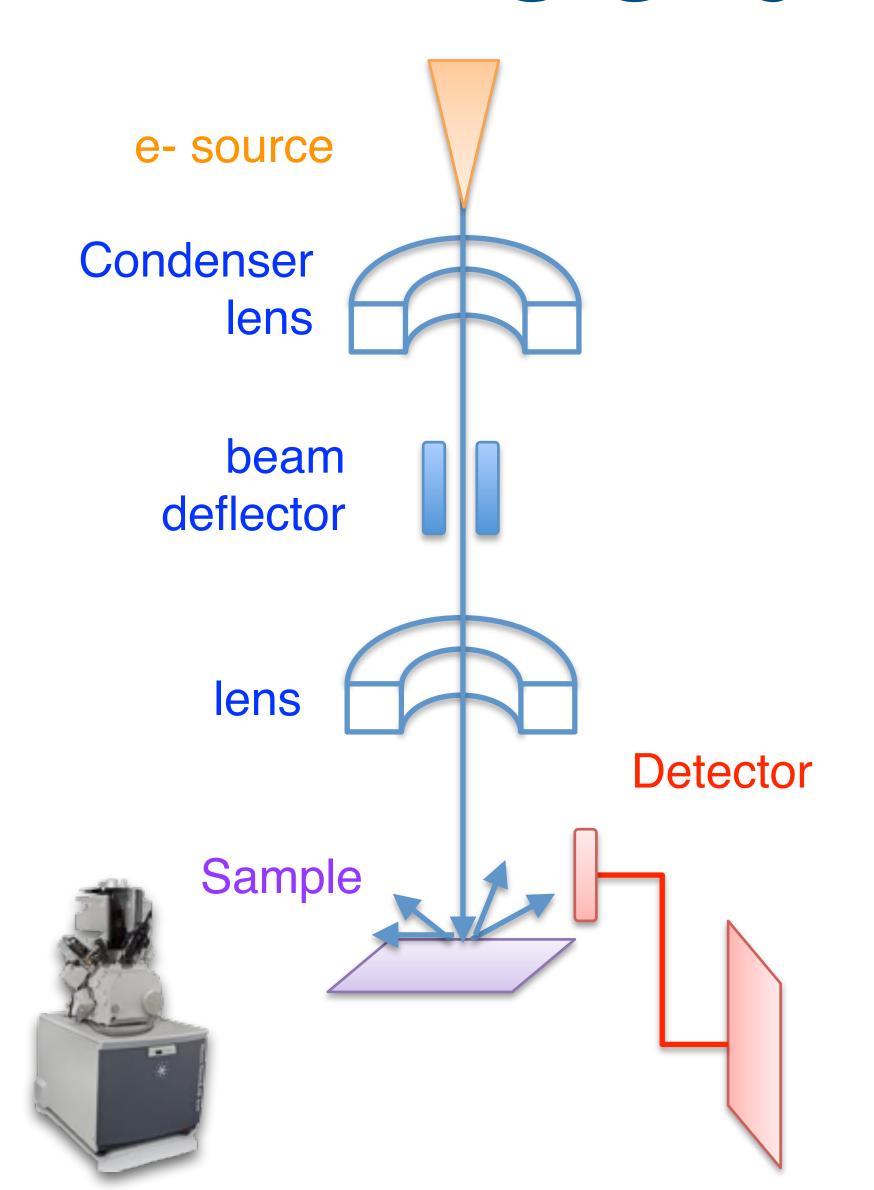


CRYOEM:

