

Appion III Workshop: Fiducial-less Cryo-ET with Appion-Protomo



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Simons Resource for Automated Molecular Microscopy
National Center for In-situ Tomographic Ultramicroscopy
Simons Electron Microscopy Center
New York Structural Biology Center

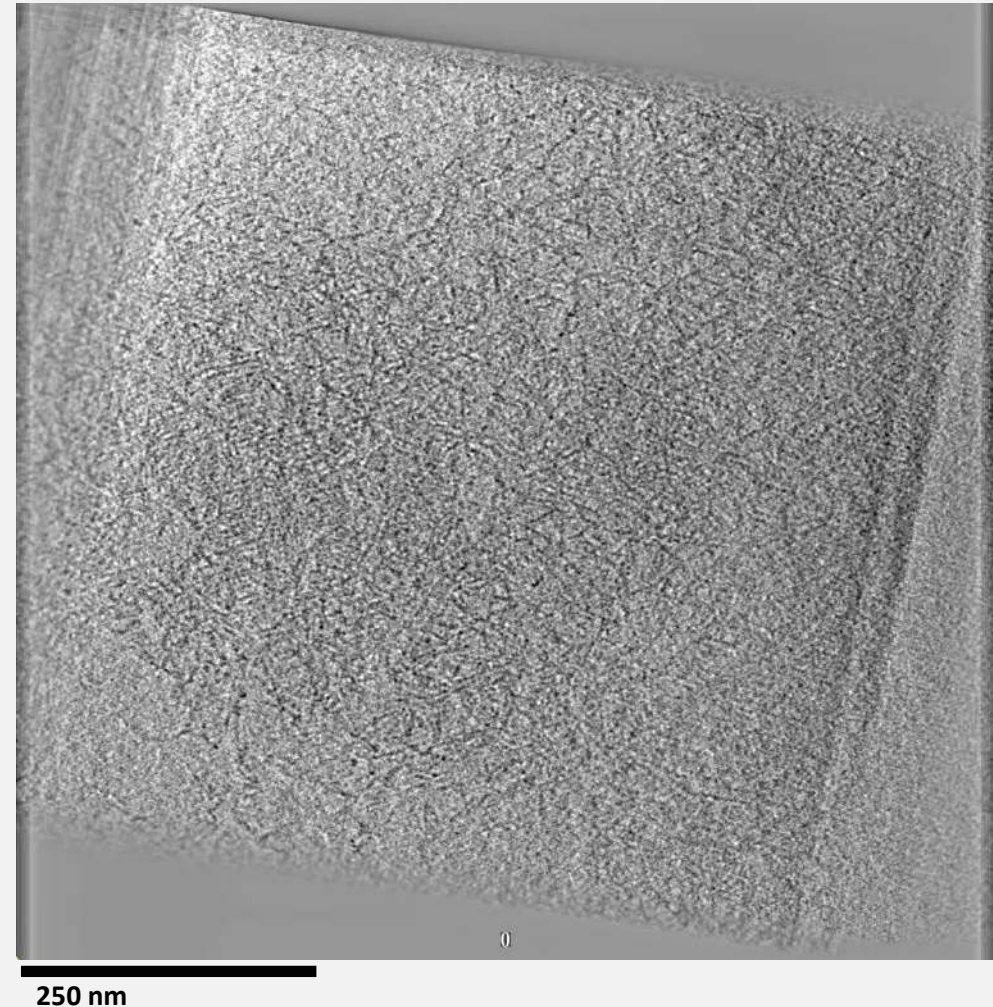
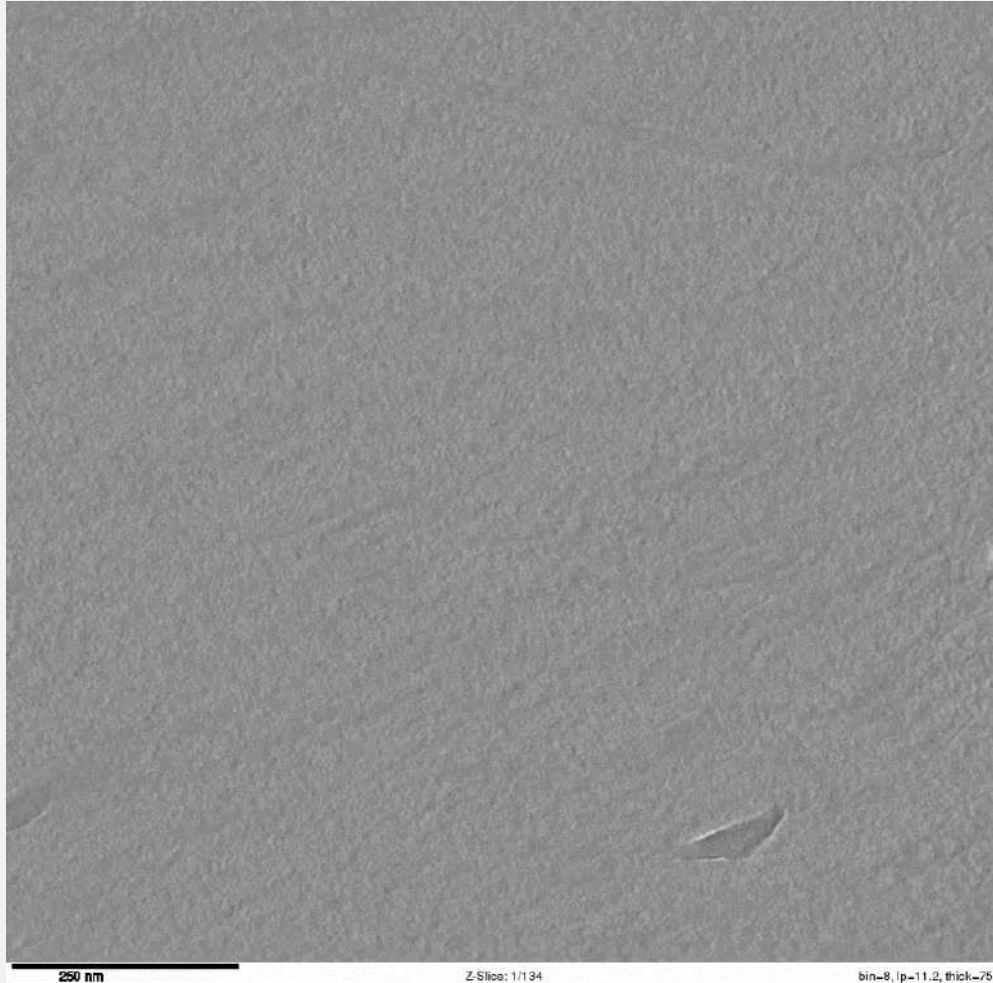


SIMONS ELECTRON
MICROSCOPY CENTER

What is CryoET?

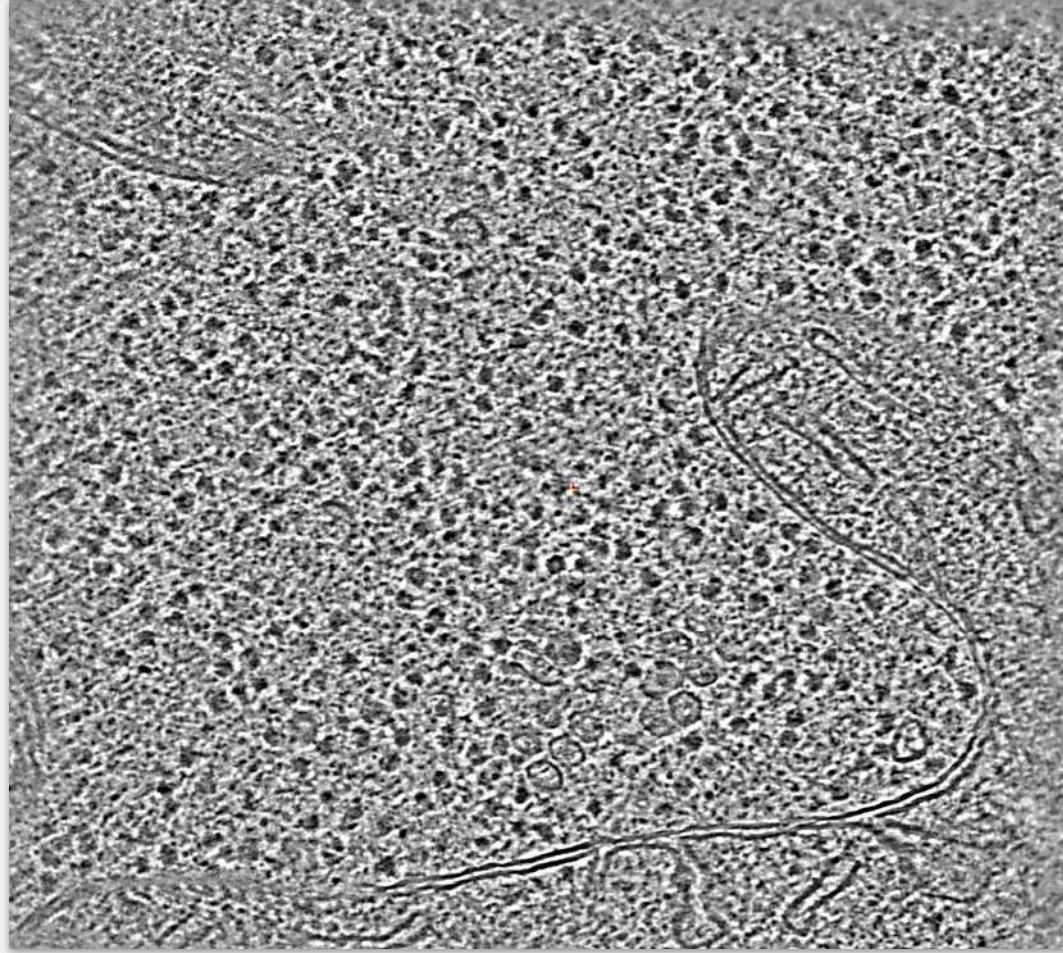
(cryo-electron tomography)

- Cells or complex reconstituted environments



What is CryoET?

(cryo-electron tomography)



250 nm



250 nm

>> CryoET is the **highest resolution method** for **native specimen**

What are we doing today?



Cryo-ET Overview

Why CryoET?

- For morphological **heterogeneity** and **unique** objects

Rough order of events

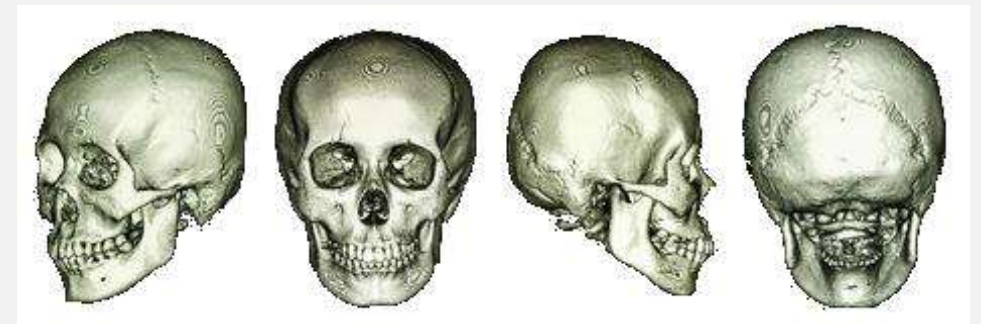
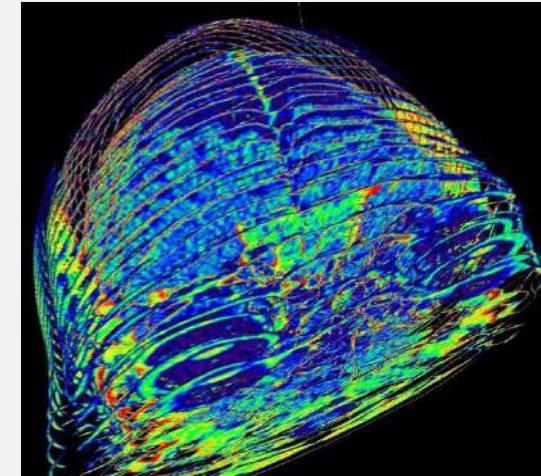
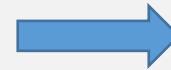
- **Tilt-series alignment** in general
- Segmentation
- Sub-tomogram processing for **repeating objects**



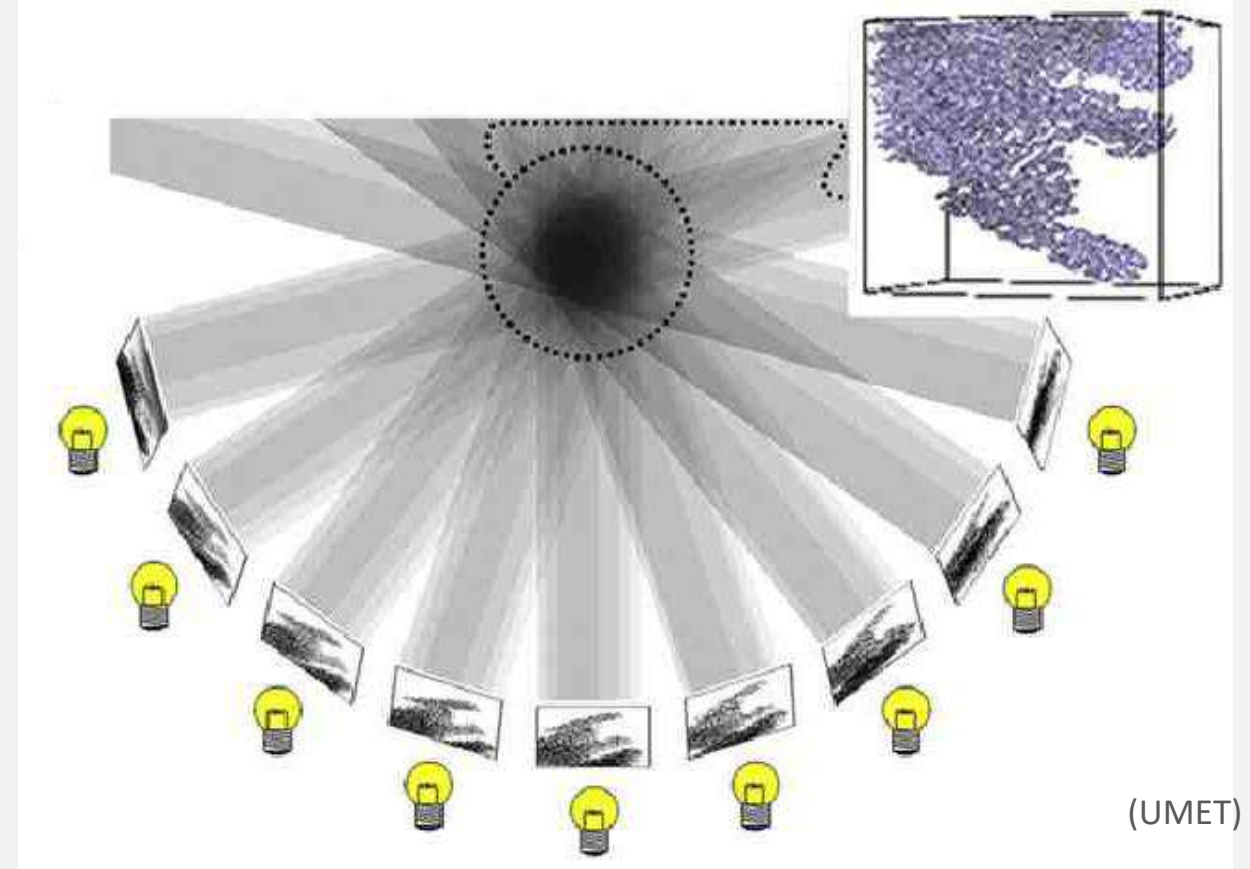
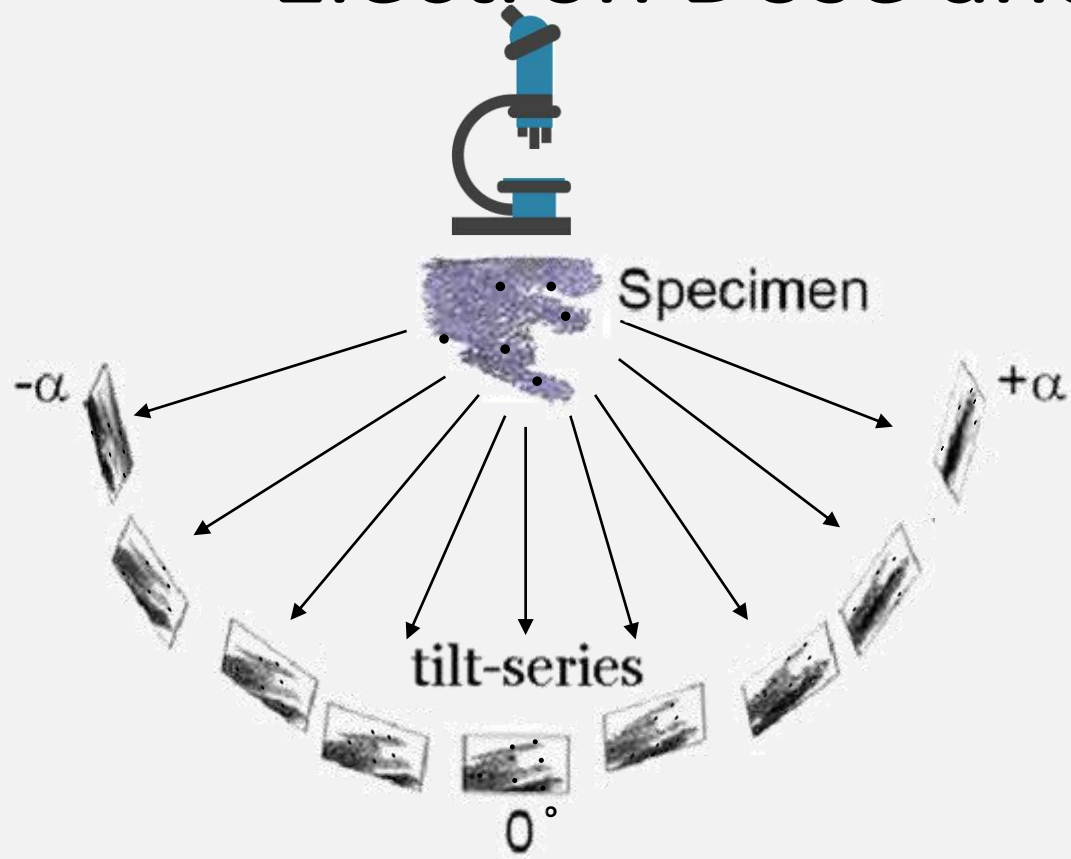
What is tomography
in general?



Tomography Overview – Consider How a CT Scan Works

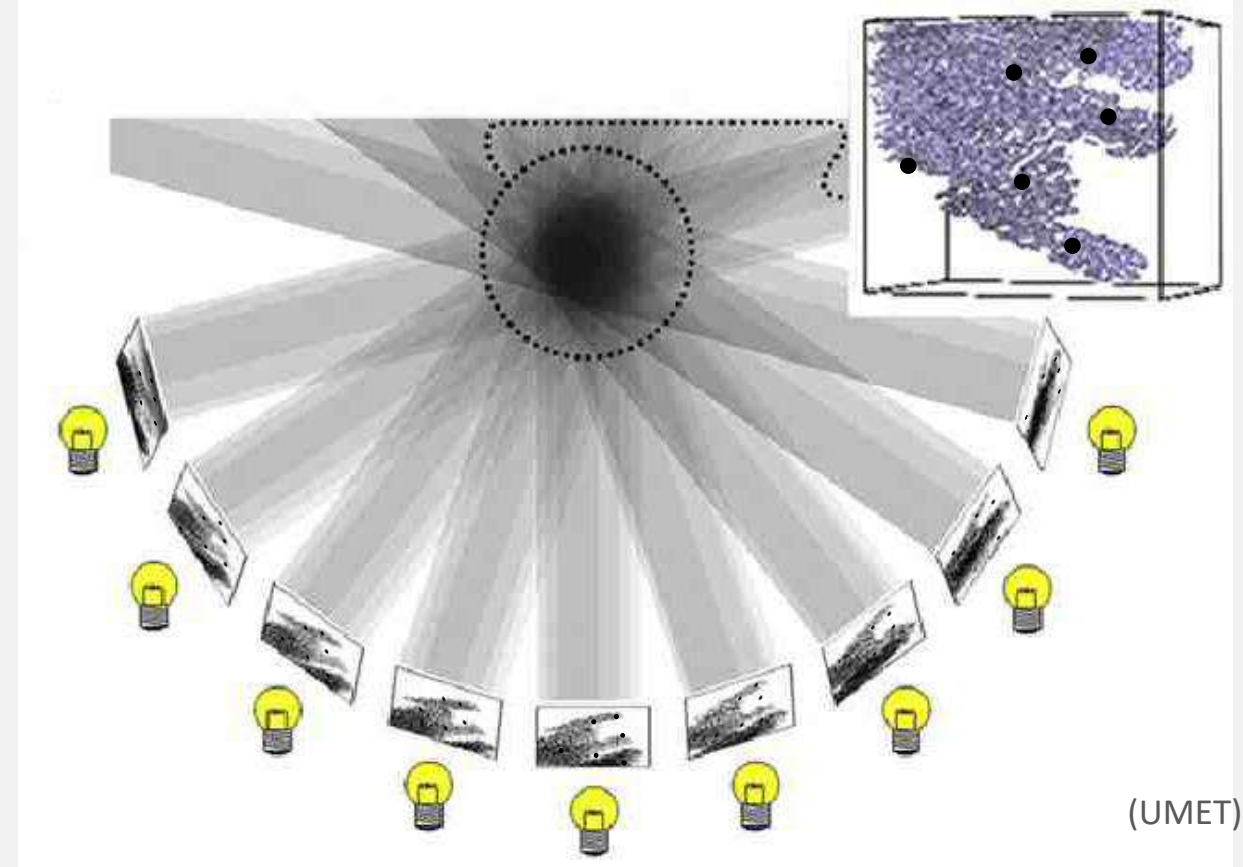
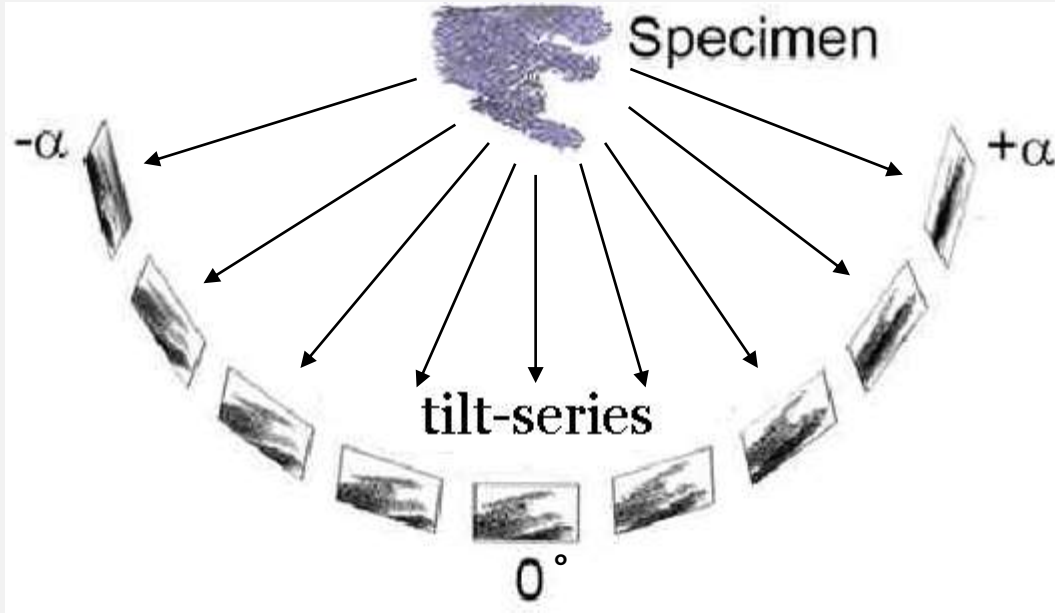
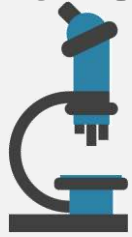


Tomogram Accuracy is Primarily Limited by Electron Dose and Tilt-Series Alignment