TOOLS



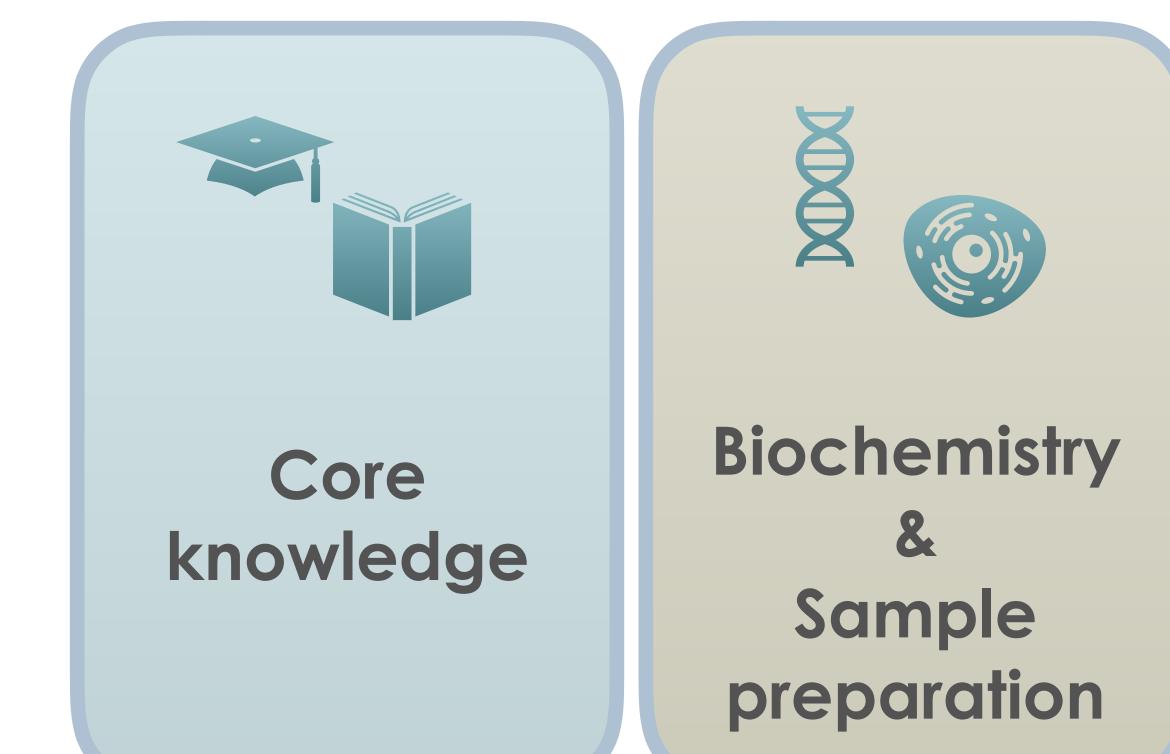


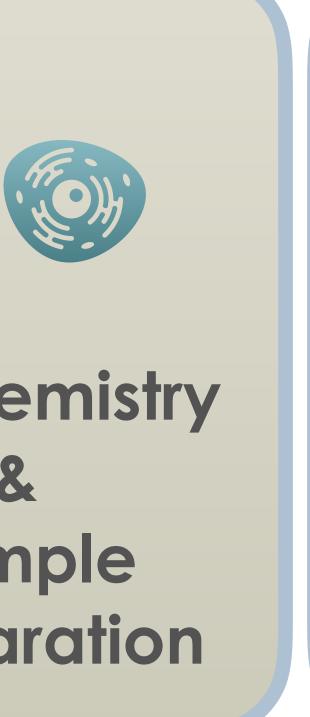


THAT'S GREAT... HOW DO I START?



THAT'S GREAT... HOW DO I START?









FOCUS ON 4 AREAS



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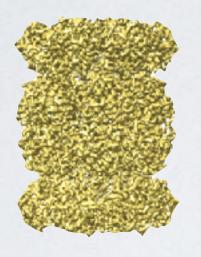


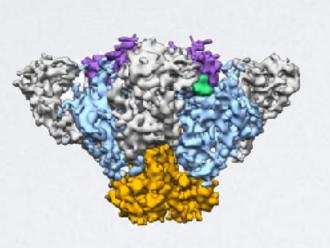
Simons Electron Microscopy Center



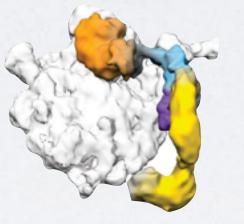
Focus on:

Molecular Structure:





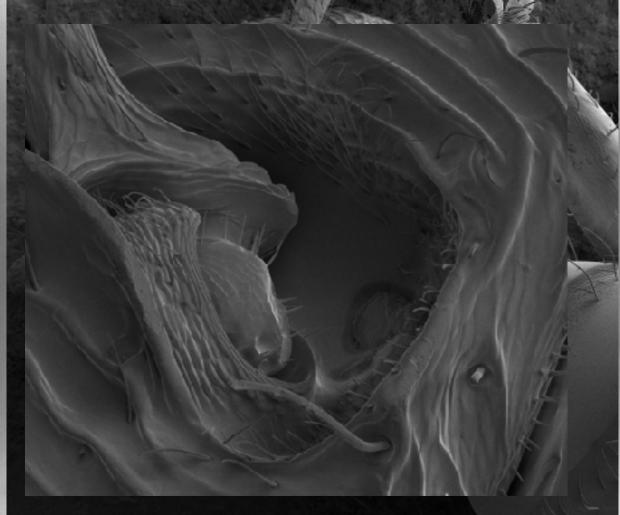














Simons Electron Microscopy Center



https://semc.nysbc.org/workshops-and-courses/

yearly

SEMC EM Course

Theory behind EM **Spring semester**

quarterly

SEMC Appion workshop

Appion data processing

monthly

SEMC New User Orientation

Sample preparation Leginon intro/use of screening

24-hr access test

All hours access test Safety training

weekly

User Project Discussion Meetings

Tue @ 3pm Thurs @3pm /@3:30pm

daily

Advanced Leginon use

Training for independent use of the microscopes

TRANSFORMATIVE HIGH RESOLUTION CRYO-ELECTRON MICROSCOPY PROGRAM



https://commonfund.nih.gov/CryoEM



The program aims to broaden access to high-resolution cryoelectron microscopy (cryoEM) for biomedical researchers, by creating

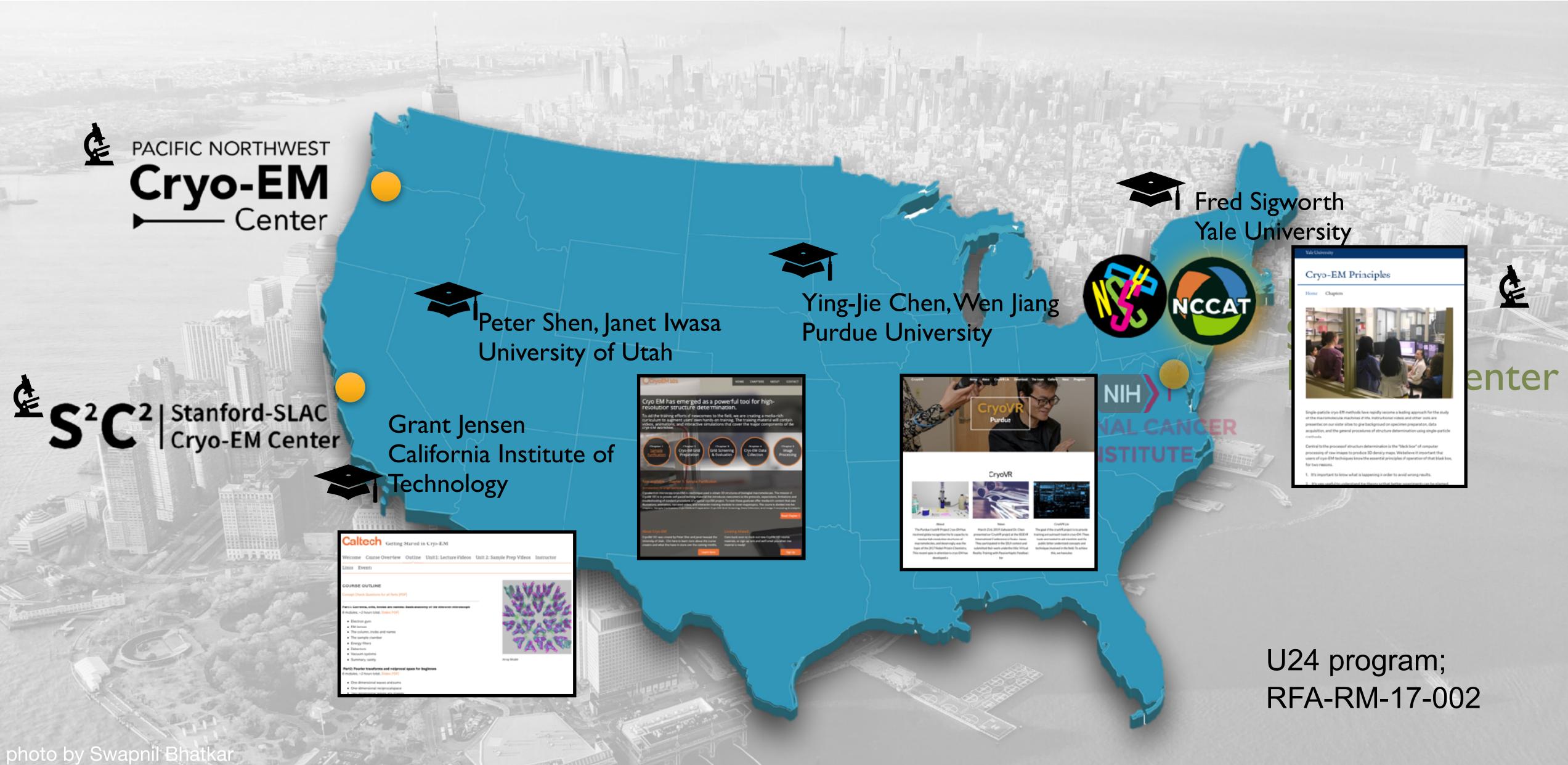
national service centers, and cultivating a skilled workforce, through the development and implementation of cryoEM training material.