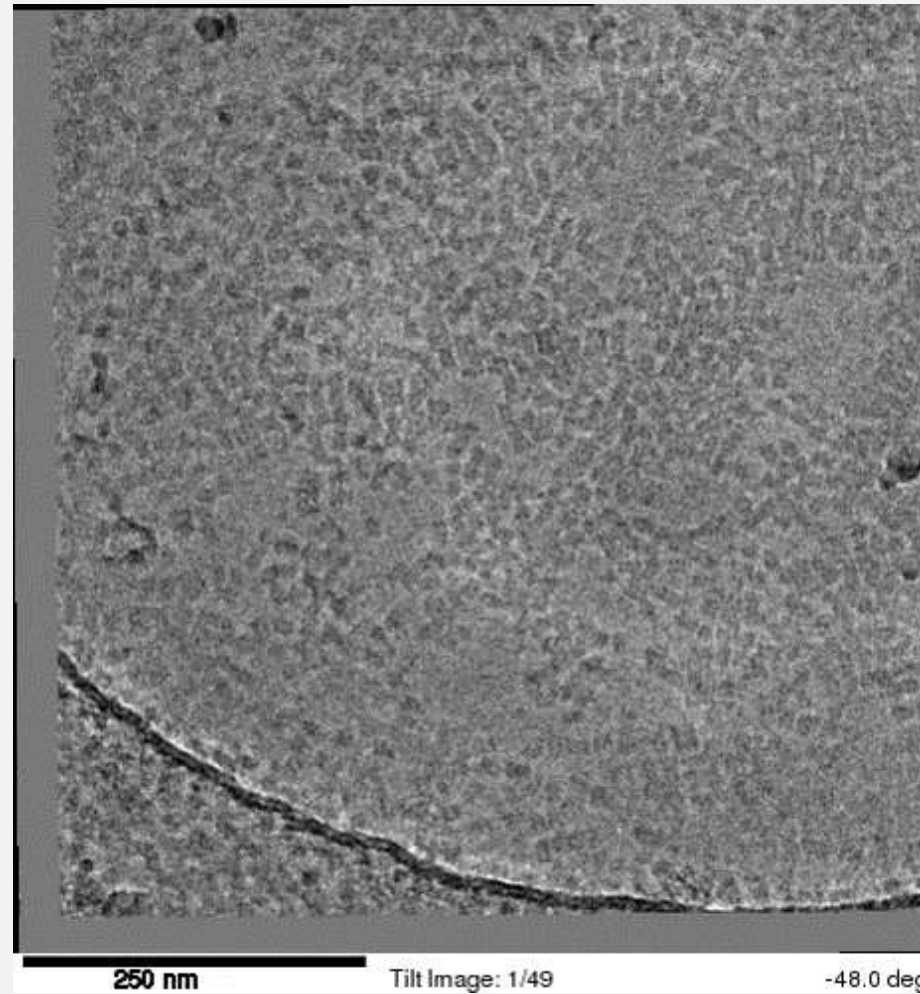
Collect —————> align —————> reconstruct

Tomogram Accuracy is Primarily Limited by Electron Dose and Tilt-Series Alignment

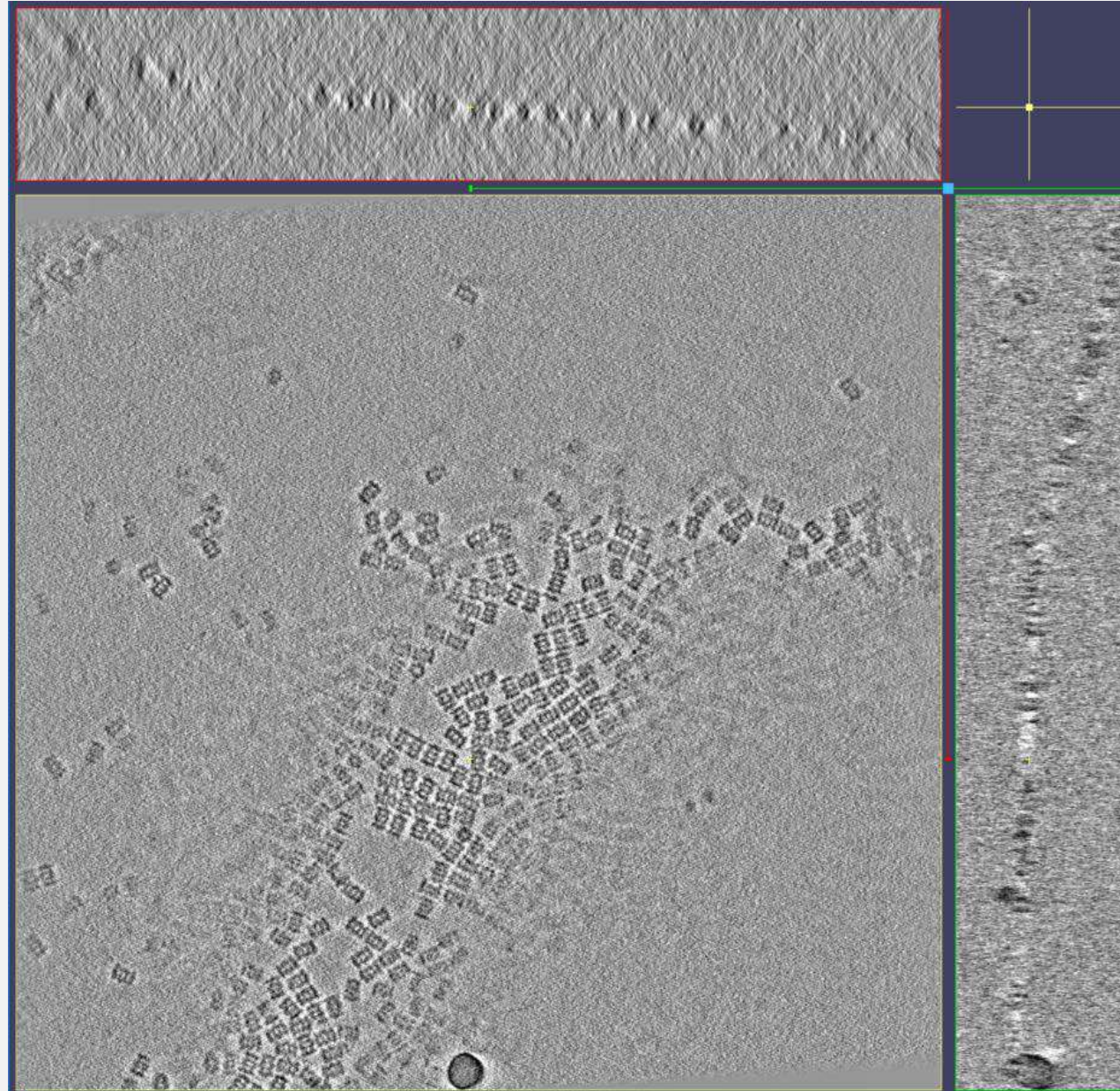


Collect → align → reconstruct

Typical tilt-series (T20S proteasome single particle)

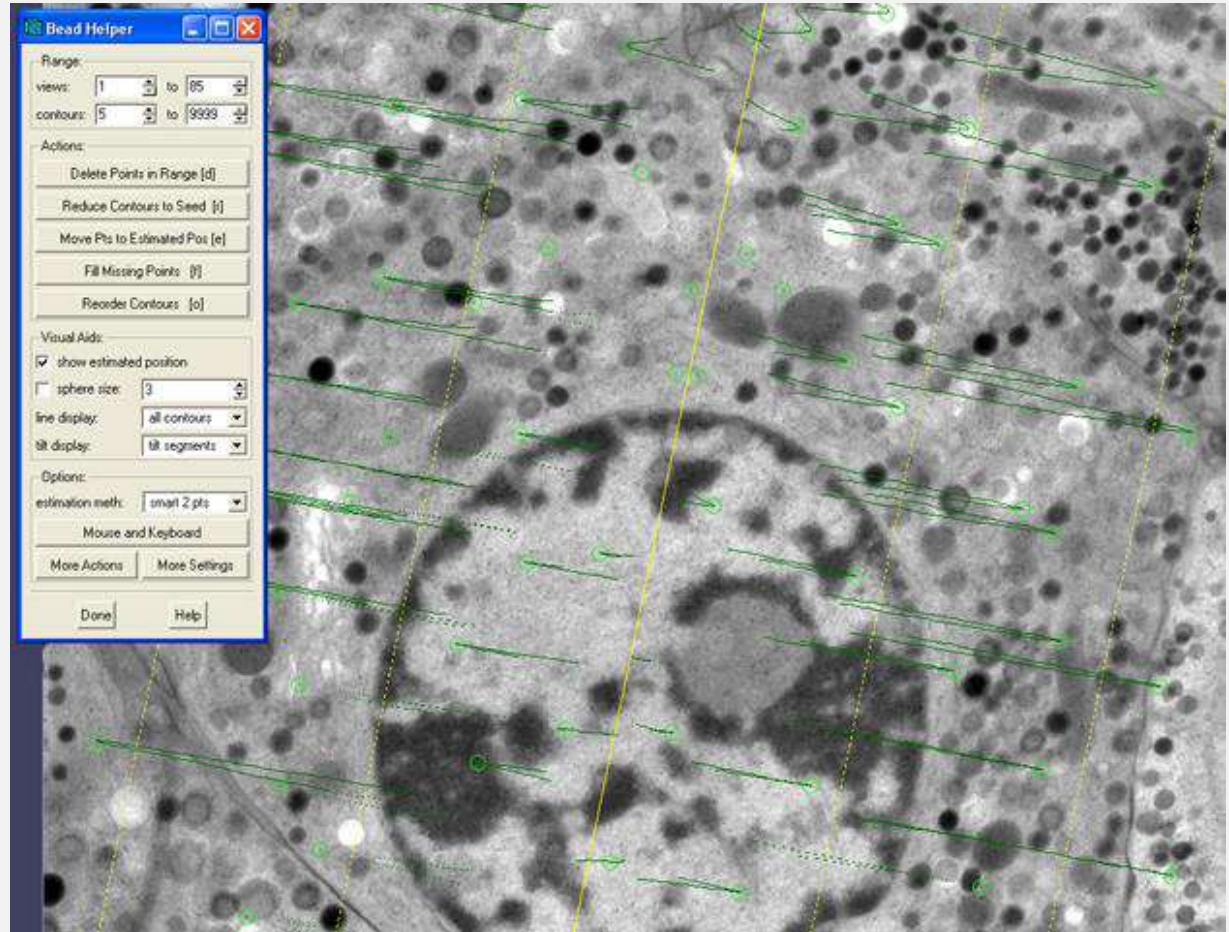


Missing Wedge Effects



Fiducial-based Tilt-Series Alignment

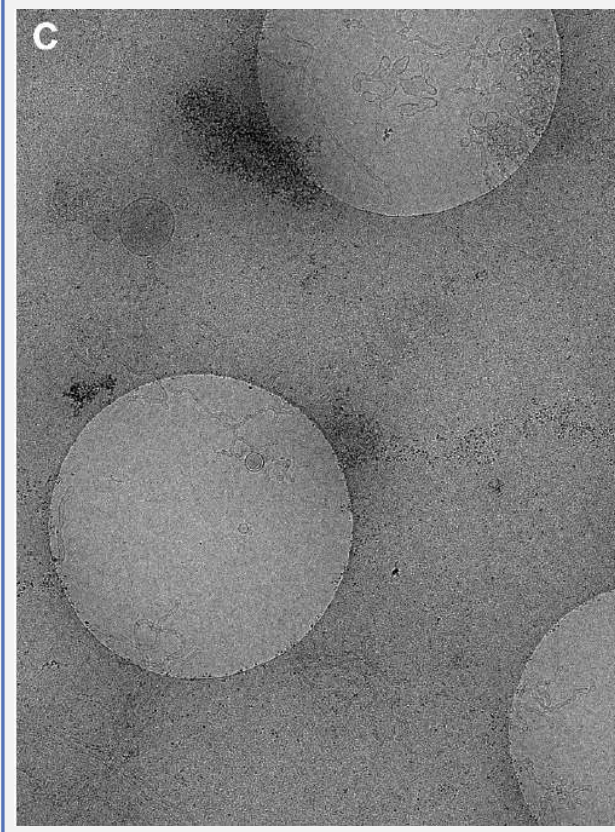
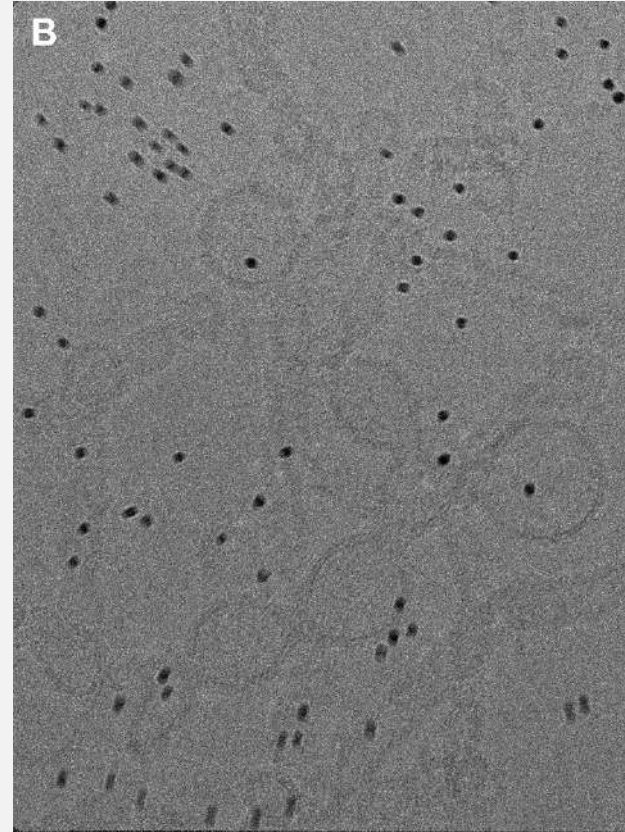
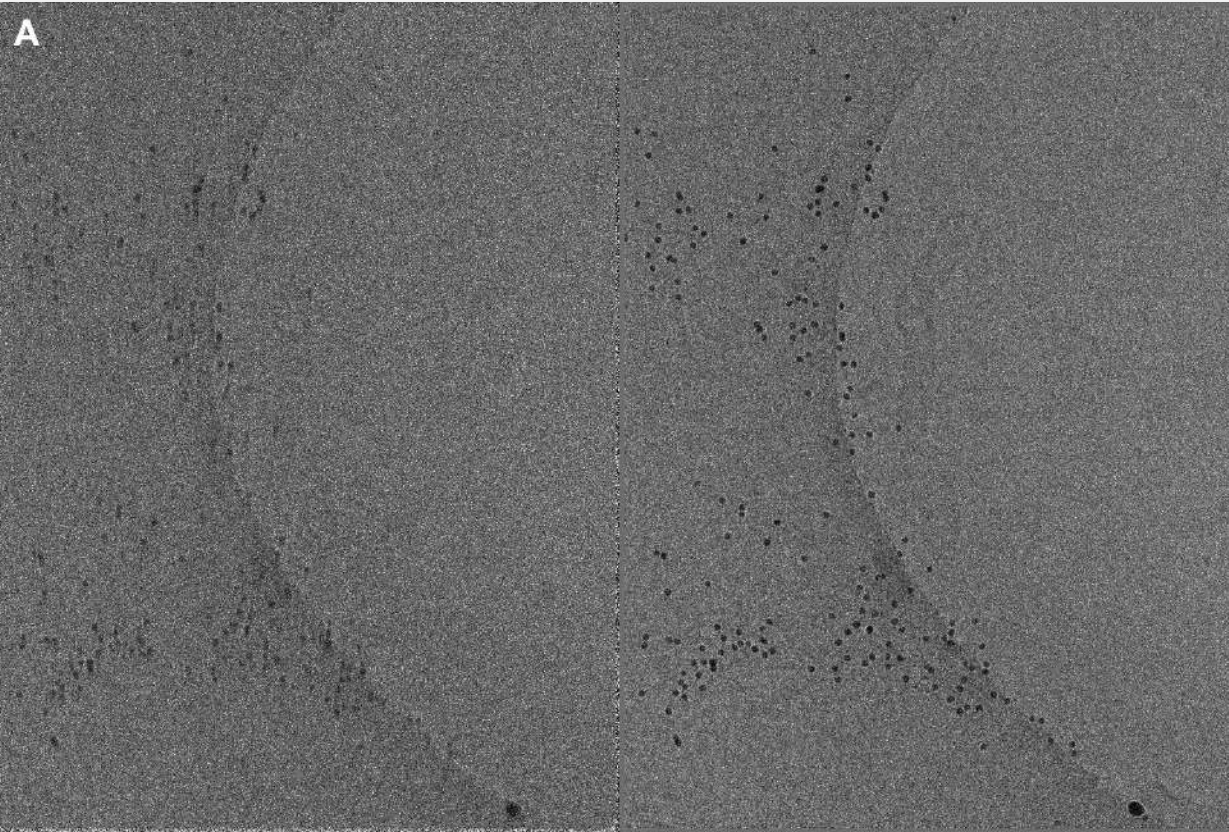
- **Requires a sufficient number of well-behaved gold beads**
- Gold beads should be **uniformly distributed** around the sample location
- Semi-automated processing



Fiducial-based Tilt-Series Alignment Issues

Isotropic and anisotropic fiducial movement relative to the sample (in 3D!)

Aggregation



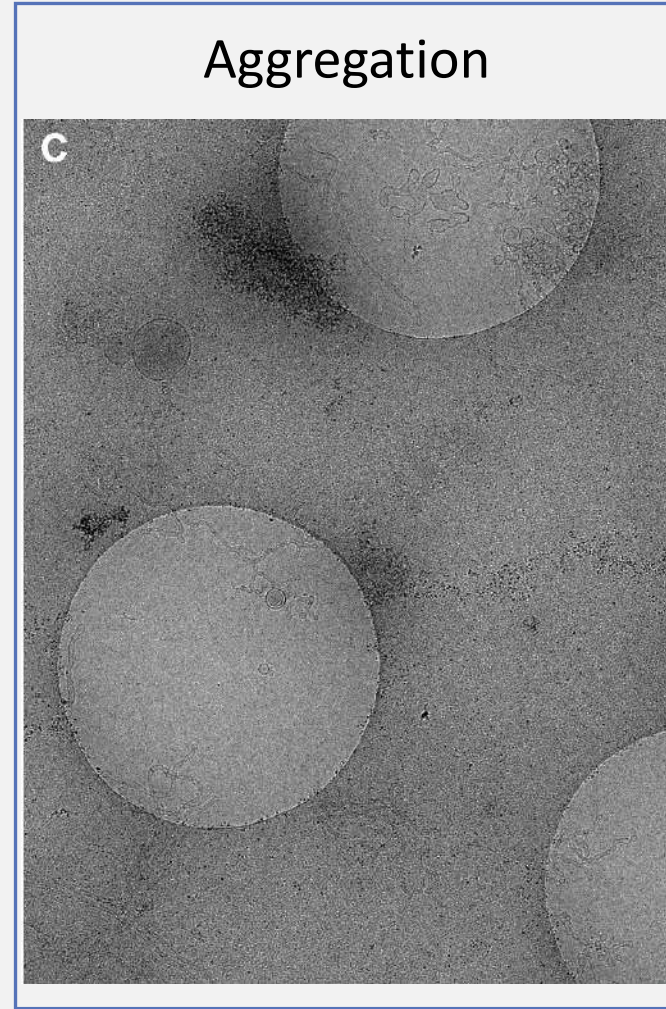
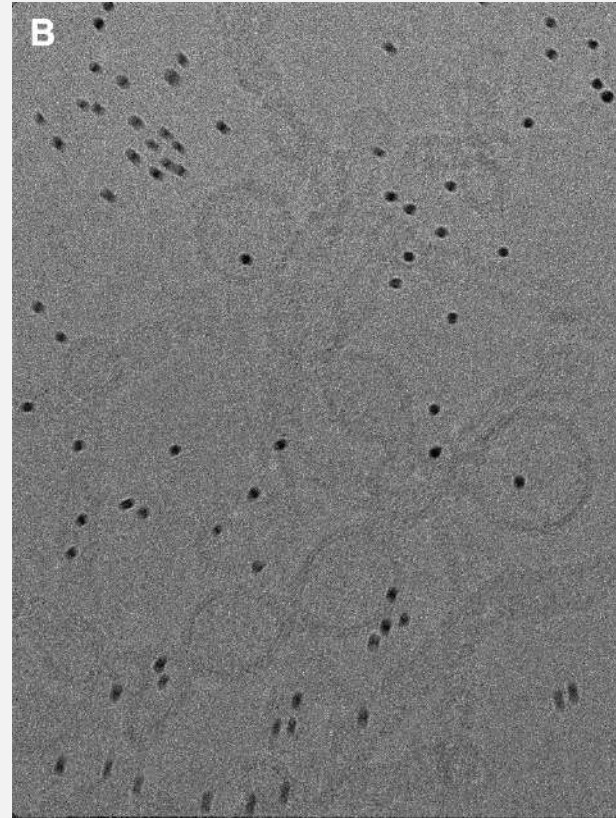
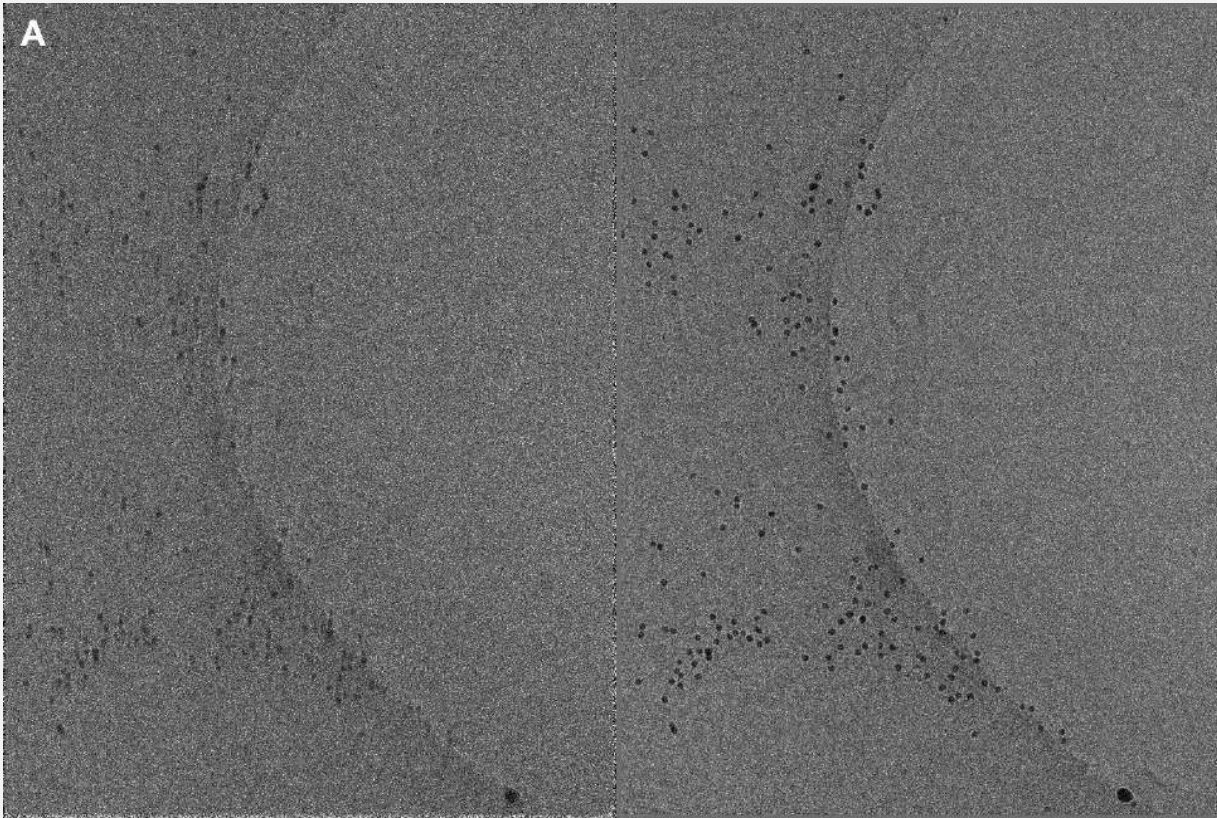
DE-20 @ 18kx; 51°, 2.34 e⁻/Å² after a cumulative dose of 60 e⁻/Å²

DE-20; 57.5 e⁻/Å², 0° exposure



Fiducial-based Tilt-Series Alignment Issues

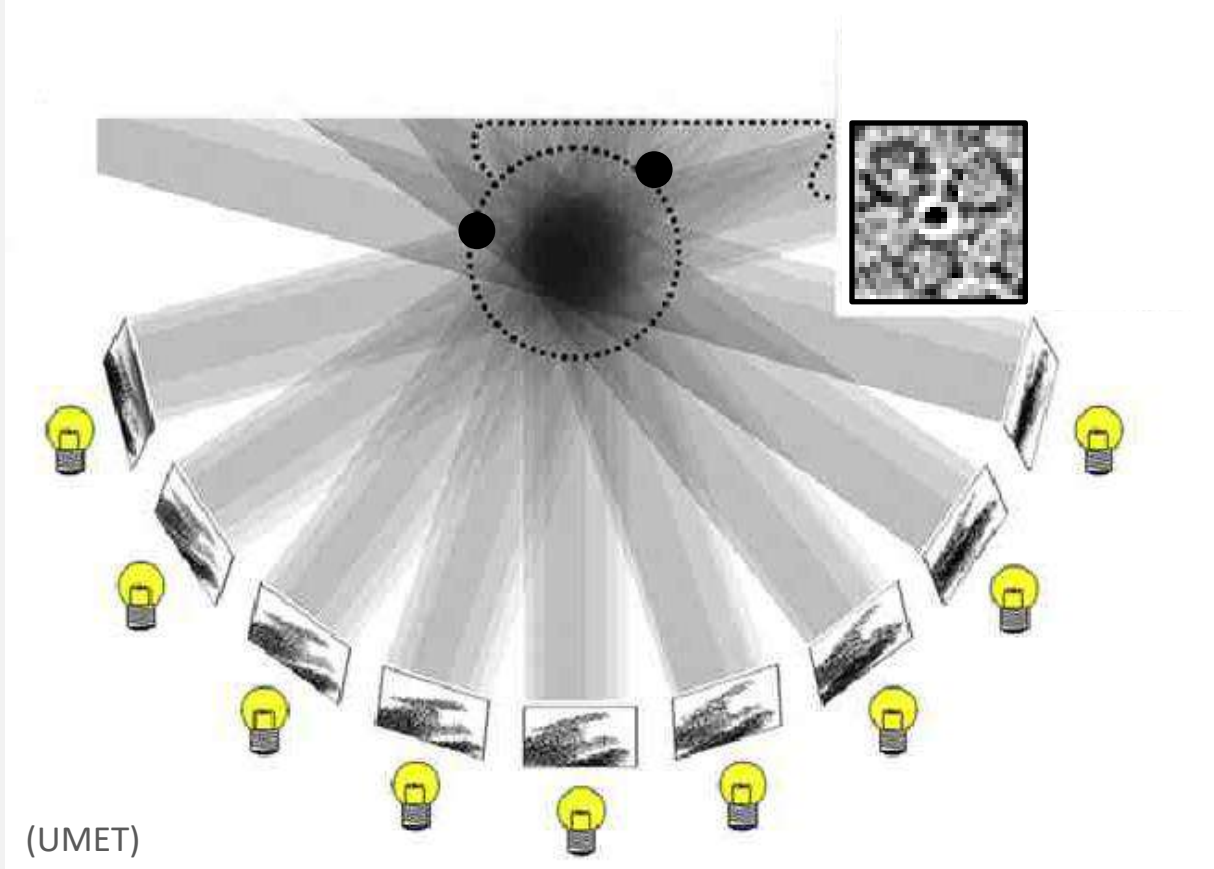
Isotropic and anisotropic fiducial movement relative to the sample (in 3D!)



DE-20 @ 18kx; 51°, 2.34 e⁻/Å² after a cumulative dose of 60 e⁻/Å²

DE-20; 57.5 e⁻/Å², 0° exposure

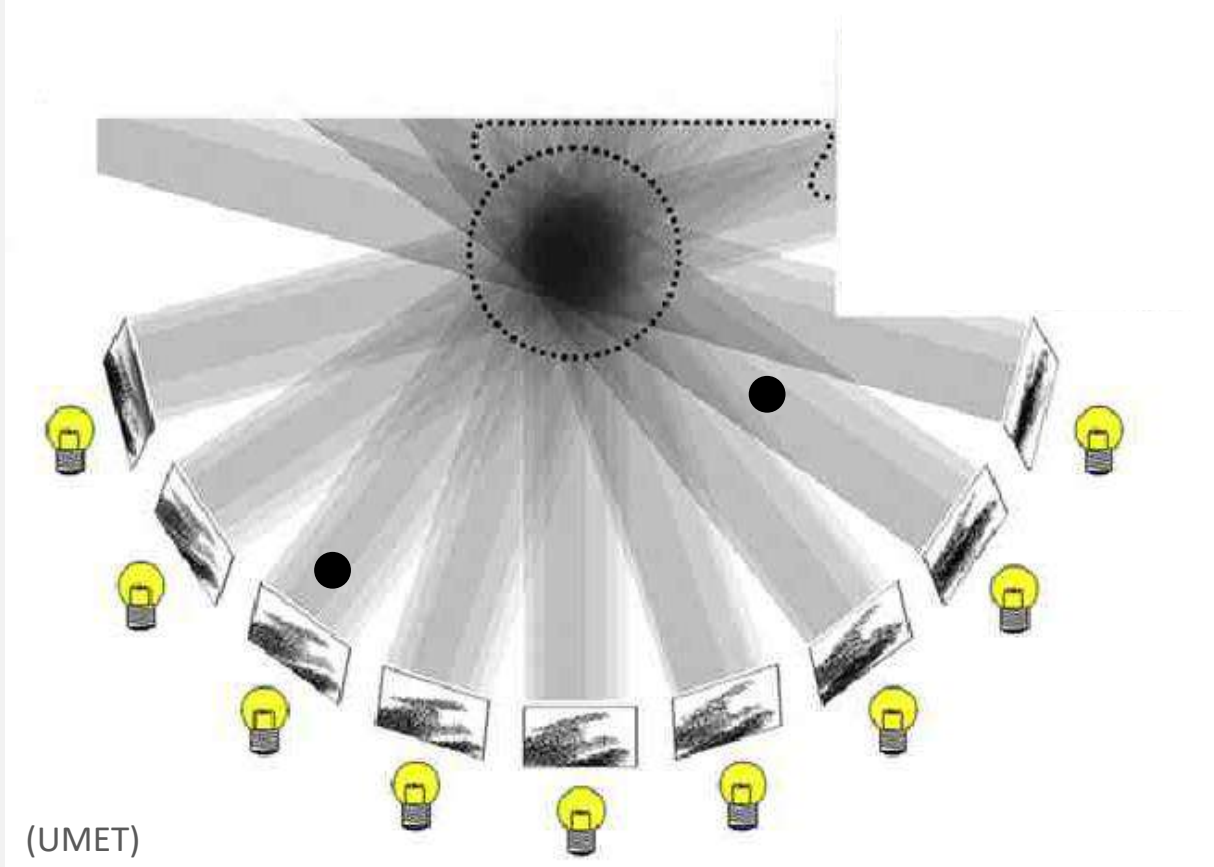
Fiducial-based Tilt-Series Alignment Issues



Nearby Fiducials Affect **Signal** and **Contrast**

- **Fiducial fringes** change the **power spectrum** of your reconstructed object.

Fiducial-based Tilt-Series Alignment Issues



Fiducials are Present in Much of the Reconstruction, *Even if You Can't See Them!*

- **Distant fiducials** can be in the **projection direction** of your extracted object of interest.
- Erasing fiducials isn't perfect.

Tilt-series alignment software

- **Software:**
 - ETomo in IMOD – **Fiducial-based** alignment (also **patch tracking**)
 - Markerauto and AuTom – Automated **fiducial-based** alignment
 - Dynamo – **Fiducial-based** alignment
 - Protomo – **Fiducial-less** alignment
 - AreTomo – **Fiducial-less** alignment, GPU-accelerated
- **Must refine** most or all of the following:
 - Tilt image shifts, rotations, defocus changed, & magnification changes
 - Tilt axis location
 - Tilt angles

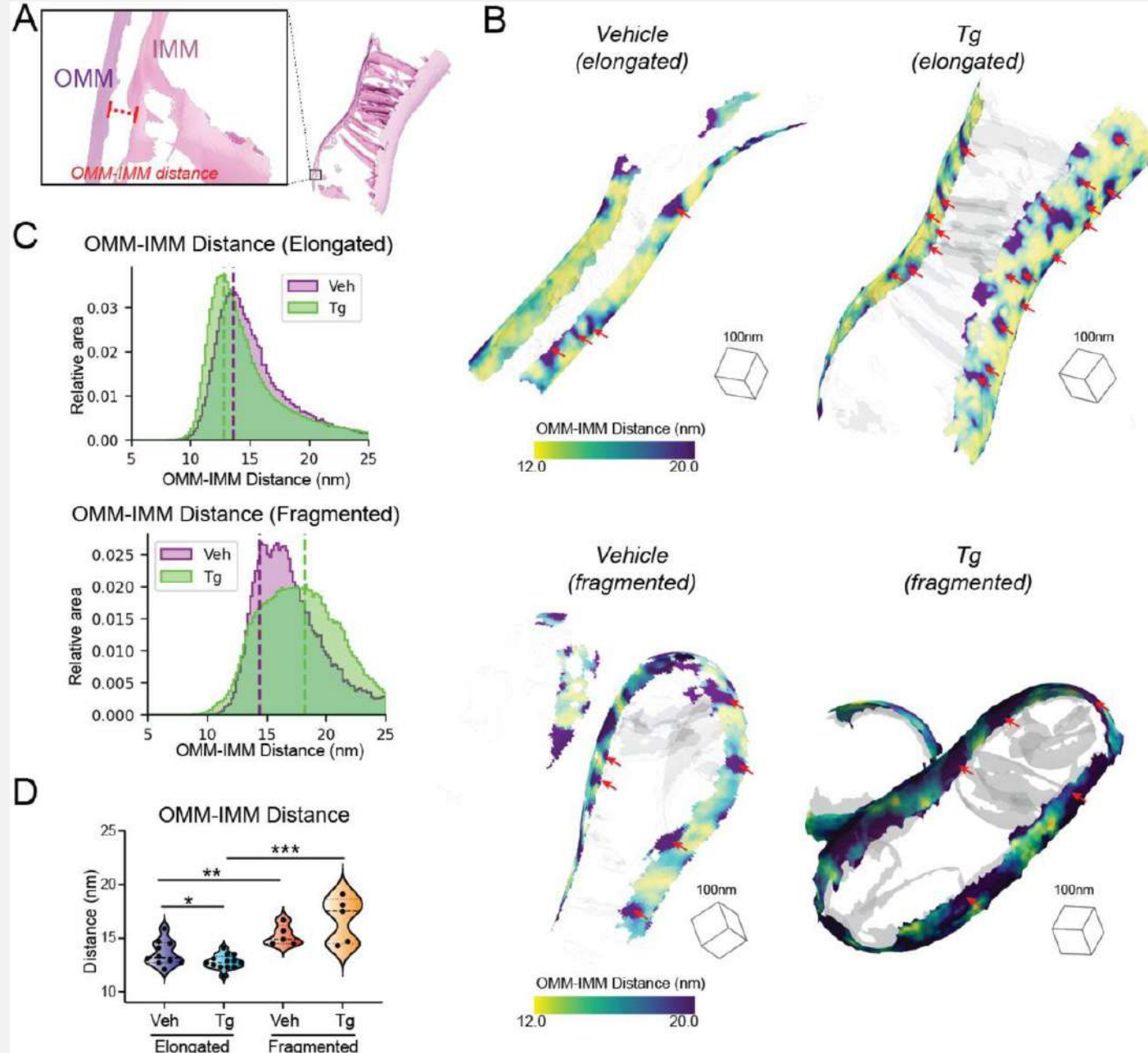


Annotation and Segmentation Software

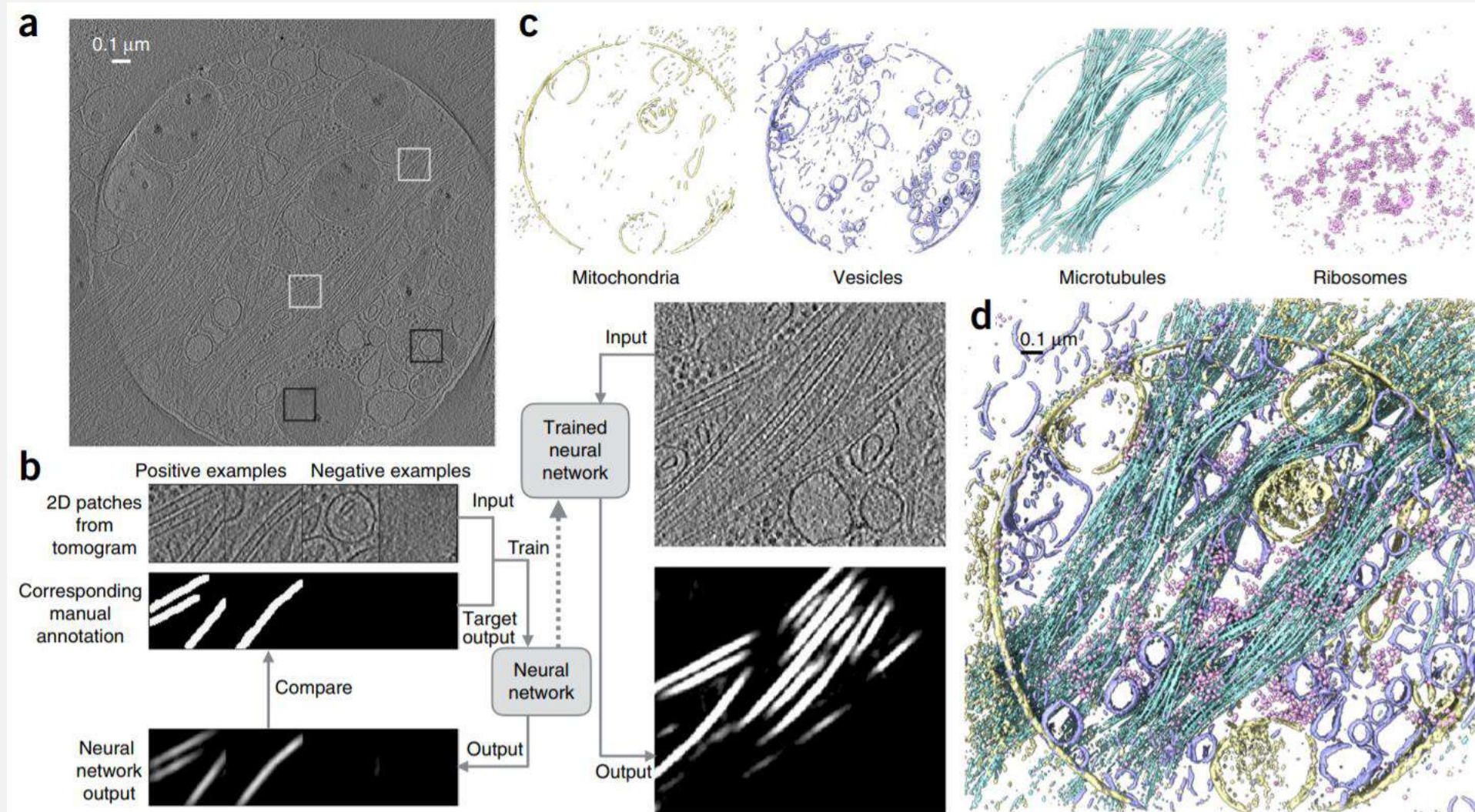
- **Surface morphometrics toolkit** – for mitochondria membrane statistics
- **EMAN2** – Neural network segmentation
- **Dynamo** – Annotate membranes, tubes, helices, crystal structures, vesicles, etc.
- Amira – Interactive segmentation and filtering suite
- Template picking (e.g. MolMatch, Dynamo)
- Difference of Gaussian (DoG) picking



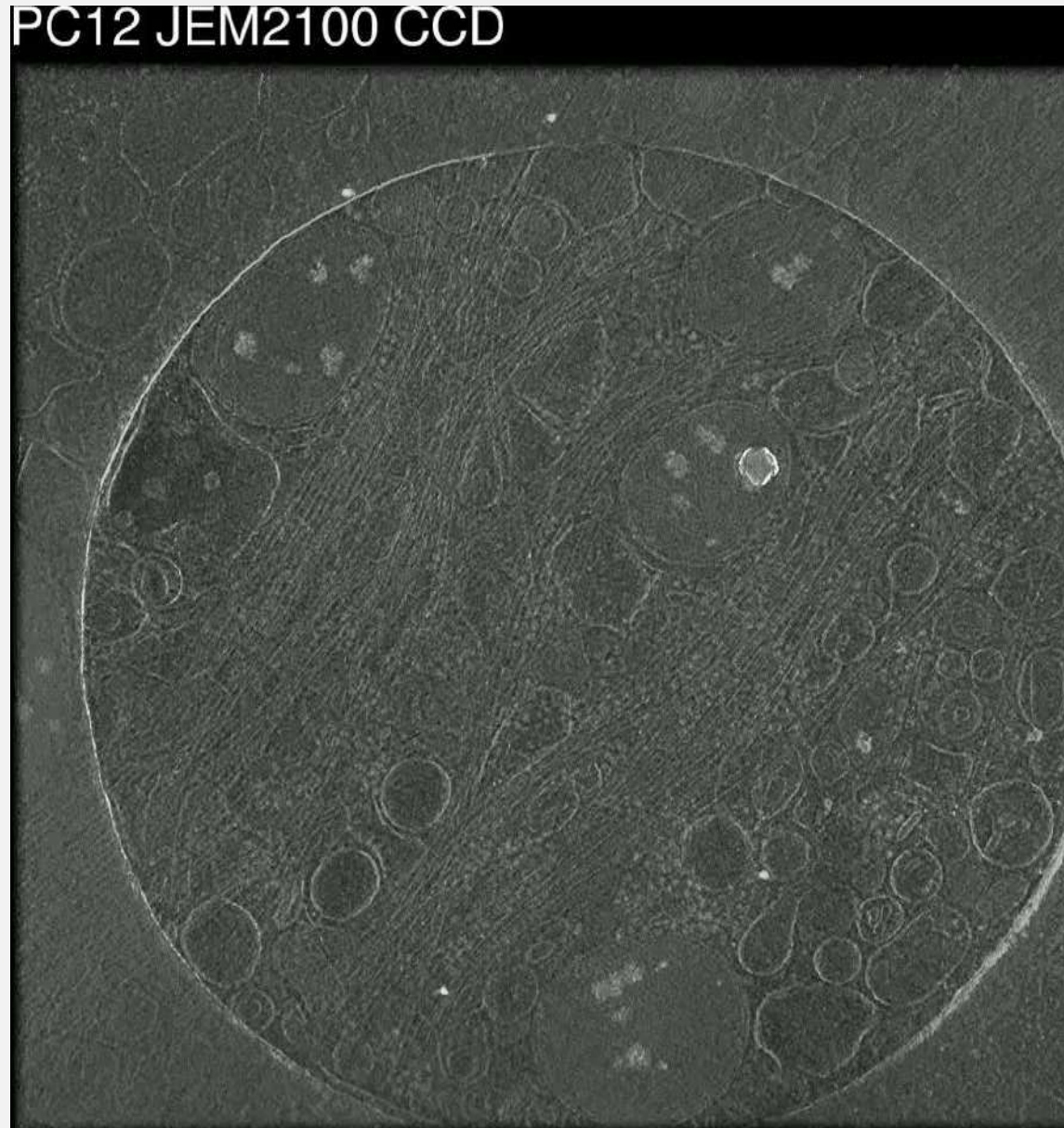
Surface morphometrics toolkit



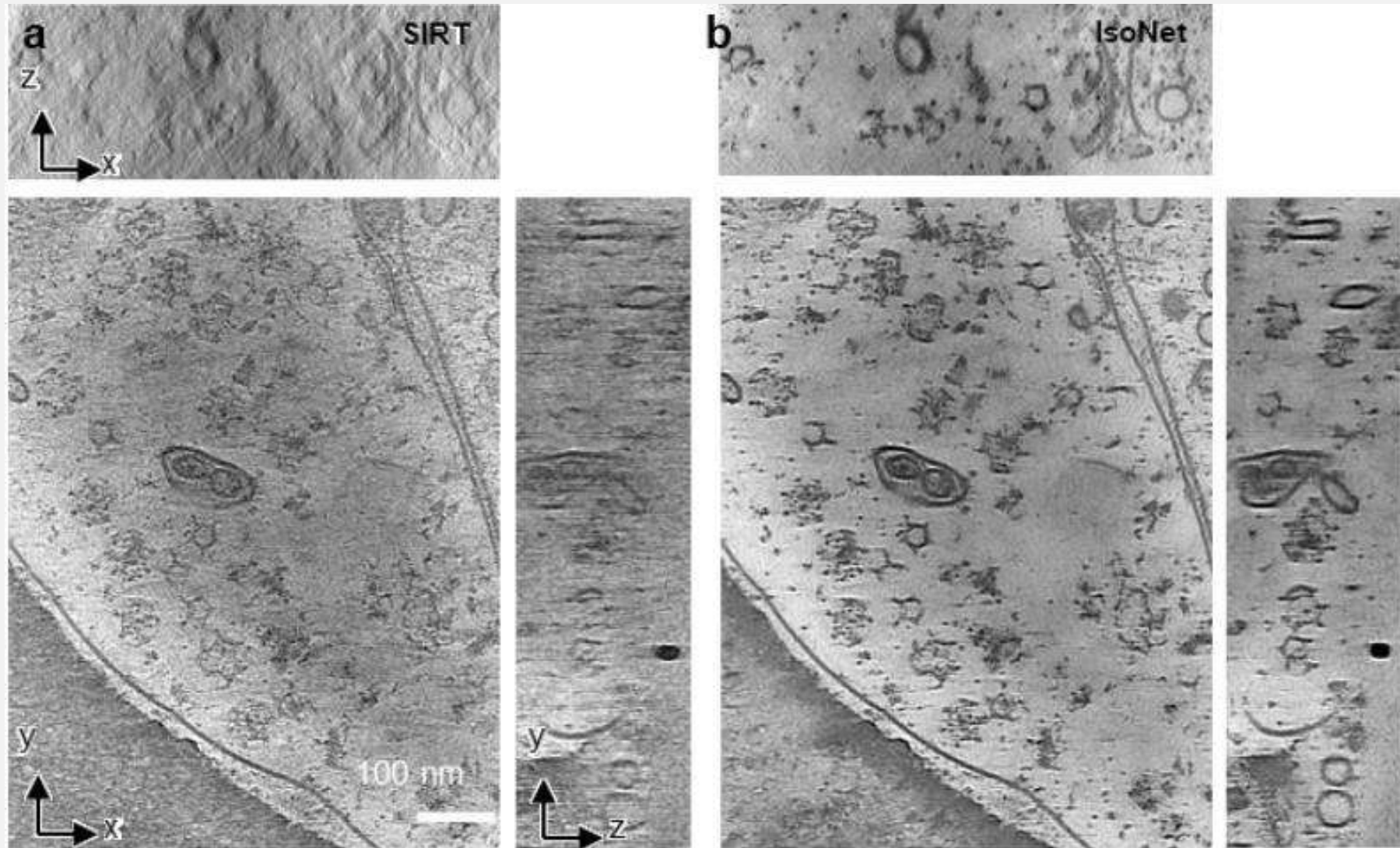
Sub-tomogram segmentation with CNNs in EMAN2



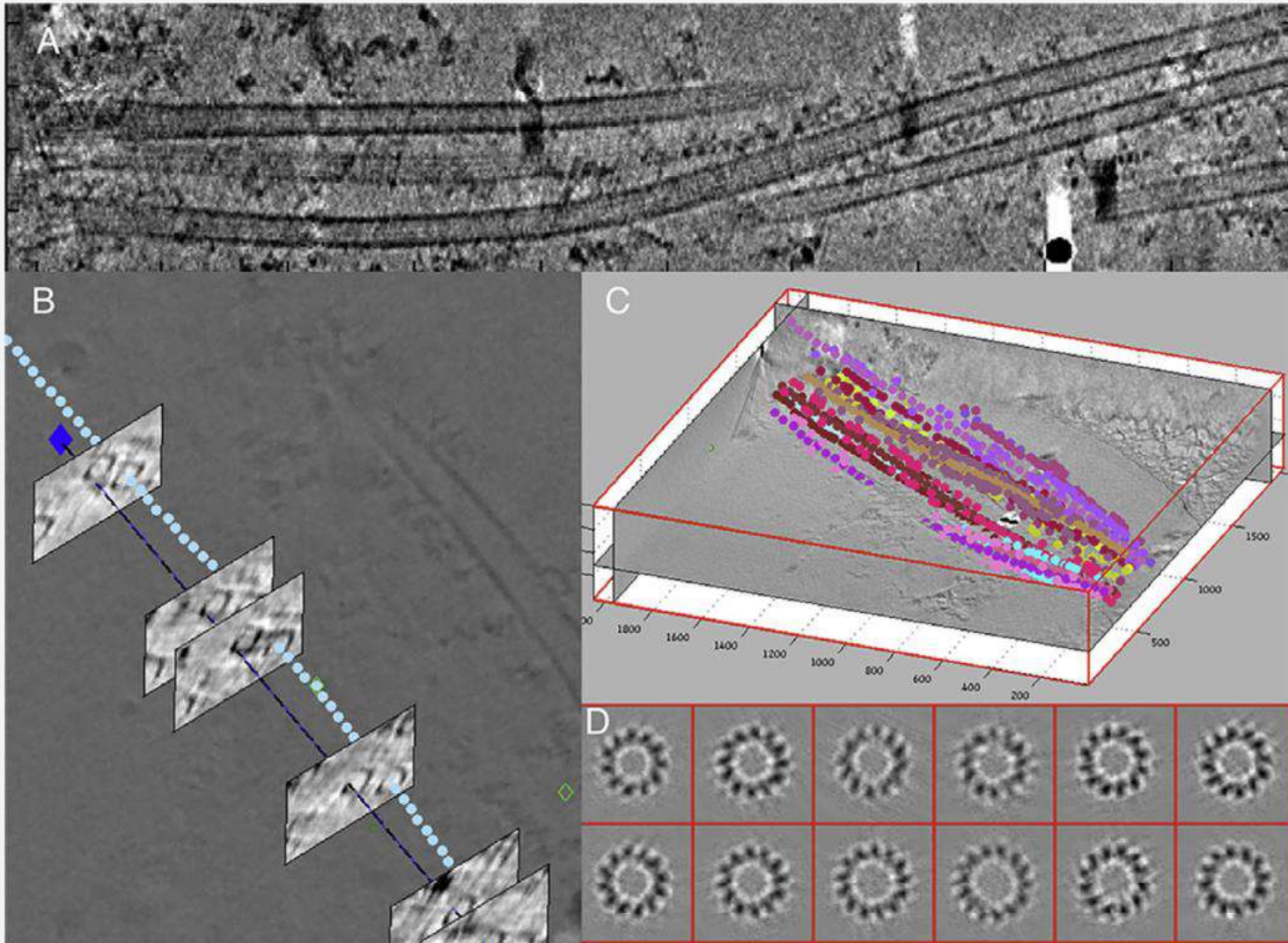
Sub-tomogram segmentation with CNNs in EMAN2