TRANSFORMATIVE HIGH RESOLUTION CRYO-ELECTRON MICROSCOPY PROGRAM





National Center for CryoEM Access and Training



Facility

Manager

Program

Krios

Chameleon
specimen
preparation

General User
Proposal (GUP)
Access

Embedded Researcher Program

Cross-Training
Proposal (TP)
Programs

Apply now



nccat.nysbc.org



@nccatinfo

MAKING CRYOEM MORE ACCESSIBLE TO THE COMMUNITY





Directors





Administrators





Scientific computing/programmers

Anchi Cheng



Shaker Krit



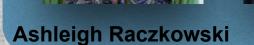
Facility and microscope operations





NYSBC SEMC NCCAT







Laura Yen



Misha Kopylov



Daija Bobe



Carolina Hernandez Robert Gheorghita Mahaira Agaron Kashyap Maruthi







Sargis Dallakyan



Anjelique Sawh Huihui Kuang



Michael Alink

Technology research



Venkat Dandey





Chase Budell

Structural biology research

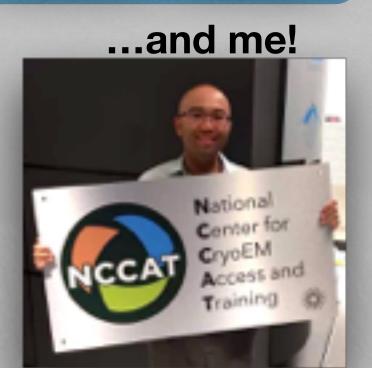


Julia Brasch





Micah Rapp





















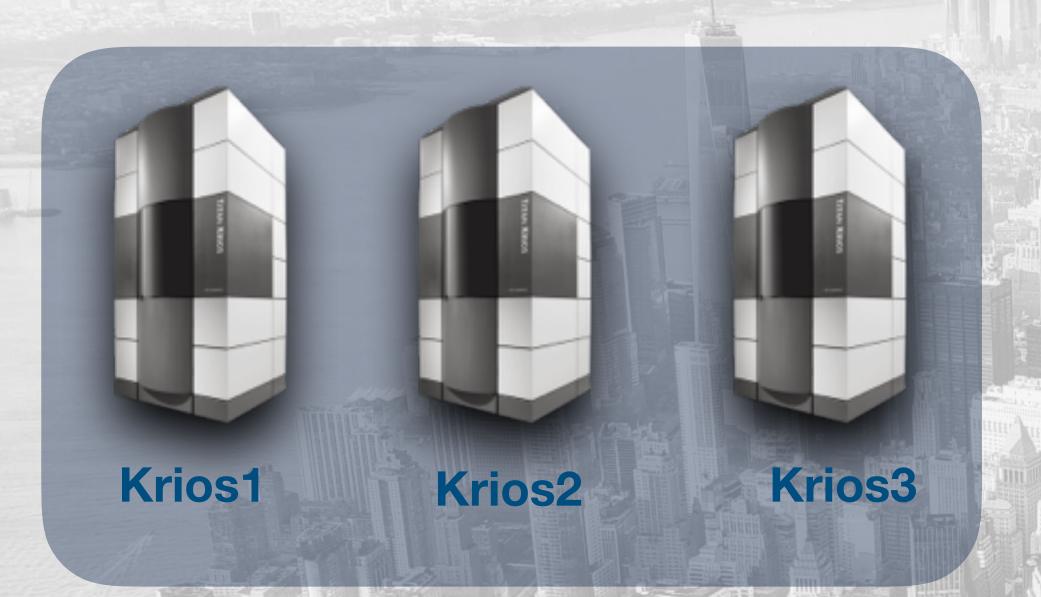








NIH P41 - National Biomedical Technology Research Resources (BTRR)















Chameleon



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