IsoNet Neural Network missing wedge restoration

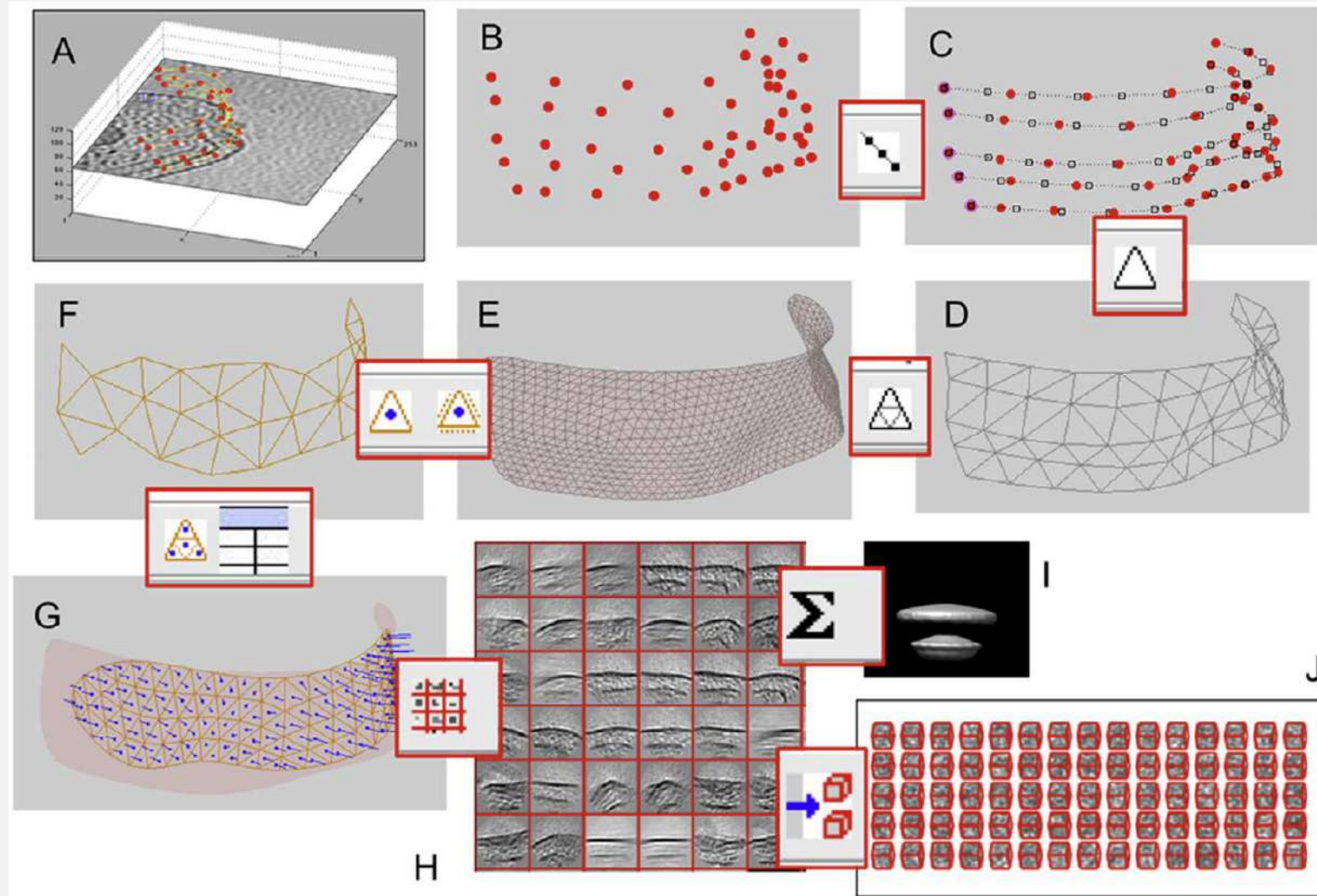


Dynamo Filament Picking

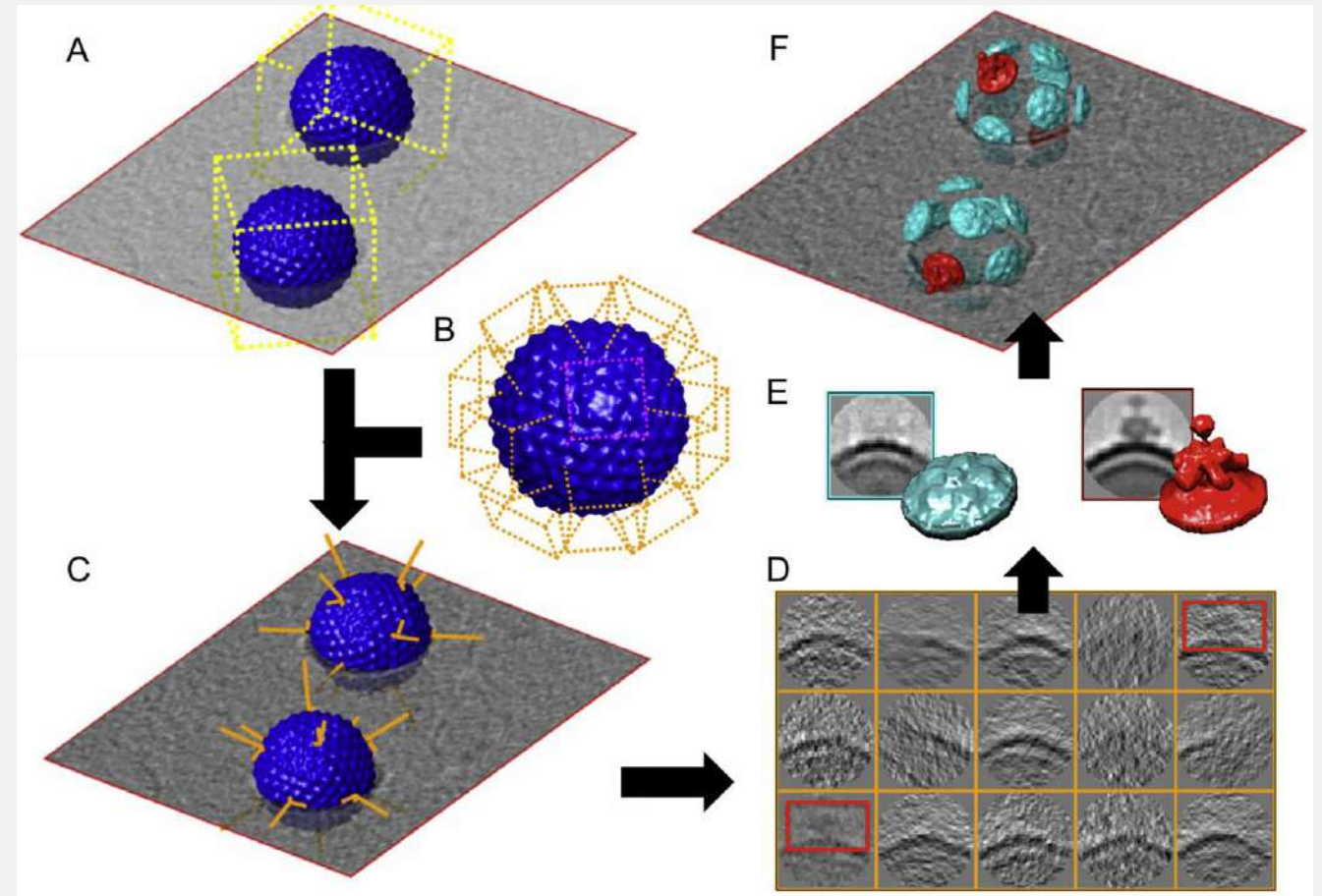
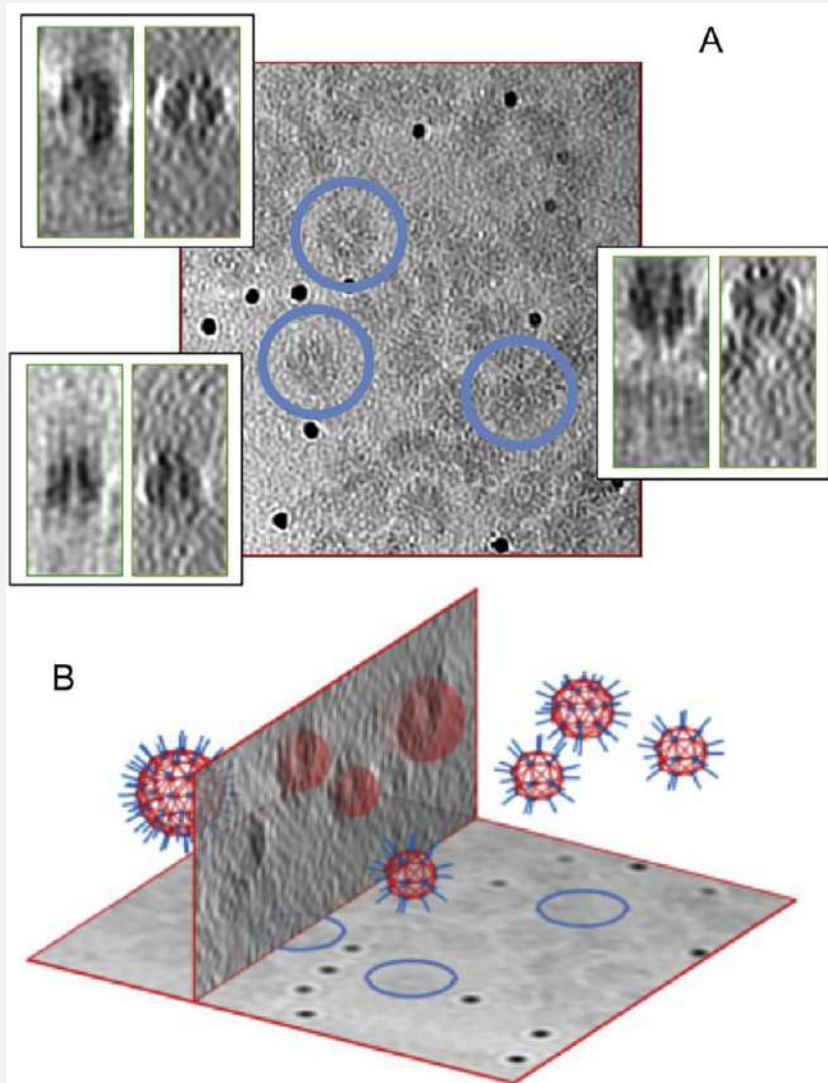


- Backbone, helical, and circumferential picking
- Helical symmetry determination

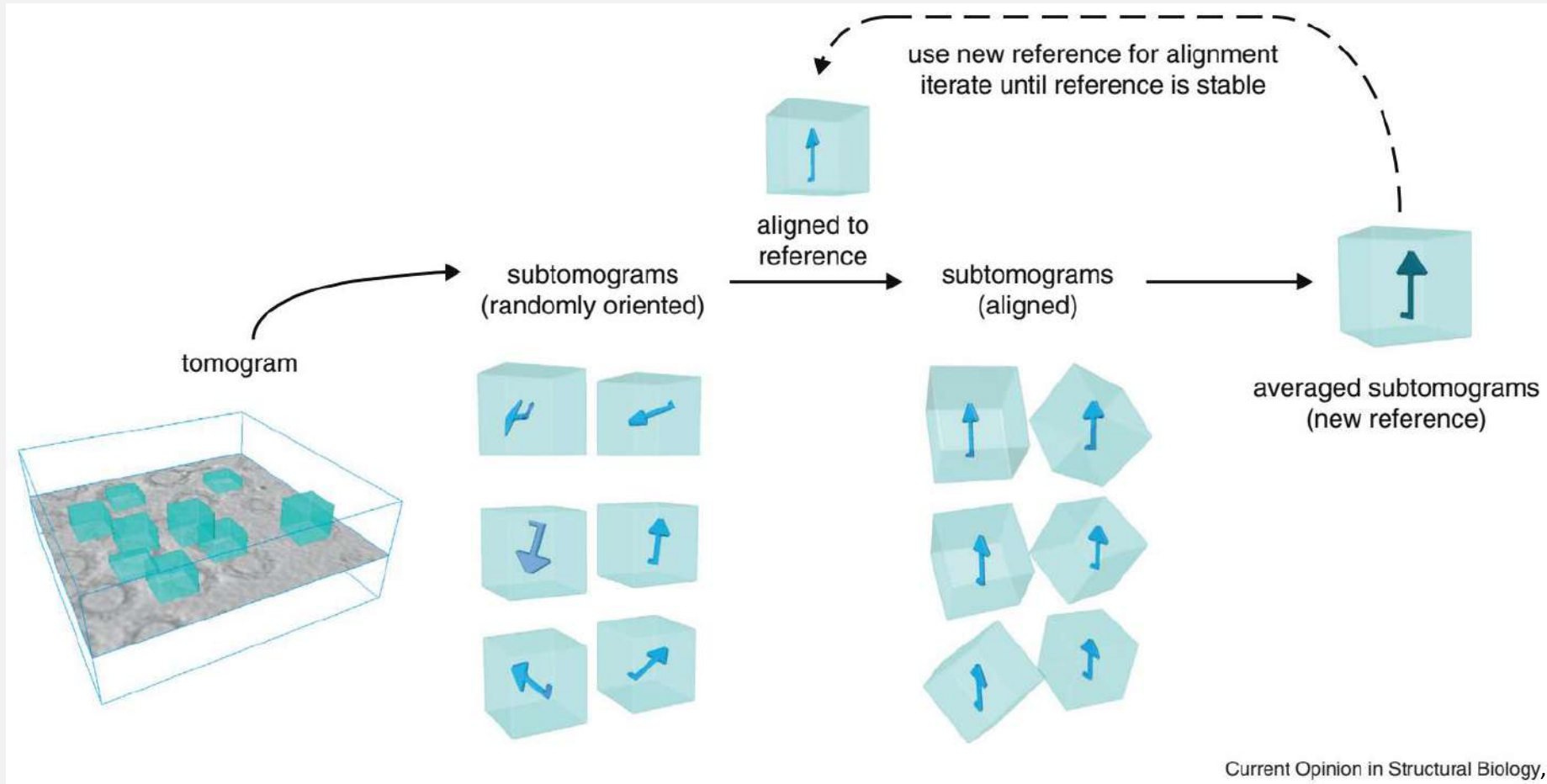
Dynamo Membrane Picking



Dynamo Vesicle and Sub-Particle Picking



Sub-tomogram Processing Workflow



- Missing wedge must be taken into account for each sub-tomogram

Sub-tomogram Processing Software

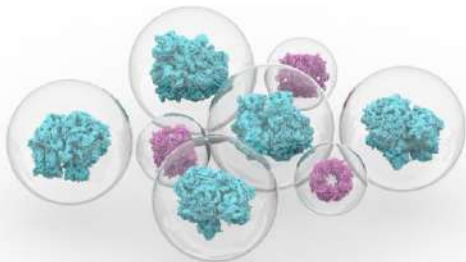
- Warp/M - Sub-tilt-series refinement, multi-species co-refinement
- Relion4 – Sub-tilt-series refinement, Bayesian approach to alignment is used
- EMAN2 – Sub-tilt-series refinement and defocus estimation
- emClarity – Sub-tilt-series refinement and defocus estimation
- Dynamo – GPU accelerated, tomogram database, extensive picking abilities
- PyTom
- PEET
- Jsubtomo
- TOM & AV3
- XMIPP



Warp/M sub-tilt multi-species refinement

a

Single particles, optimized separately

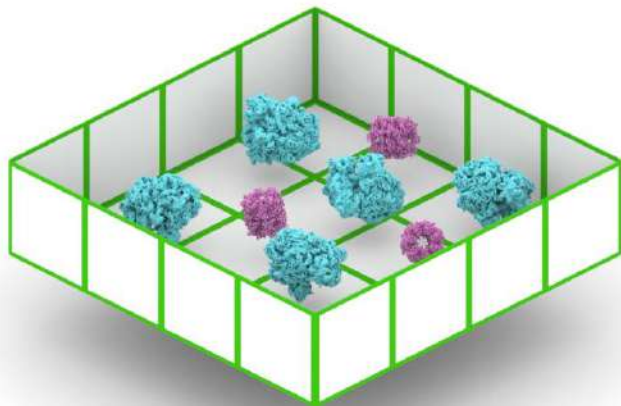


$$S_1 = \text{Projection}(\text{pose}_1) \cdot \text{Image}_1$$

\vdots

$$S_n = \text{Projection}(\text{pose}_n) \cdot \text{Image}_n$$

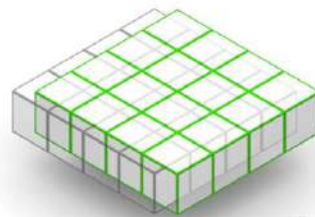
Multi-particle system, optimized simultaneously



$$M = \sum_s^{N_{\text{species}}} \sum_p^{N_{\text{particles}}} \sum_f^{N_{\text{frames}}} \text{Projection}(\text{pose}_{s,p,f} + \text{correction}_{s,p,f}) \cdot \text{Image}_{s,p,f}$$

b

Translation



Rotation

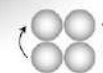
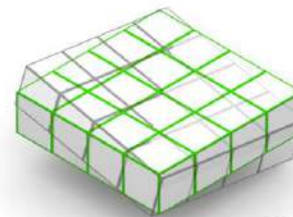
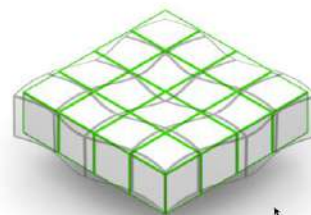
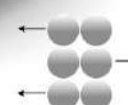
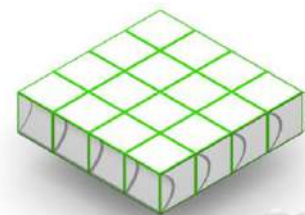


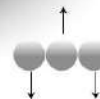
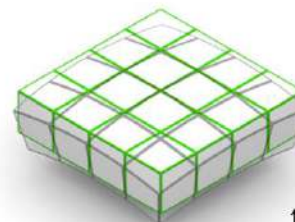
Image-space
warping



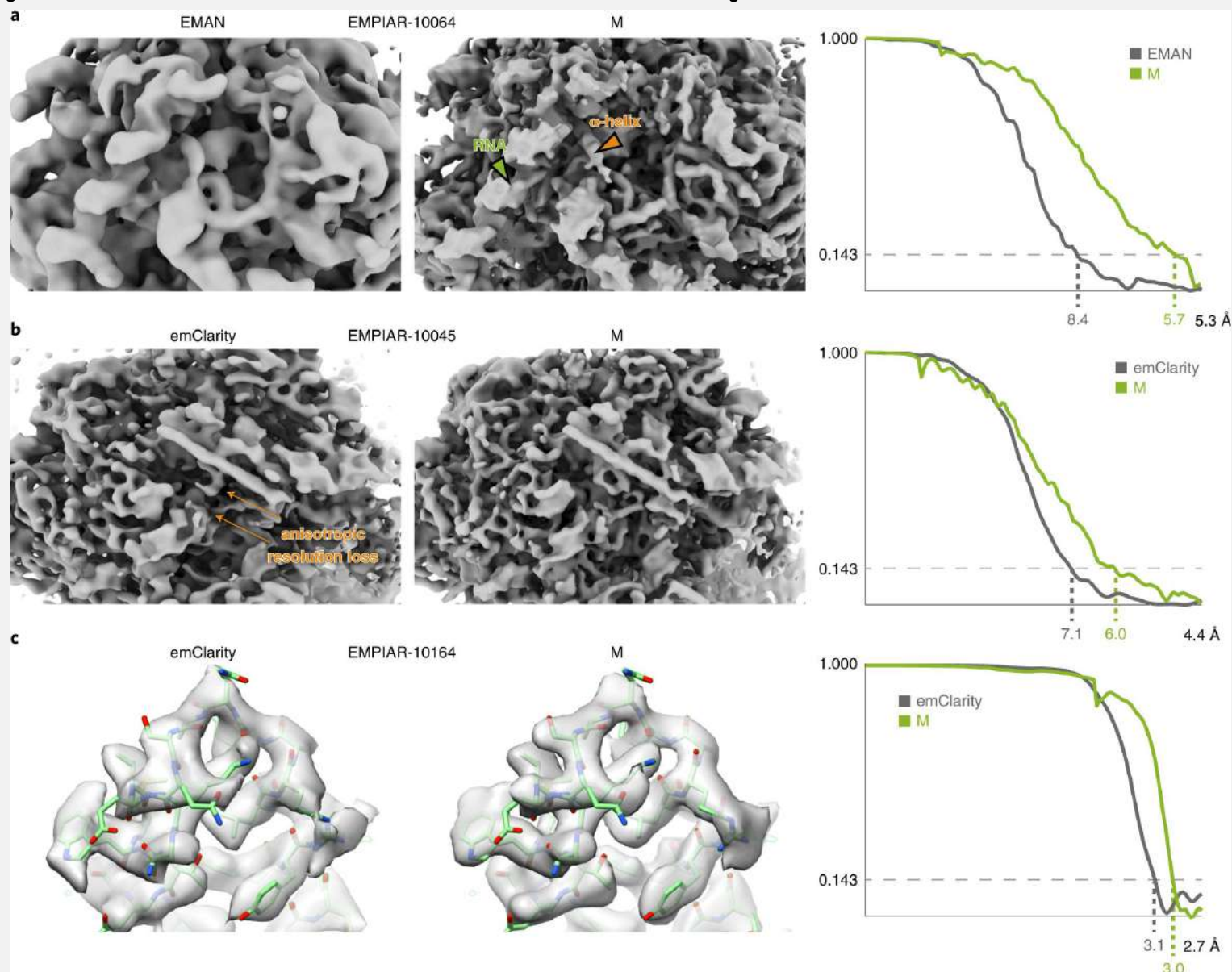
Volume-space
warping



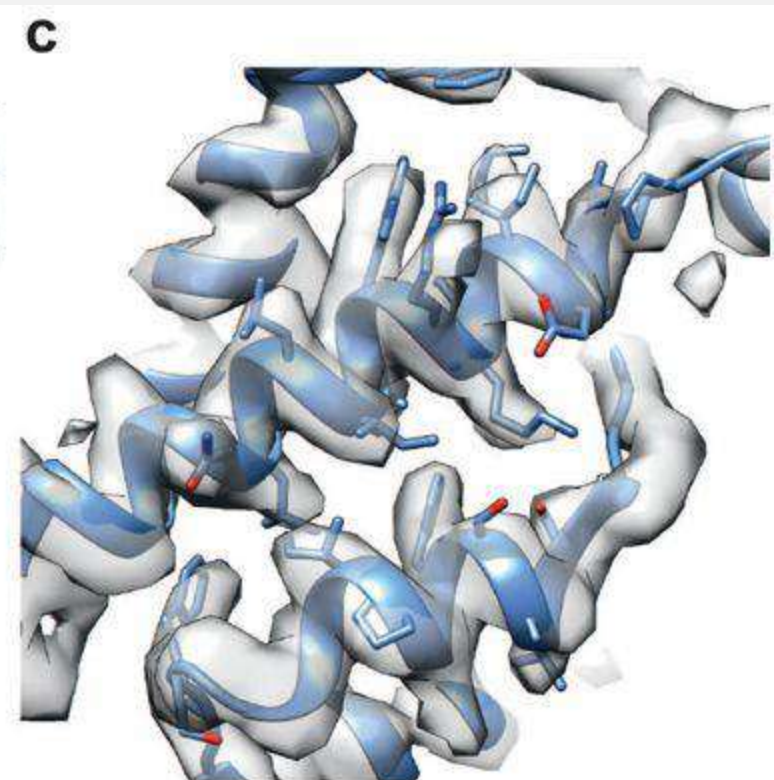
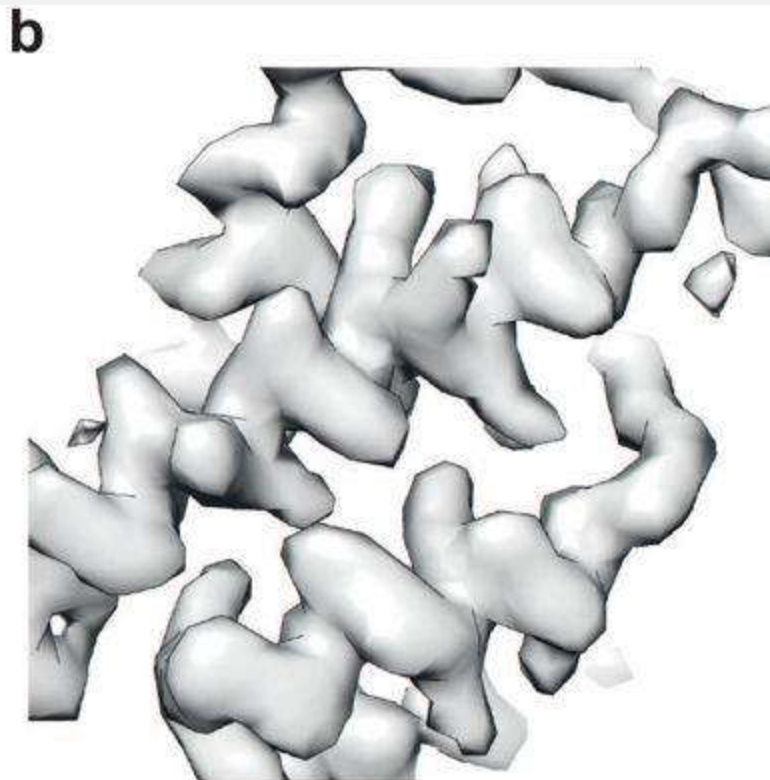
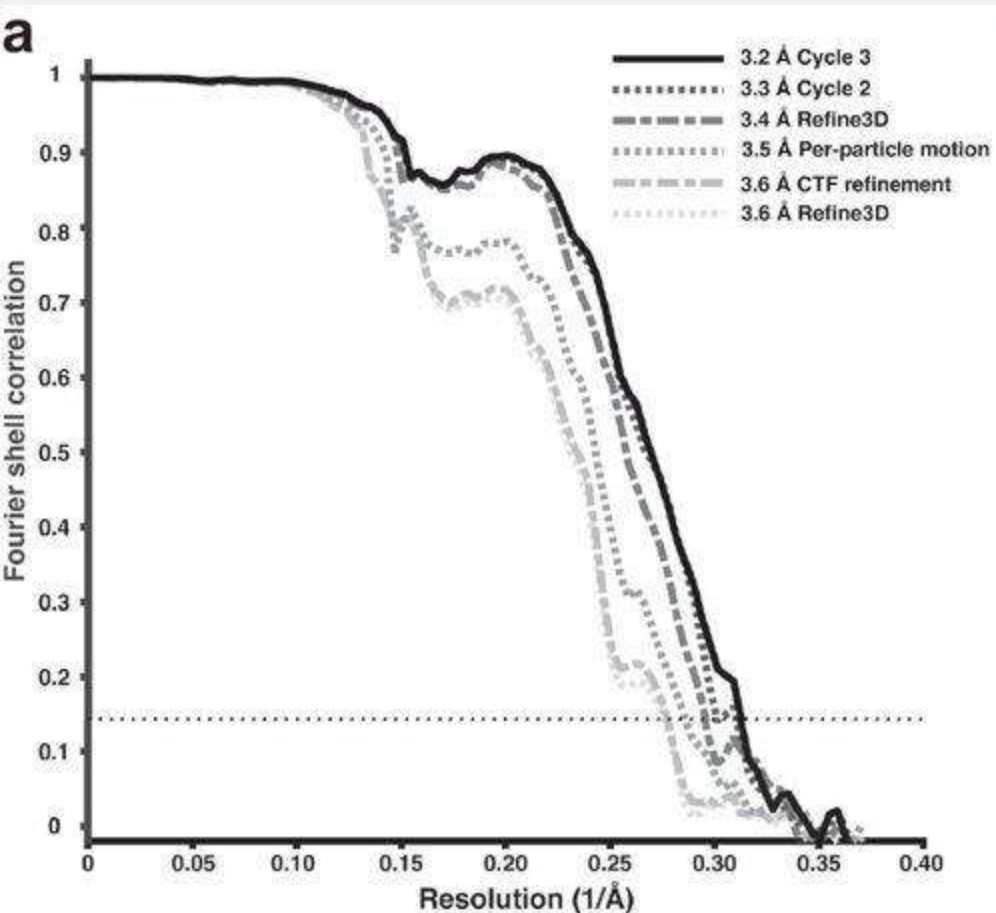
Doming



Warp/M sub-tilt multi-species refinement



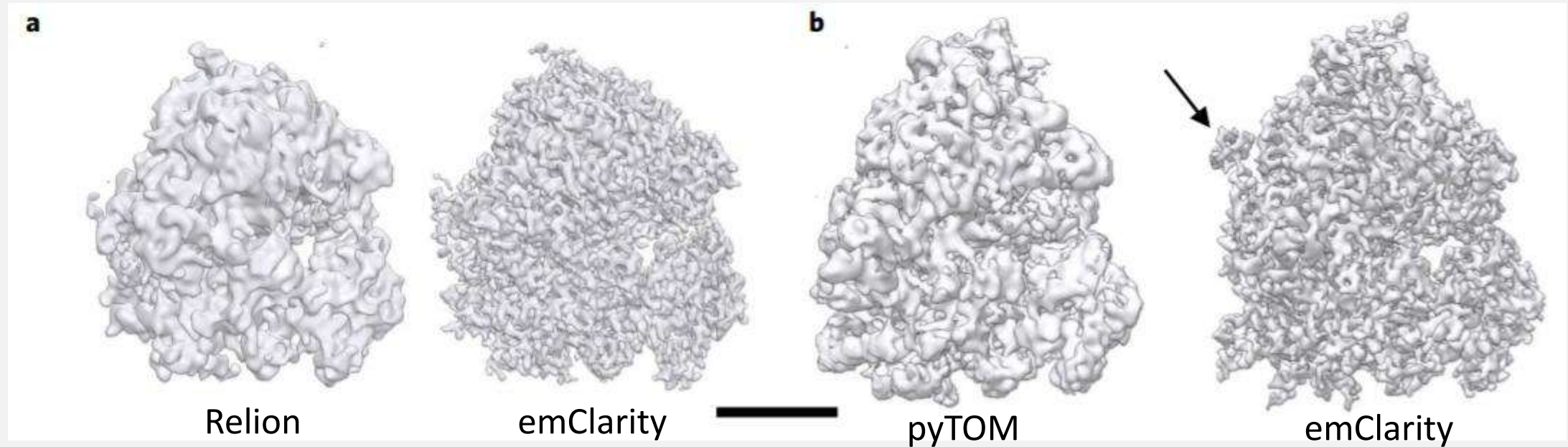
Relion4 sub-tilt refinement



emClarity sub-tilt series refinement

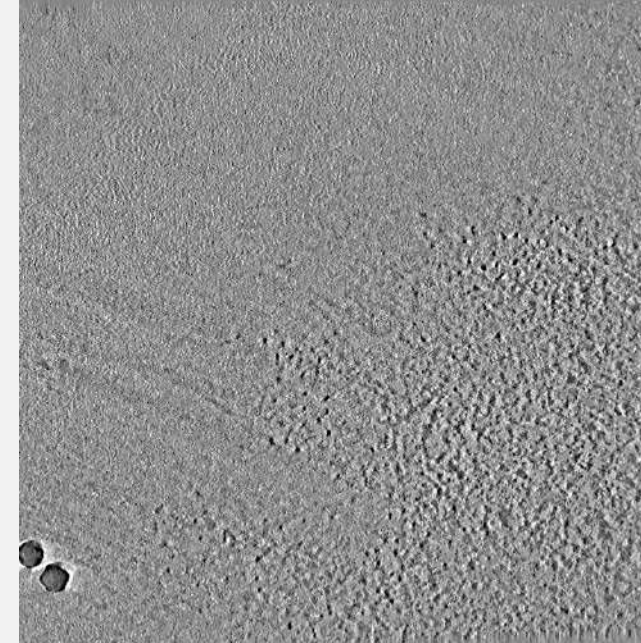
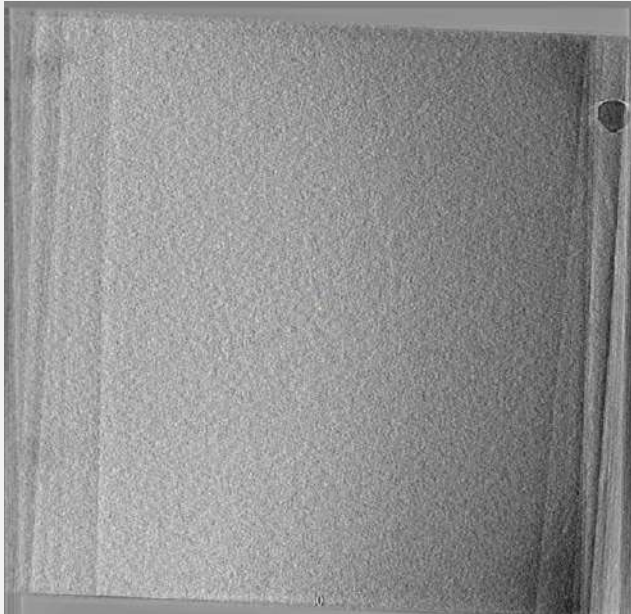
Yeast 80S ribosome

Rabbit 80S ribosome

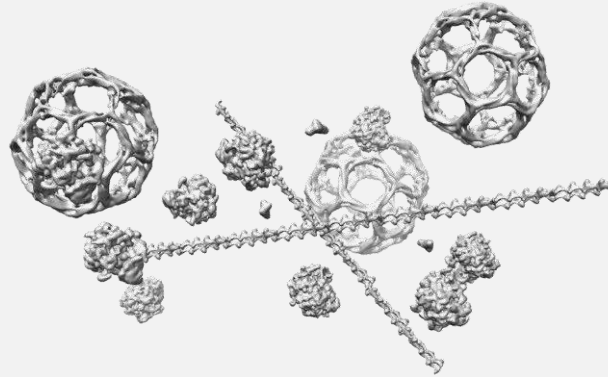
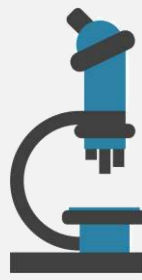


Questions?

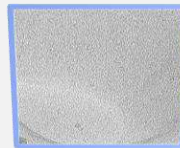
Next: Appion-Protomo description



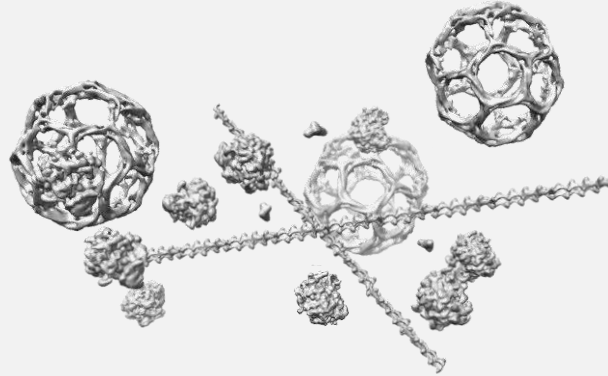
3D specimen movement during collection



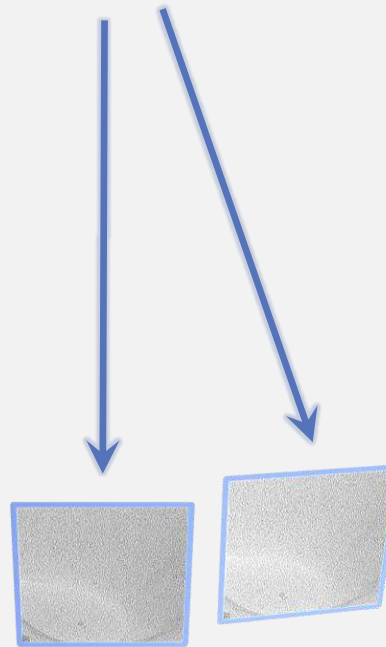
(movements are exaggerated)



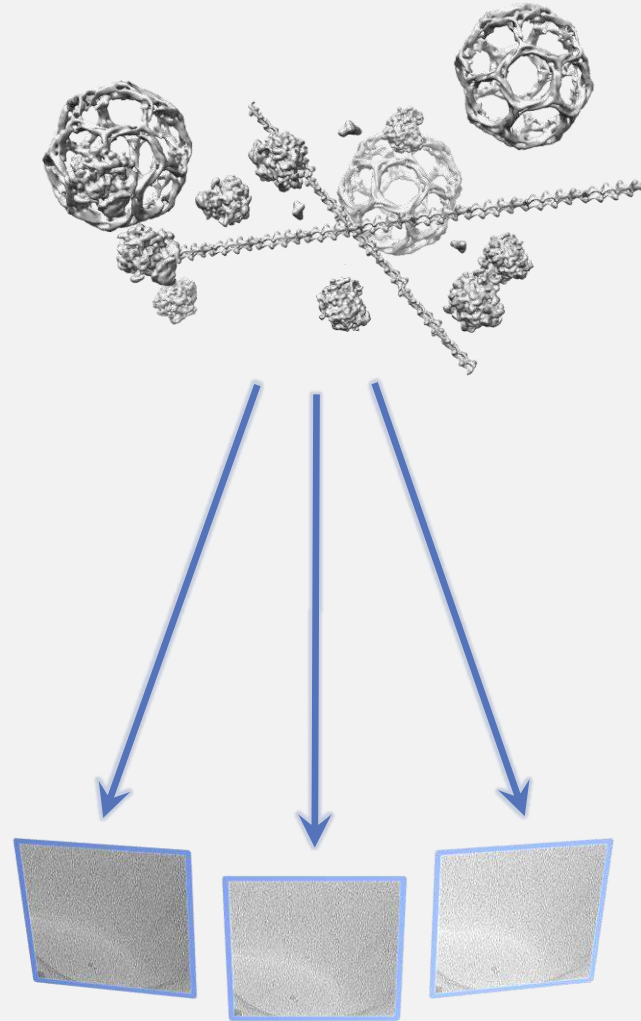
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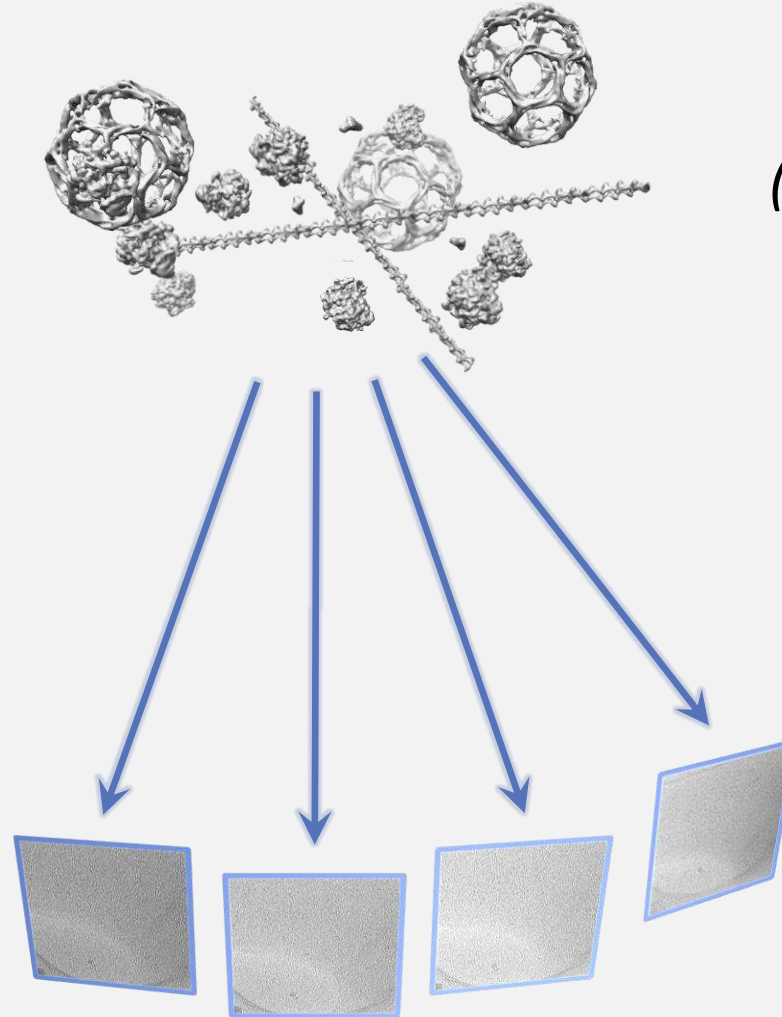


3D specimen movement during collection



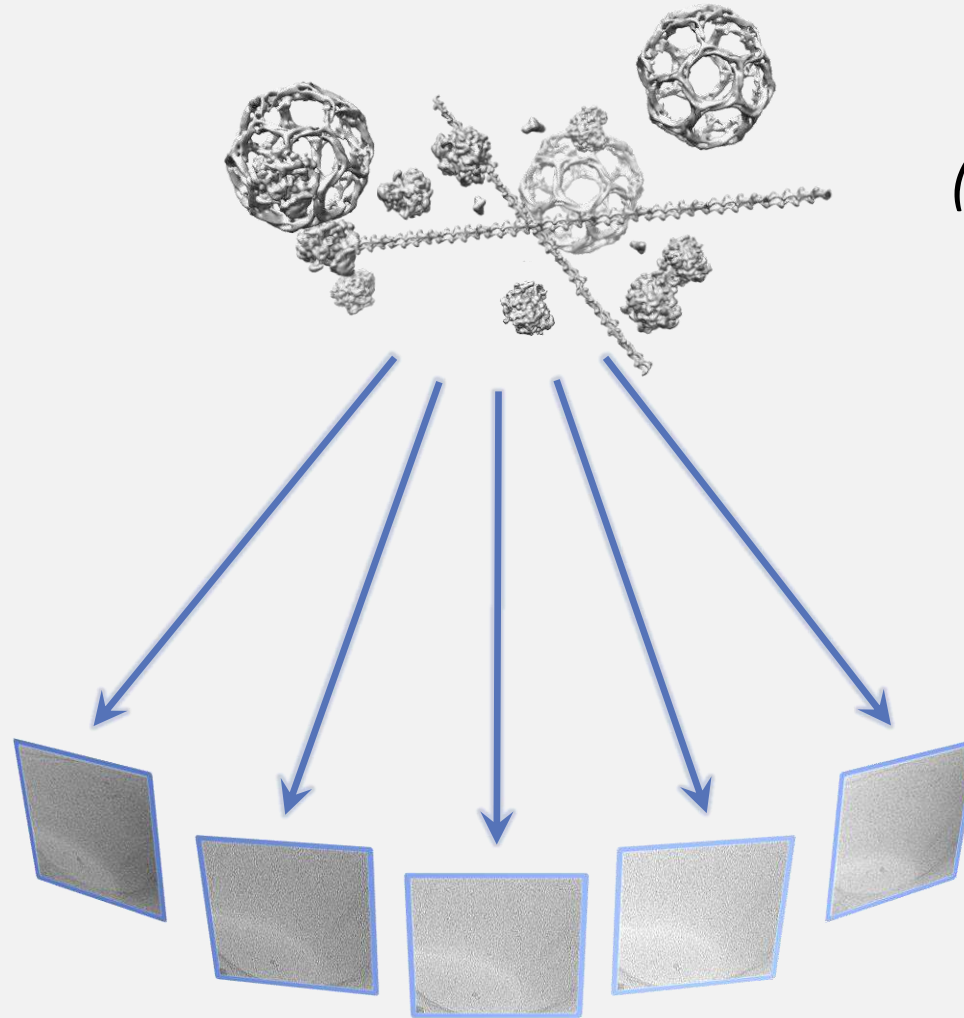
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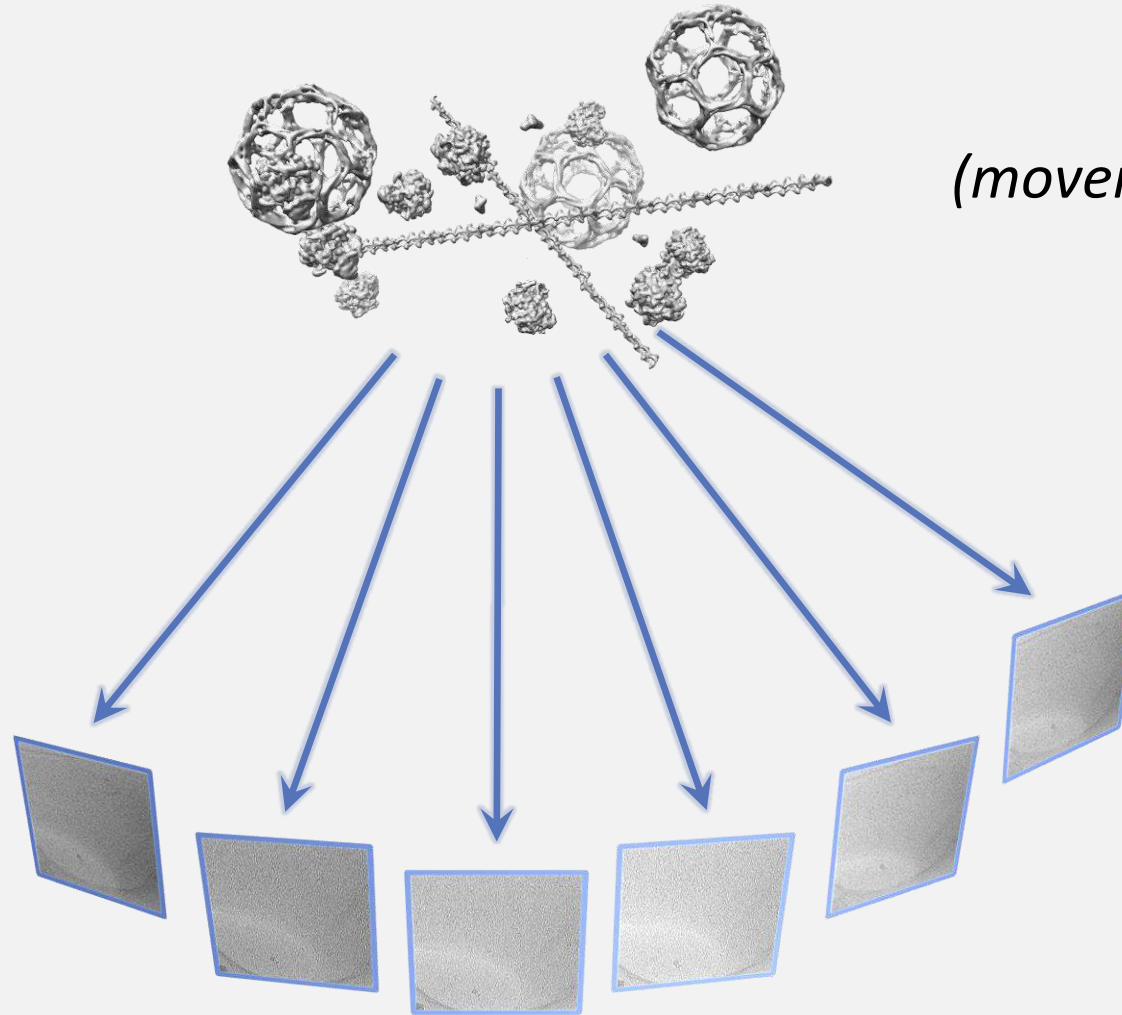
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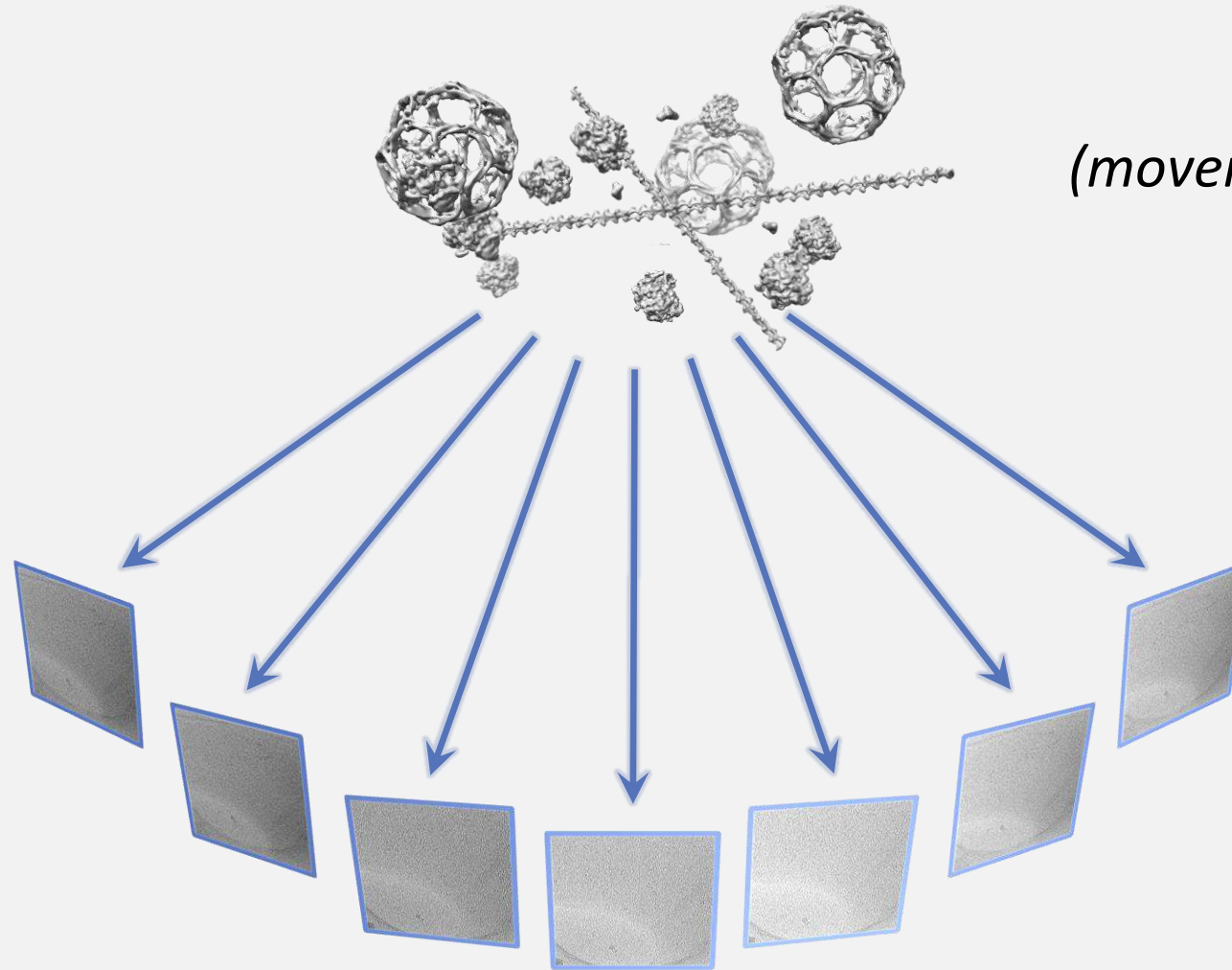
(movements are exaggerated)

3D specimen movement during collection



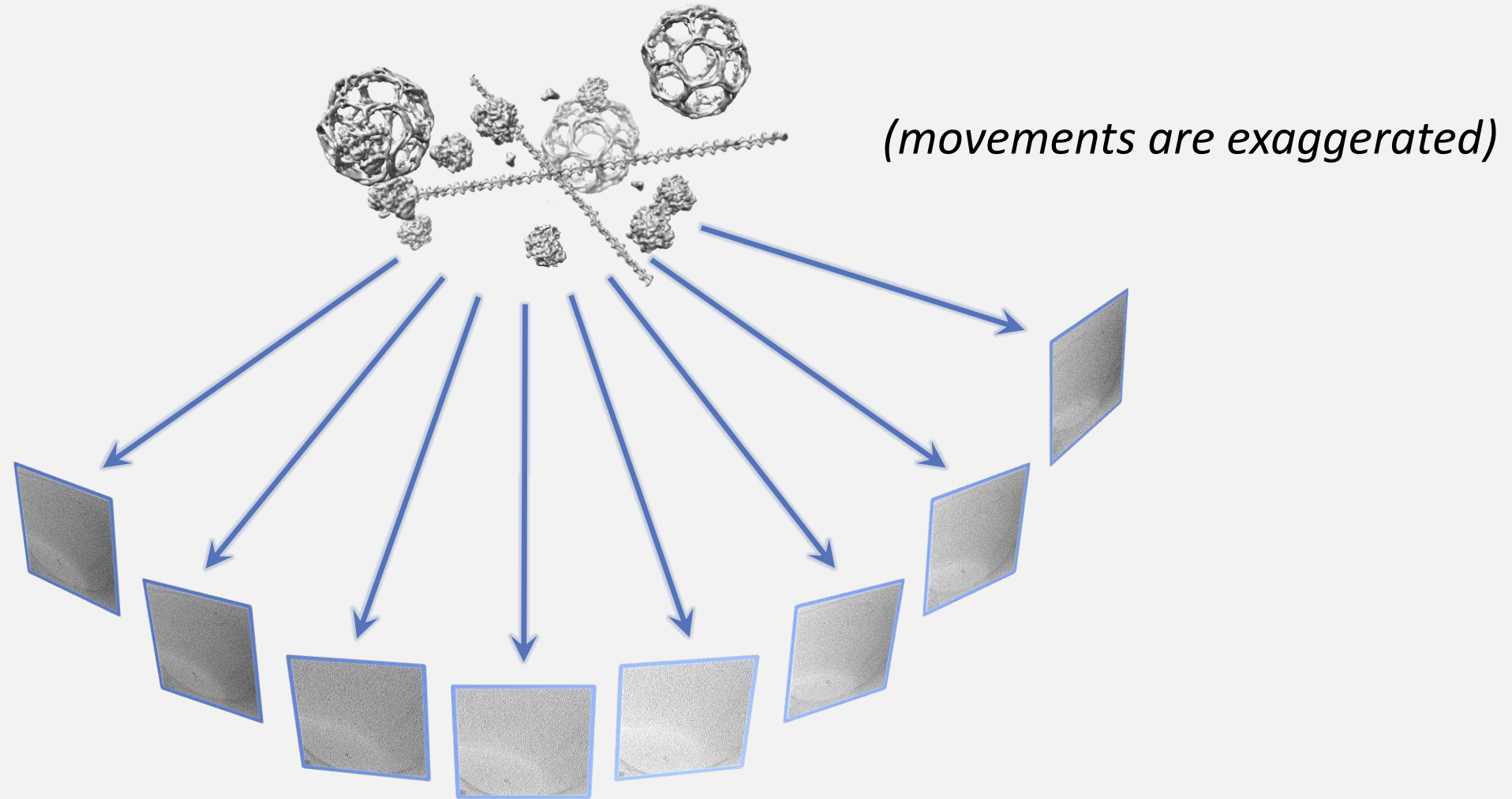
(movements are exaggerated)

3D specimen movement during collection

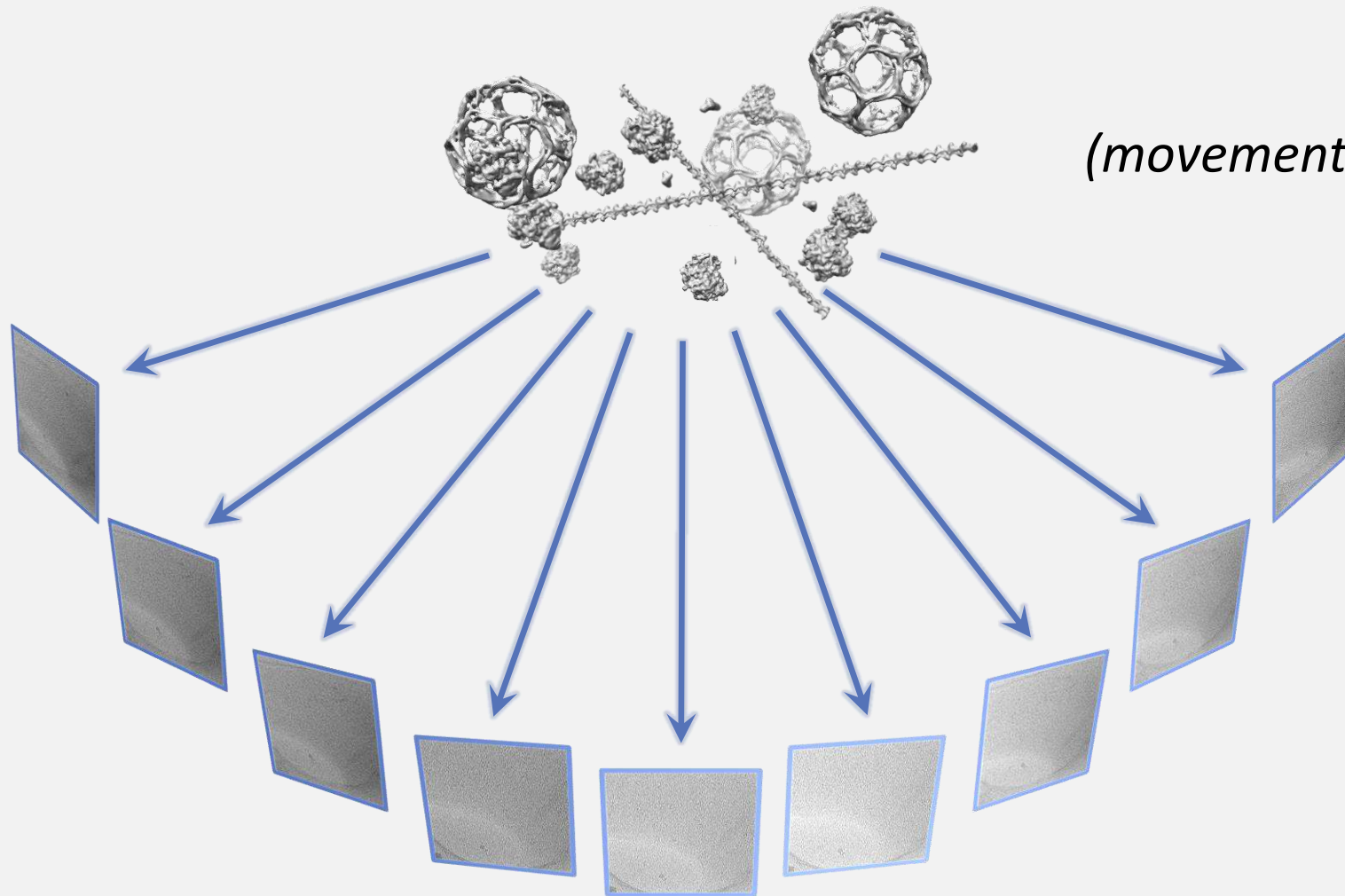


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3D specimen movement during collection



3D specimen movement during collection



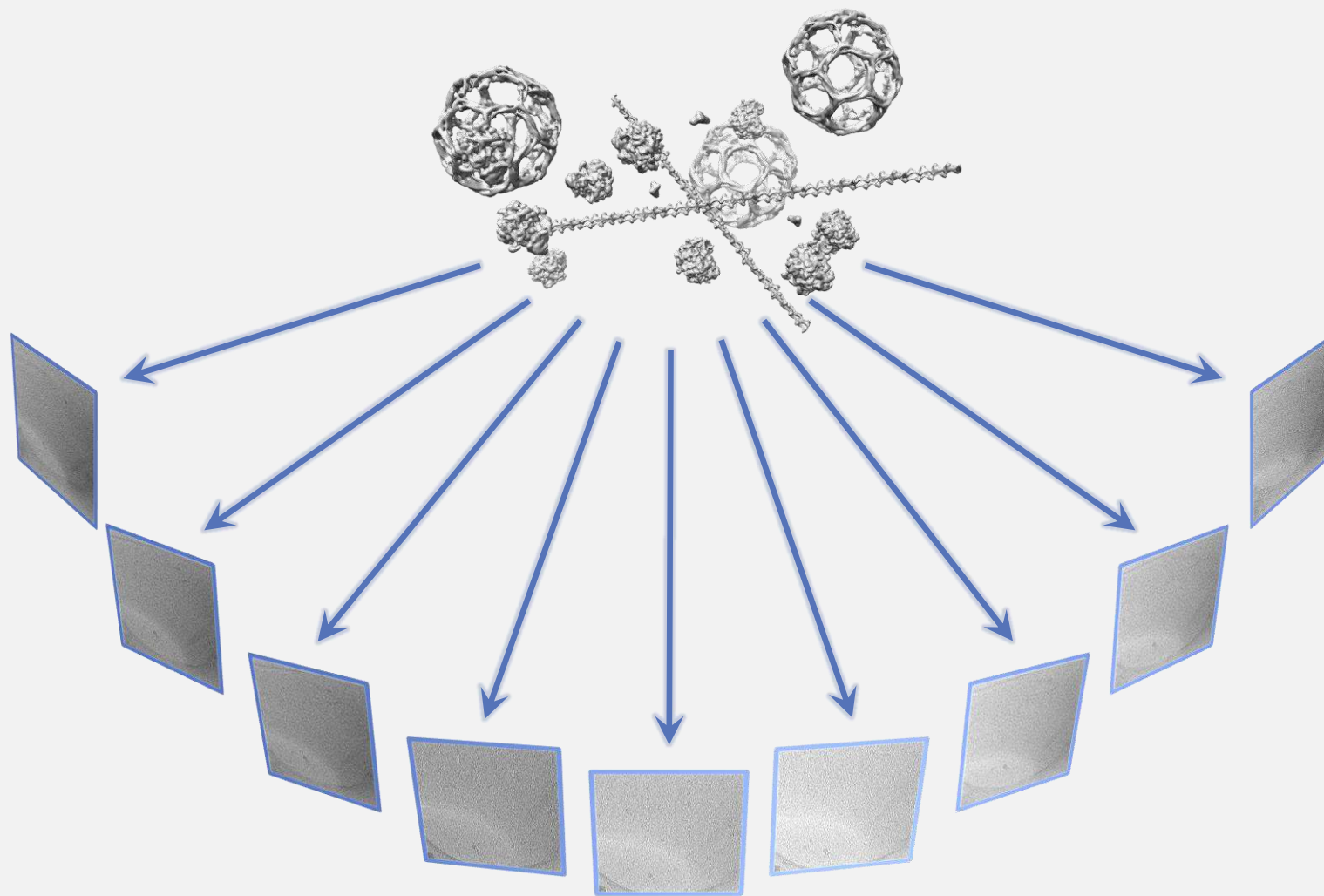
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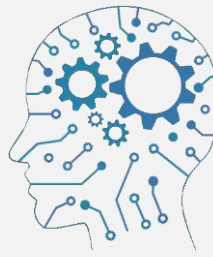
How does alignment in Protomo work?



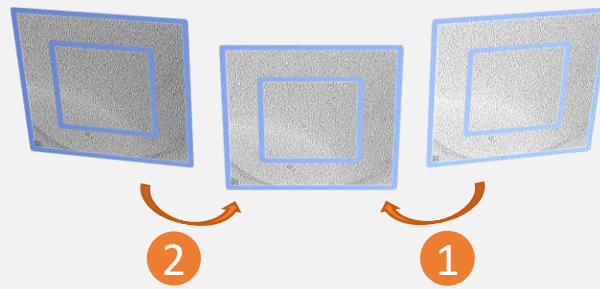


Collect a tilt-series

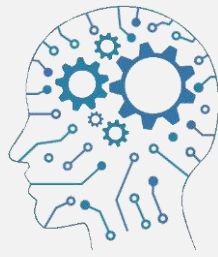




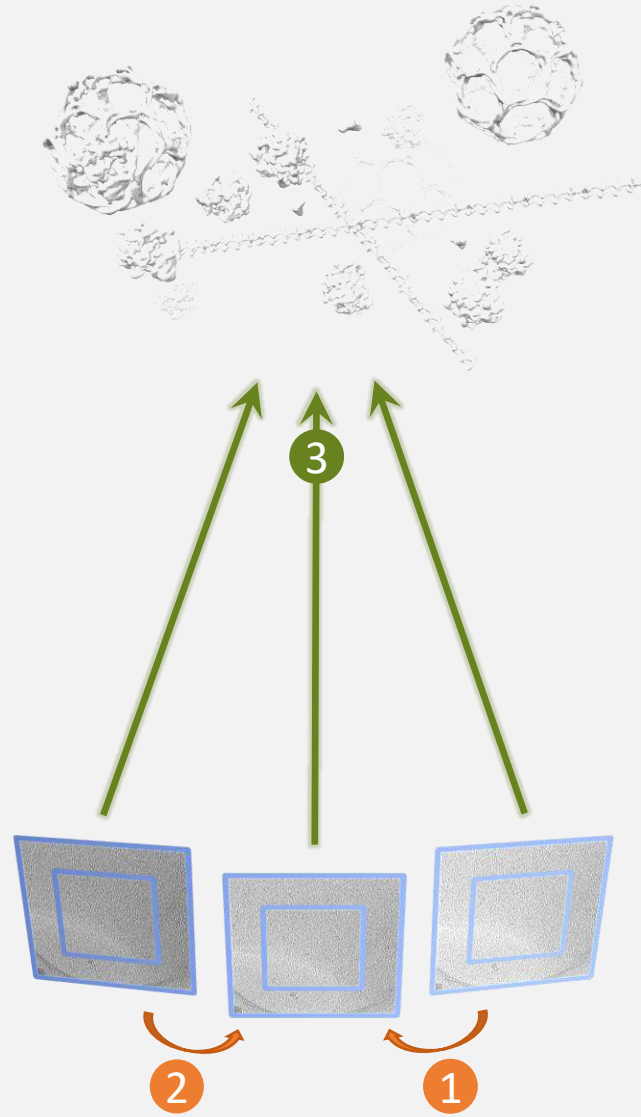
Protomo alignment



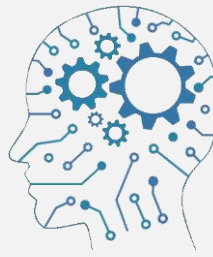
● Nearest-neighbor correlation



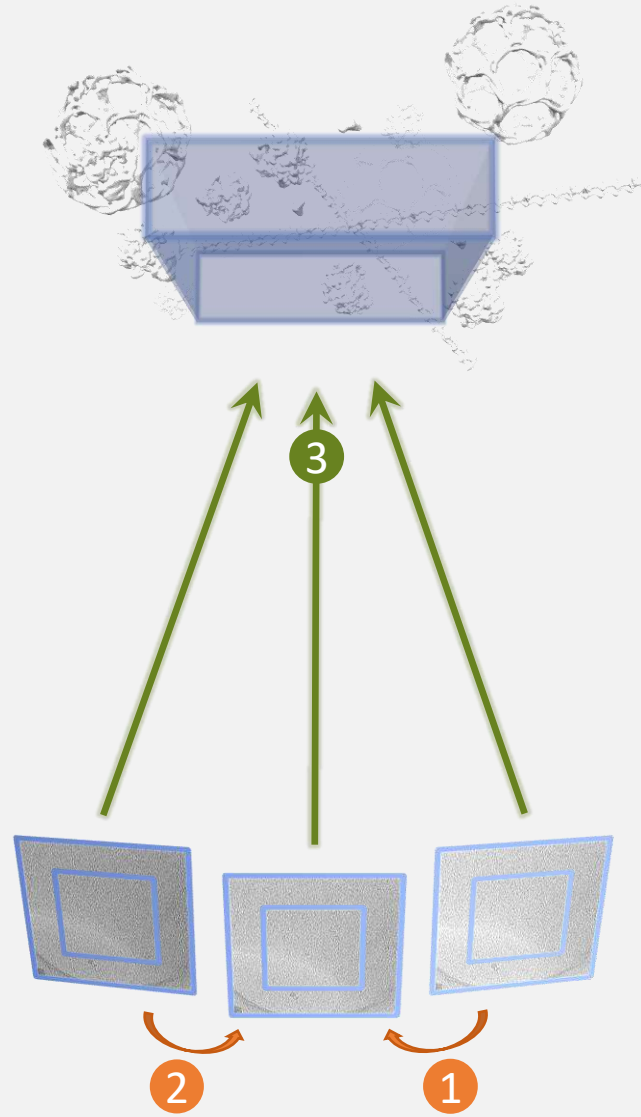
Protomo alignment




- Nearest-neighbor correlation
- Weighted back-projection

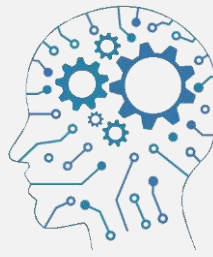


Protomo alignment

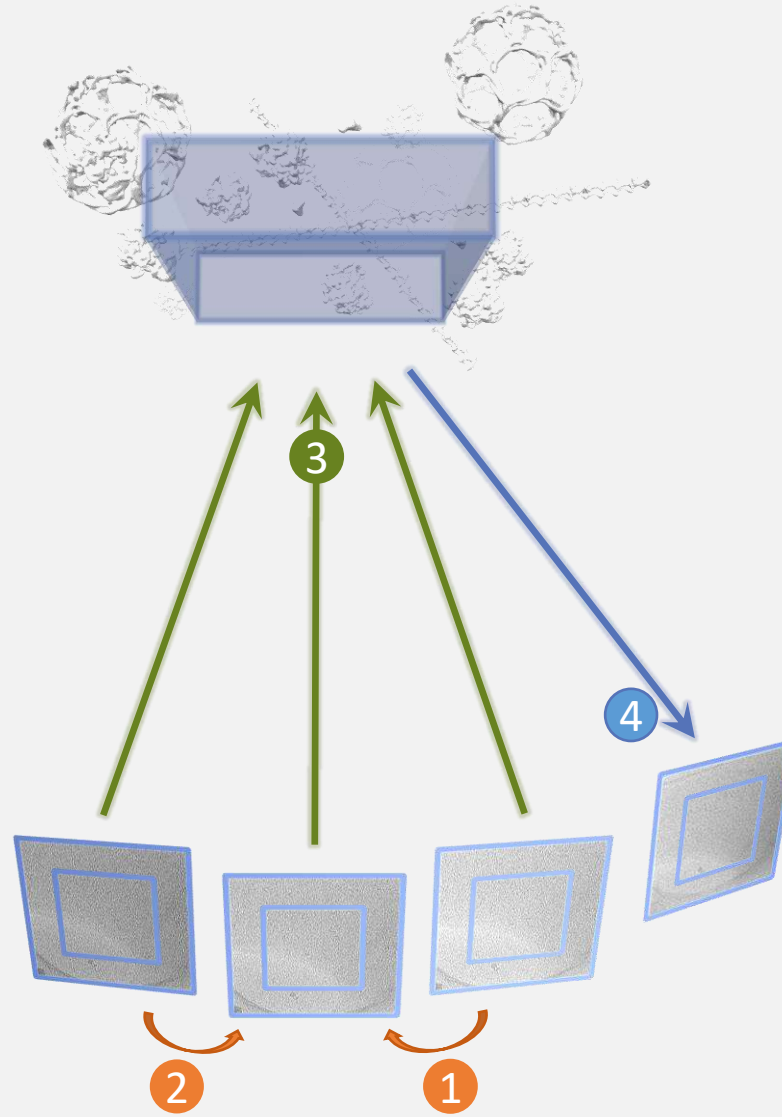


- Nearest-neighbor correlation
- Weighted back-projection

alignment thickness = z  Volume to be re-projected




Protomo alignment

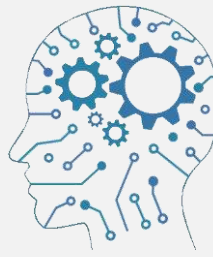


● Nearest-neighbor correlation

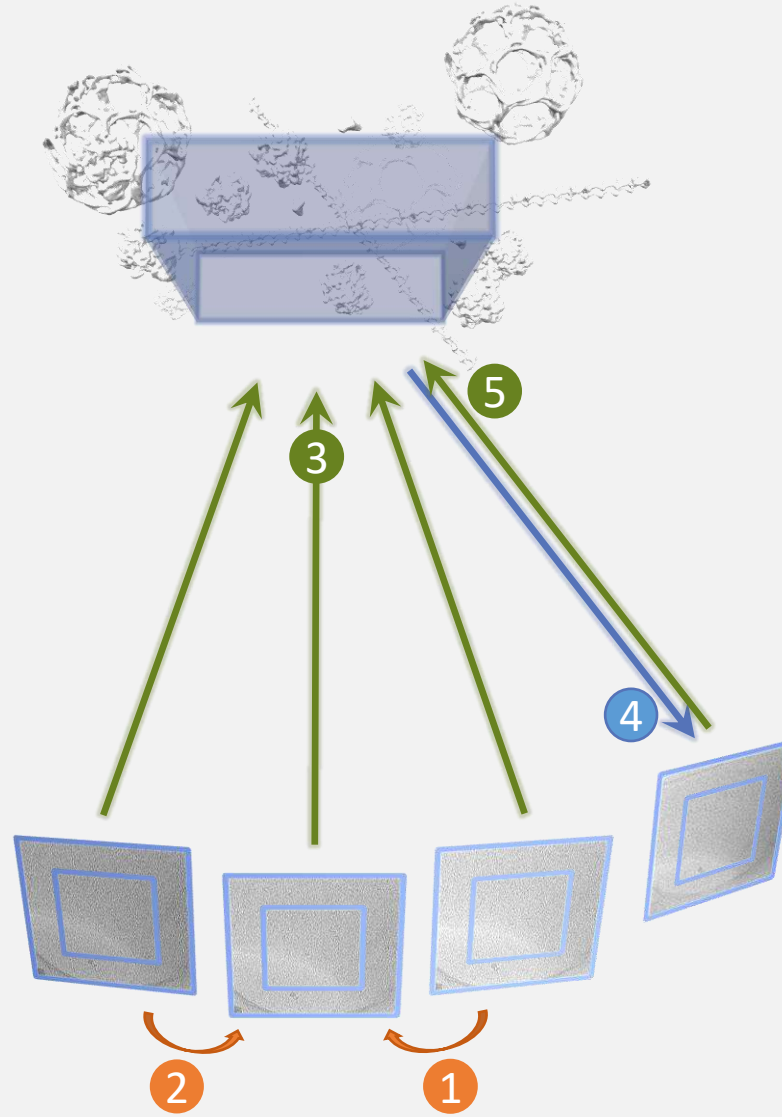
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

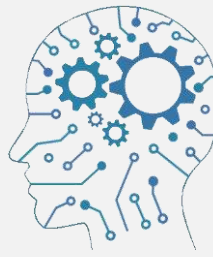


● Nearest-neighbor correlation

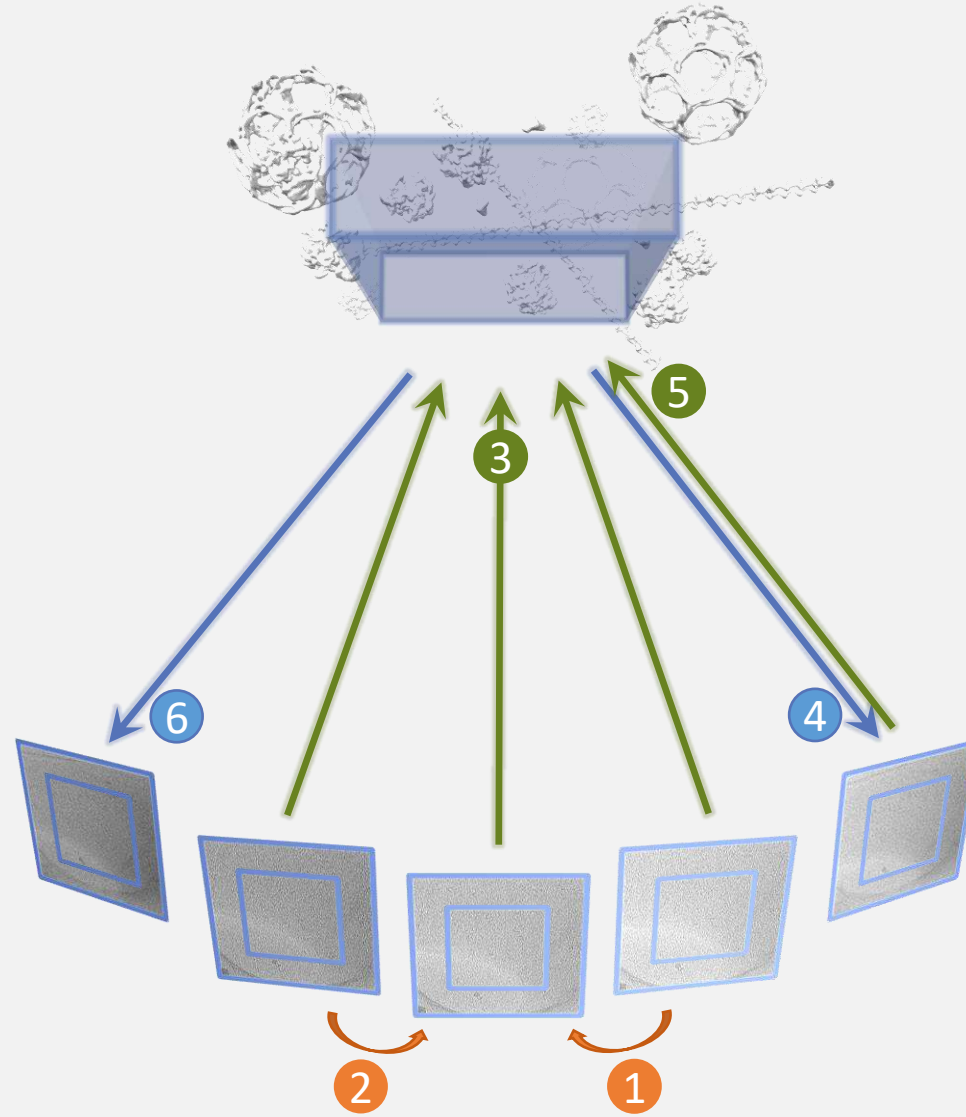
● Weighted back-projection

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


Protomo alignment

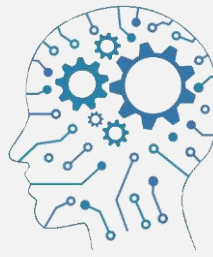


● Nearest-neighbor correlation

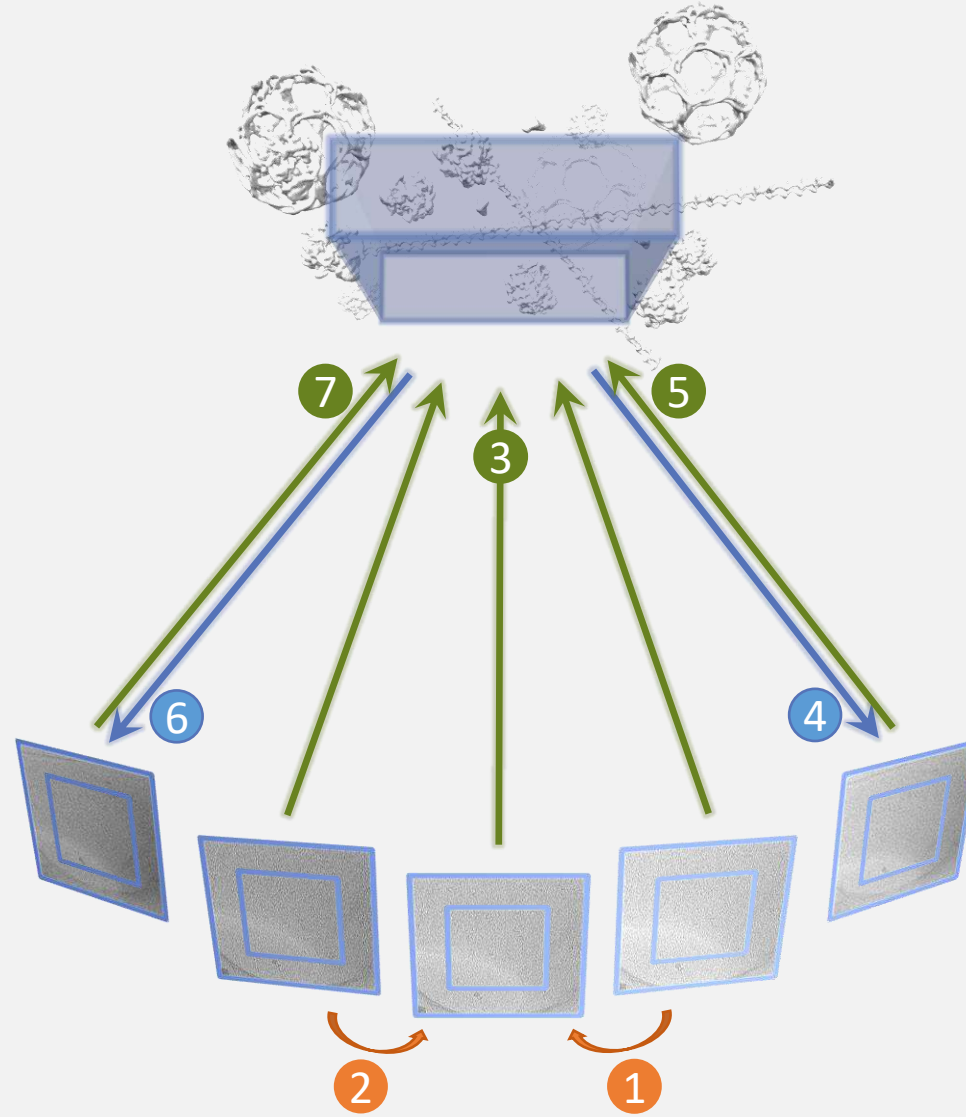
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

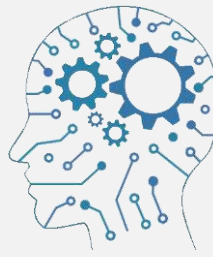


● Nearest-neighbor correlation

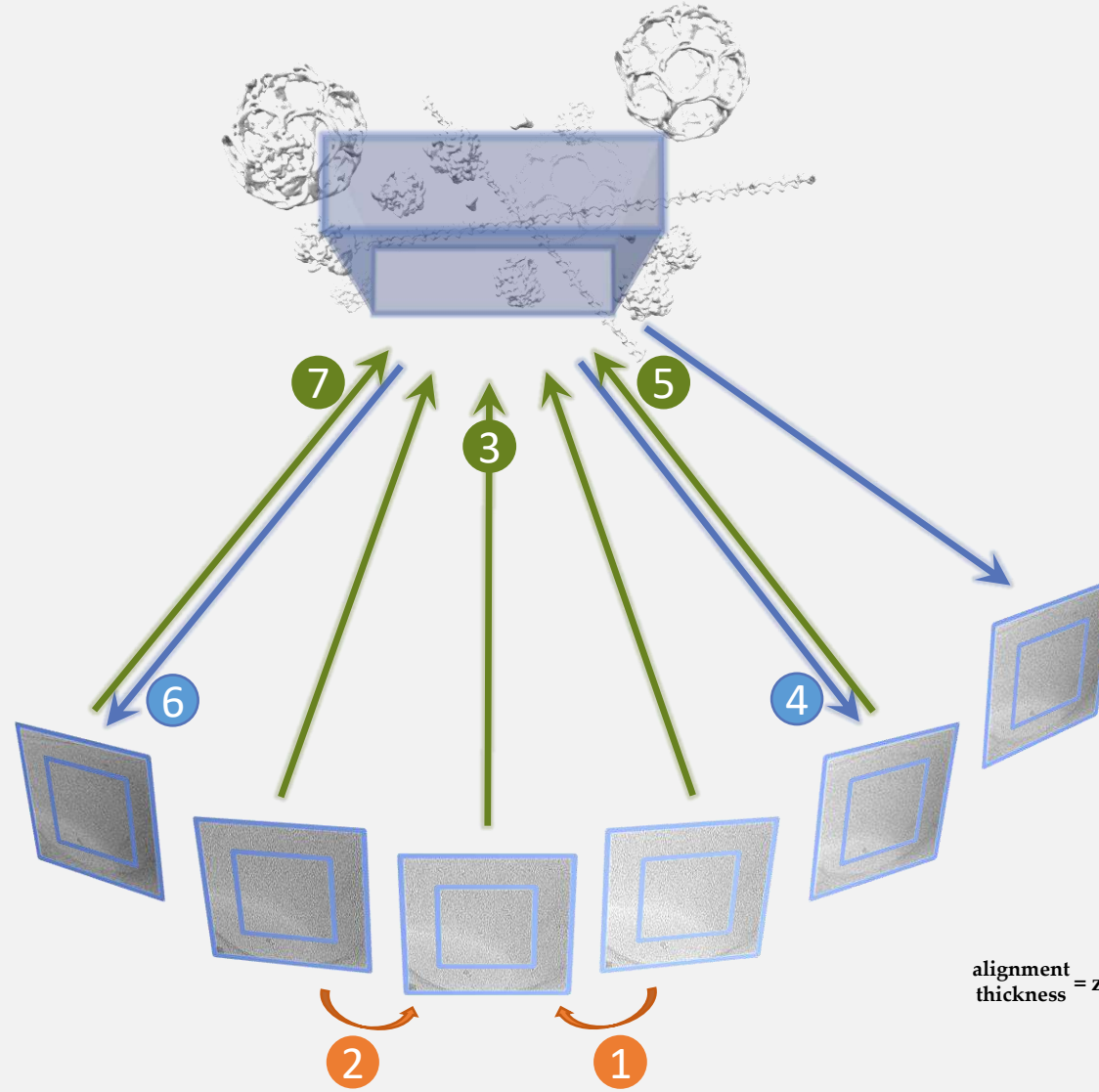
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

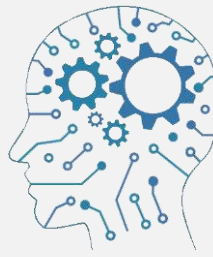


● Nearest-neighbor correlation

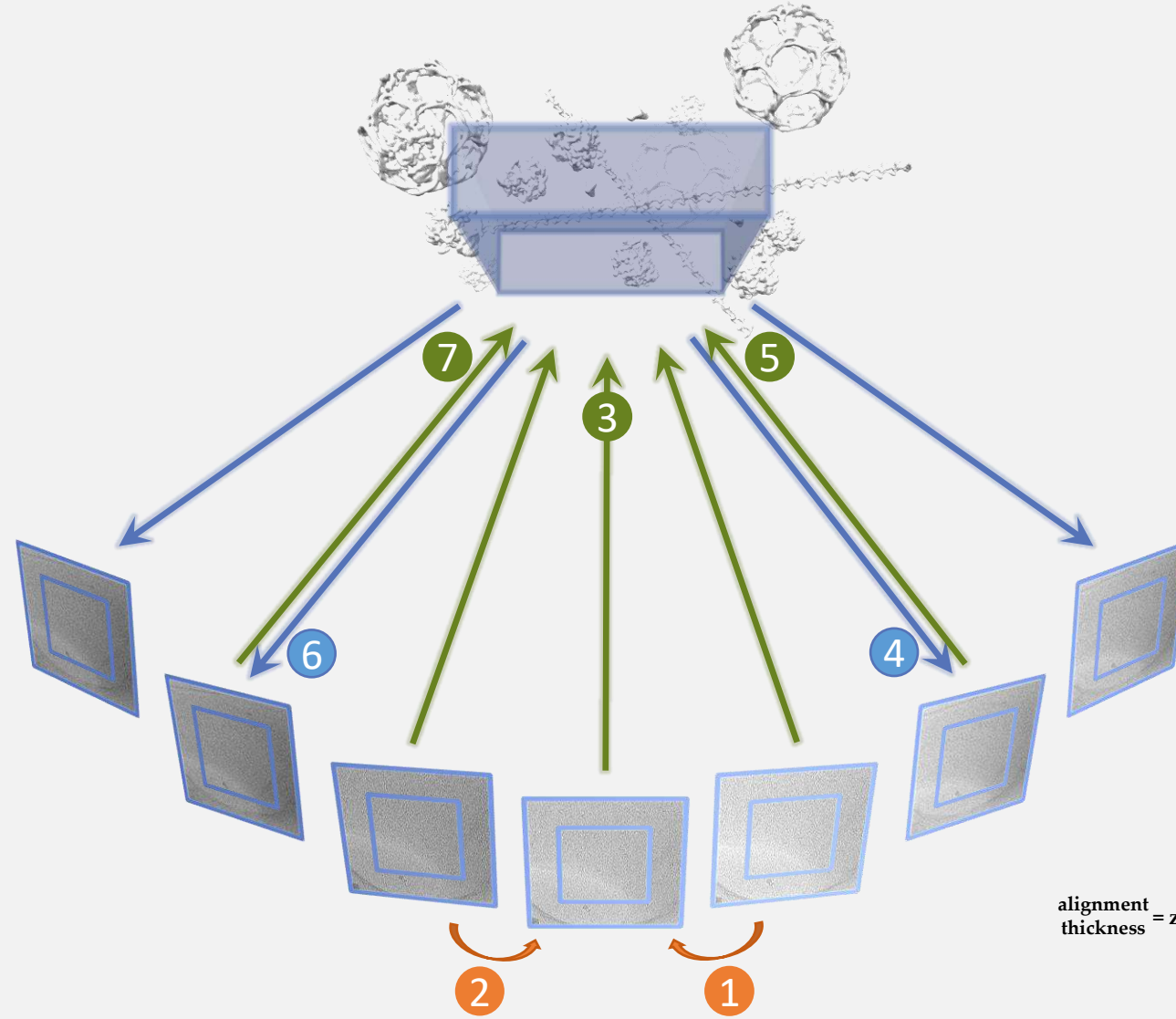
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment



● Nearest-neighbor correlation

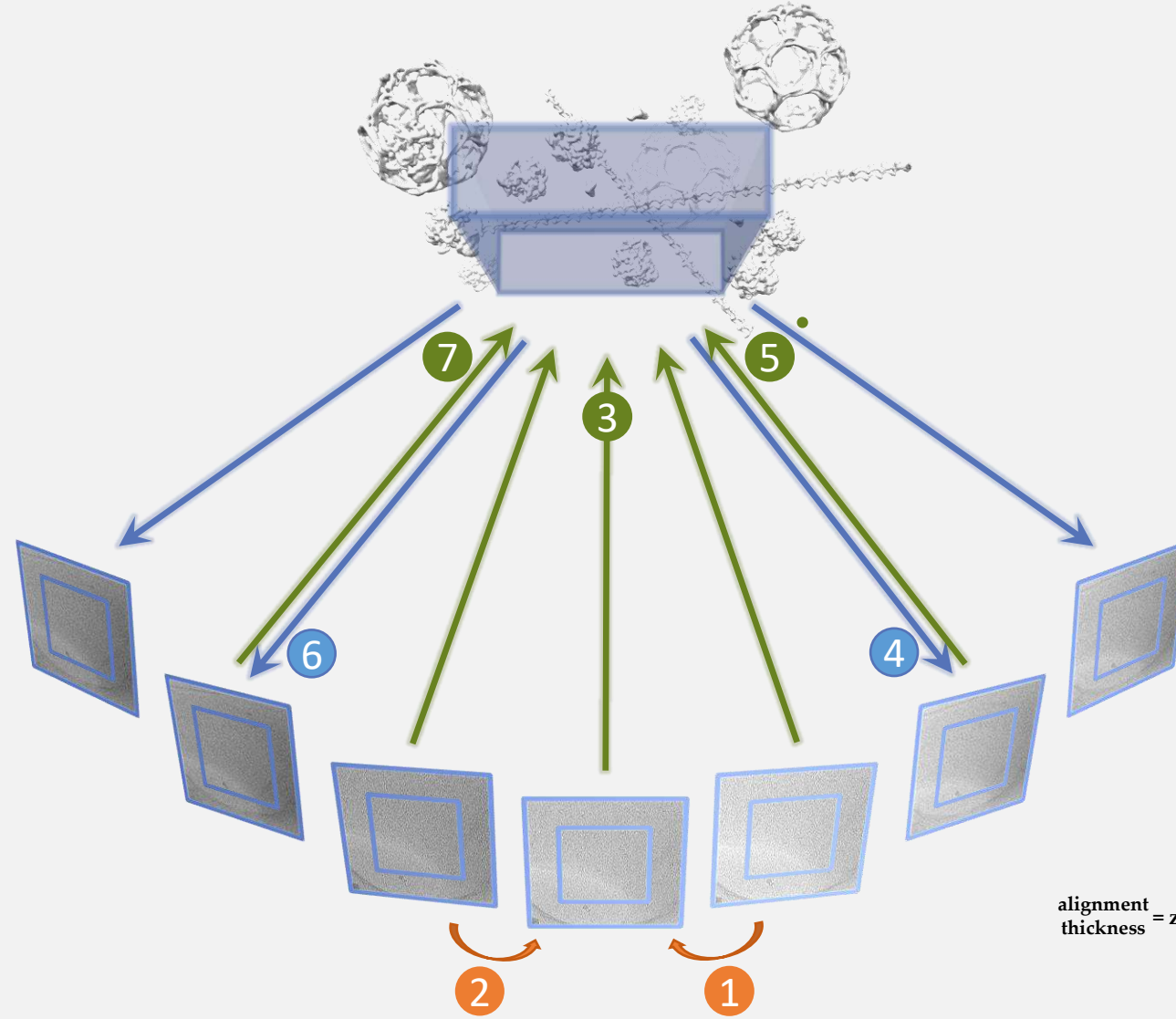
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

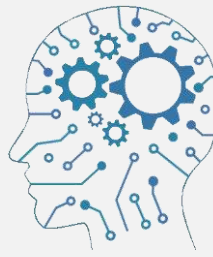


● Nearest-neighbor correlation

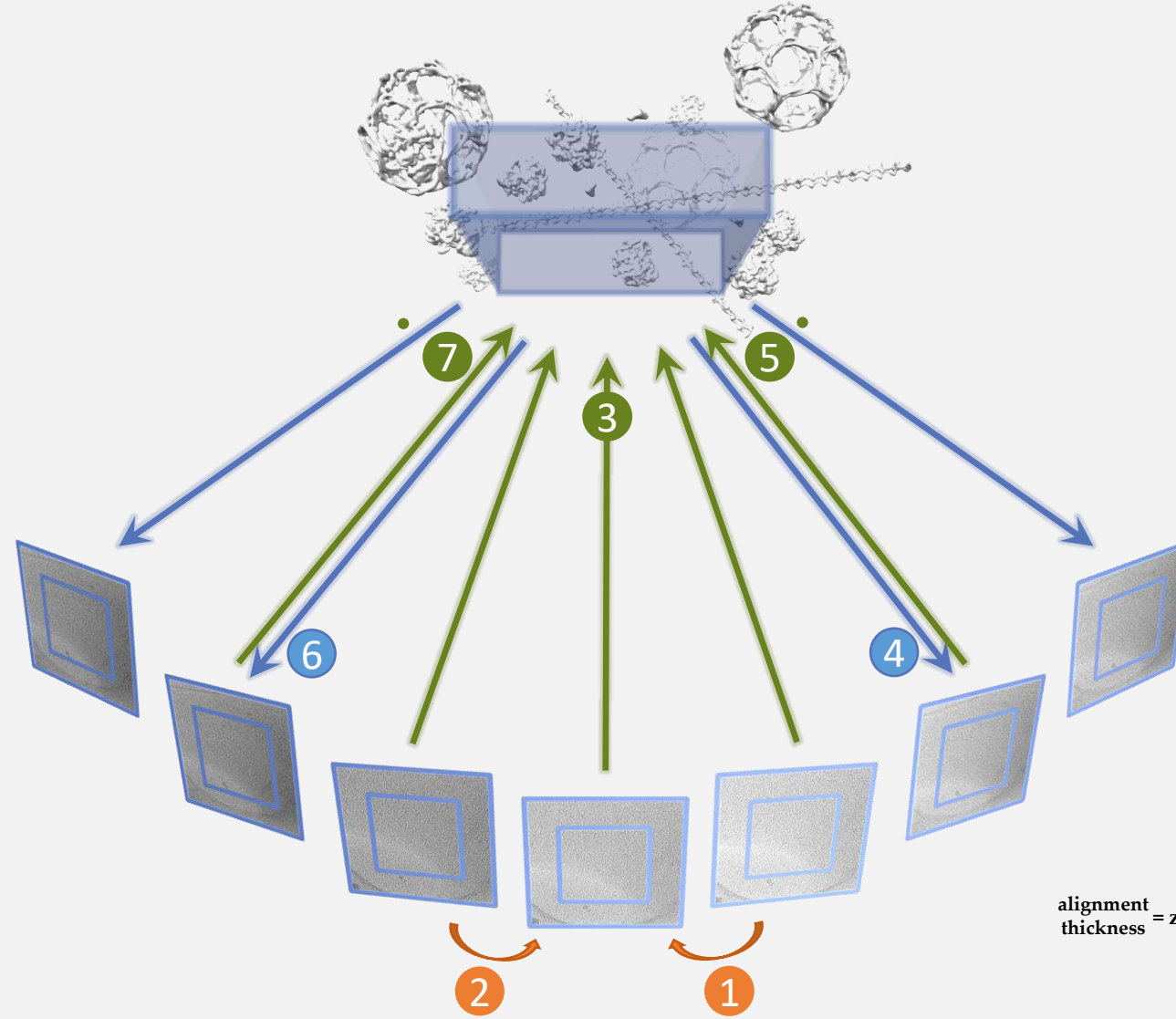
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

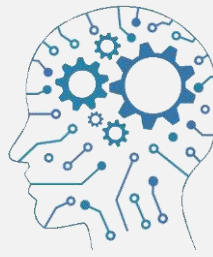


● Nearest-neighbor correlation

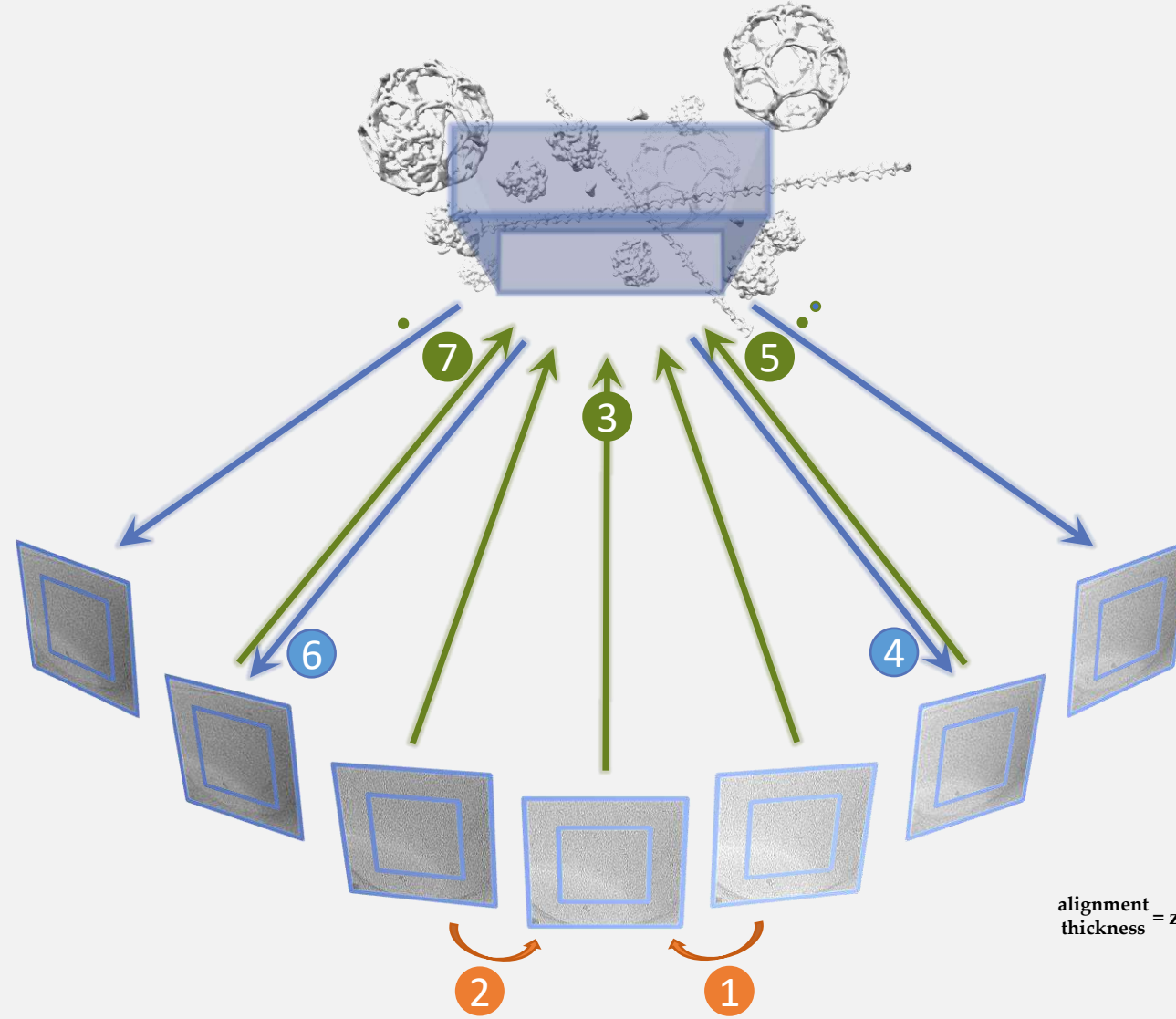
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

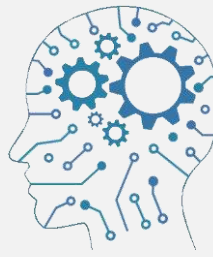


● Nearest-neighbor correlation

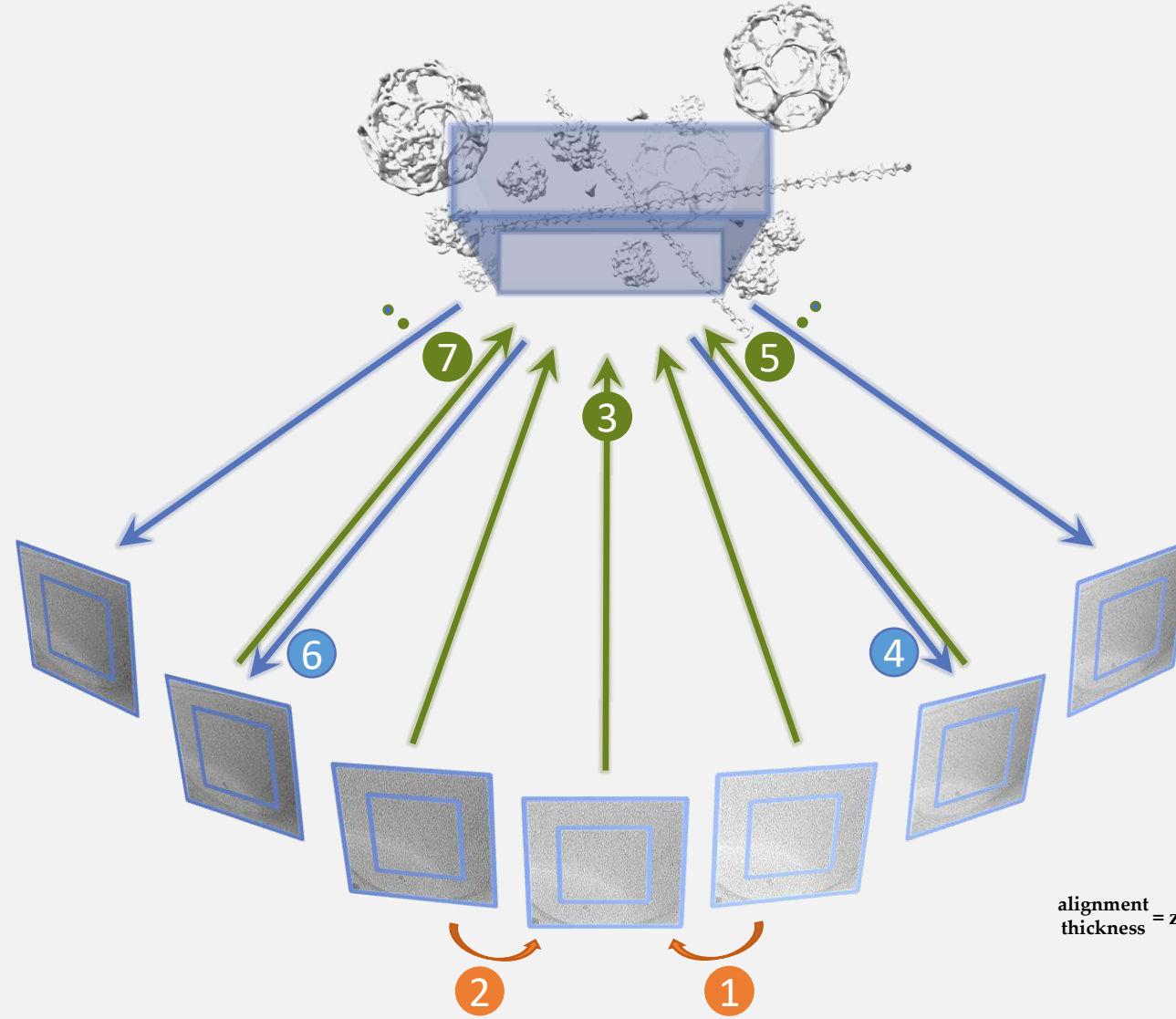
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

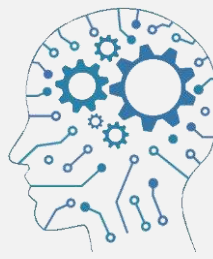


● Nearest-neighbor correlation

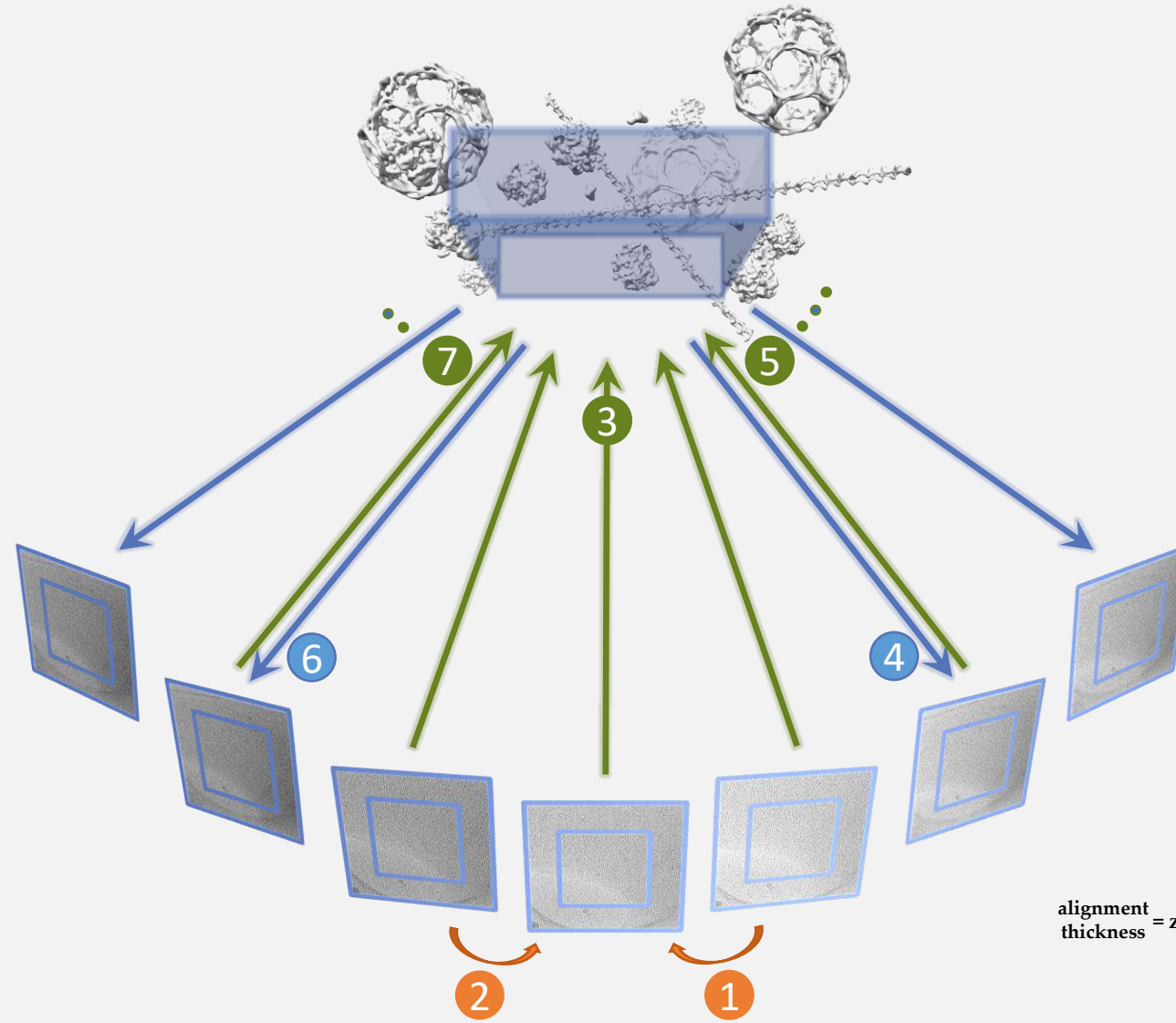
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

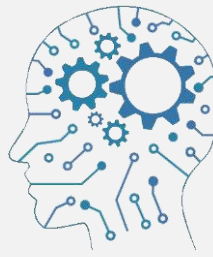


● Nearest-neighbor correlation

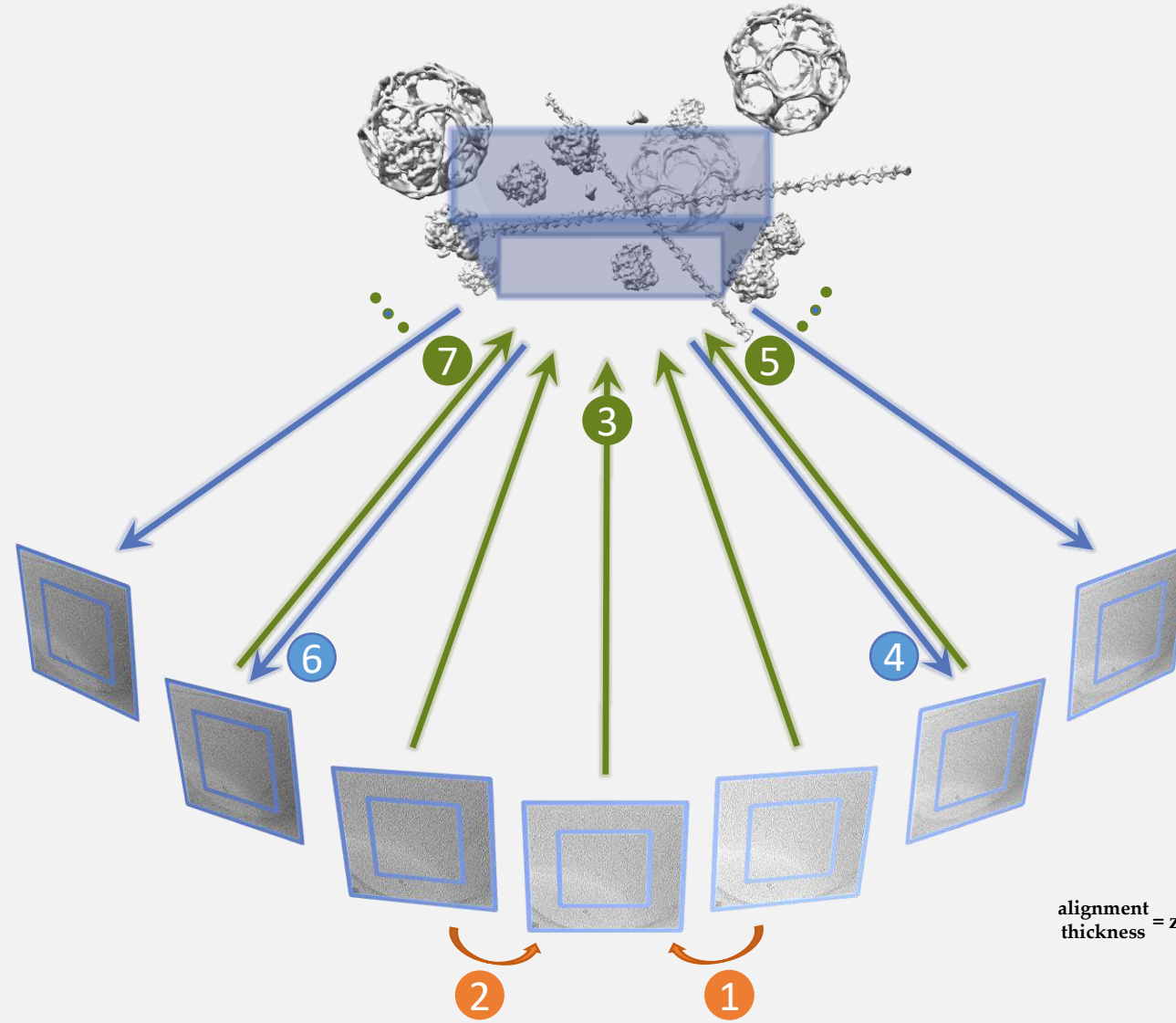
● Weighted back-projection

alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation




Protomo alignment

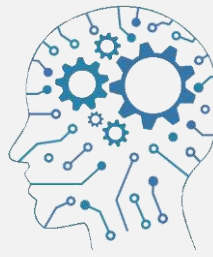


● Nearest-neighbor correlation

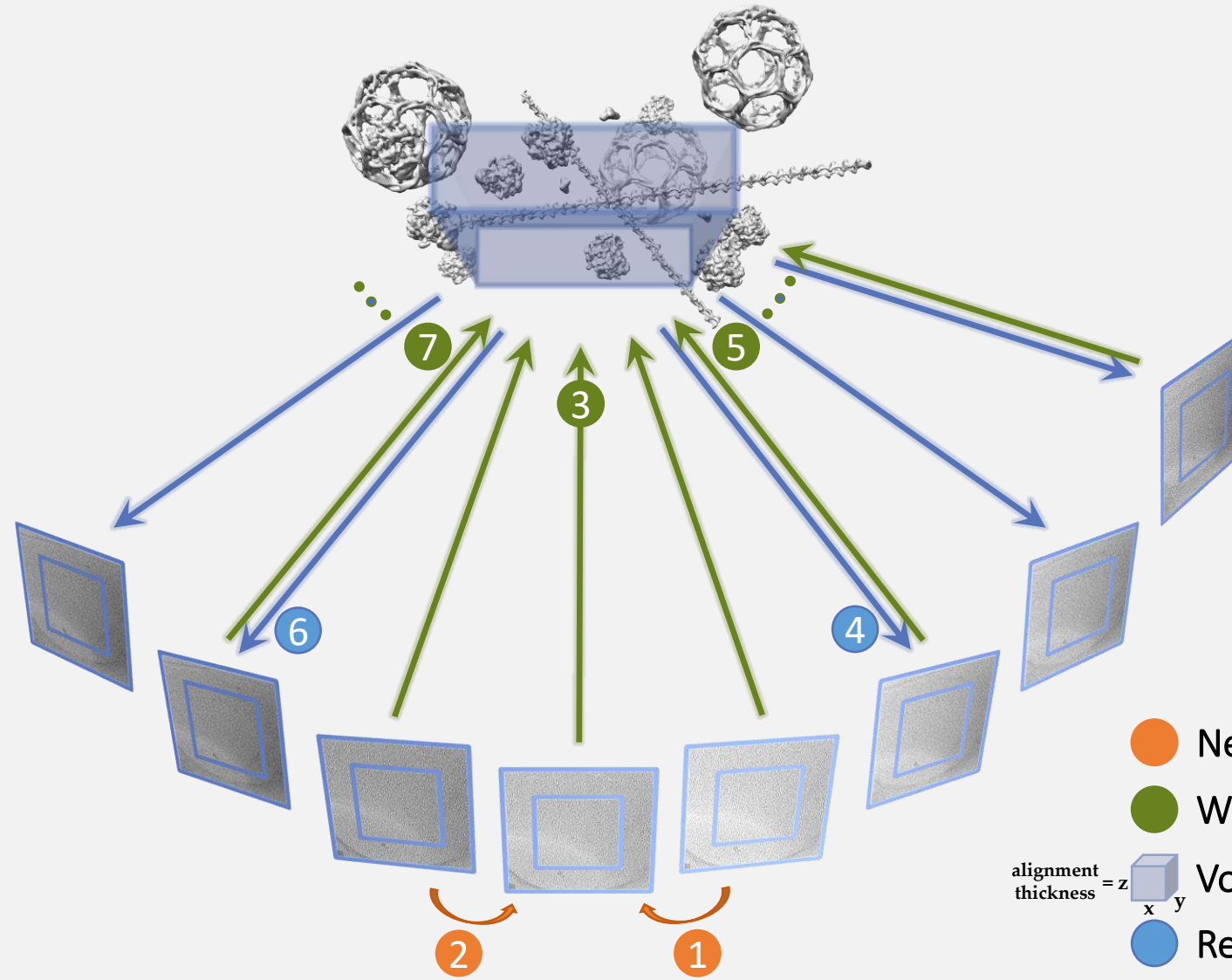
● Weighted back-projection

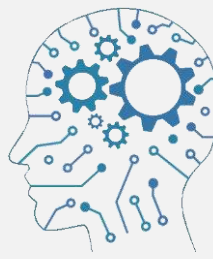
alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation

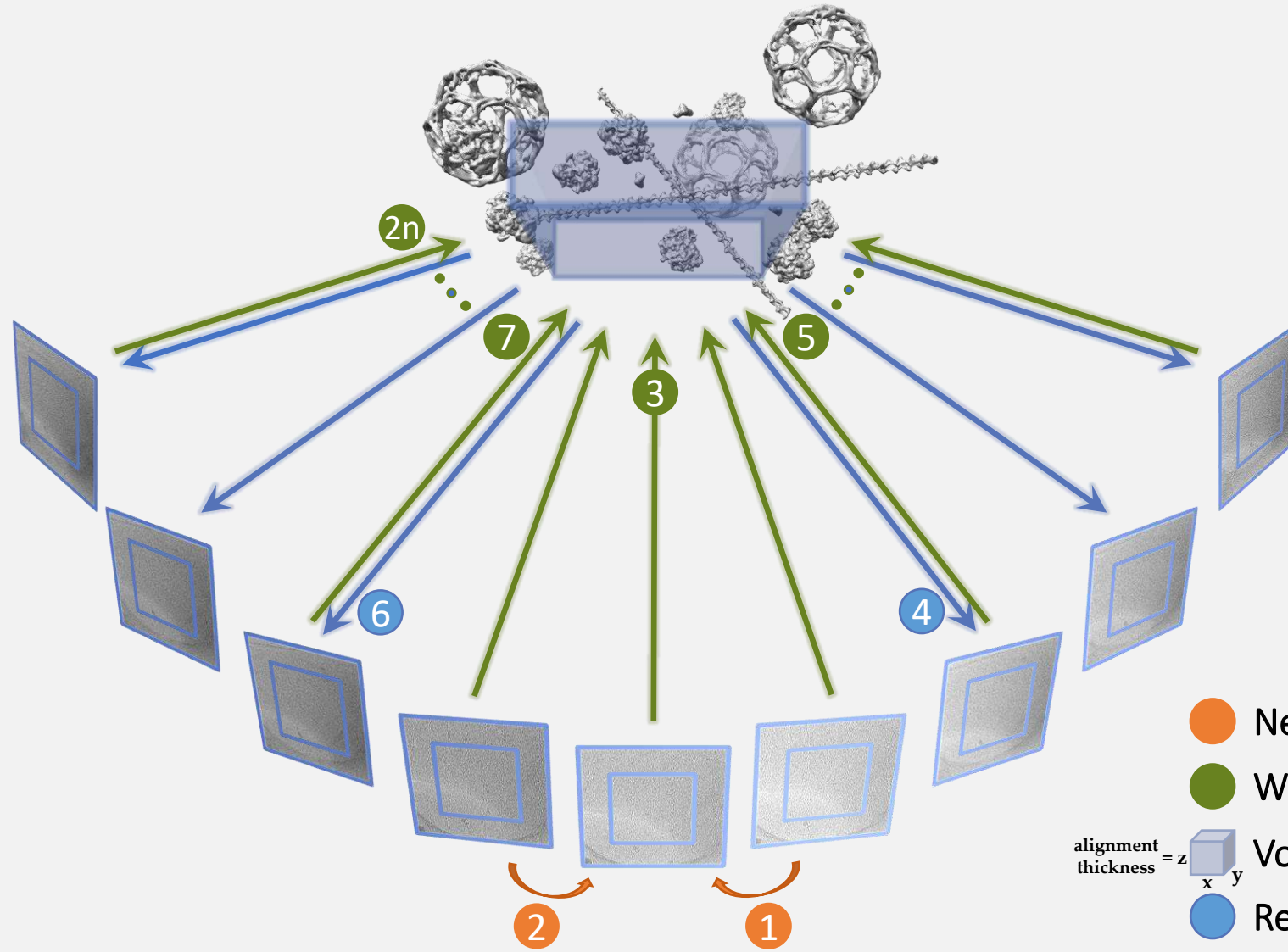


Protomo alignment






Protomo alignment

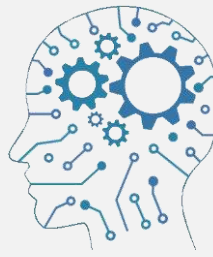


● Nearest-neighbor correlation

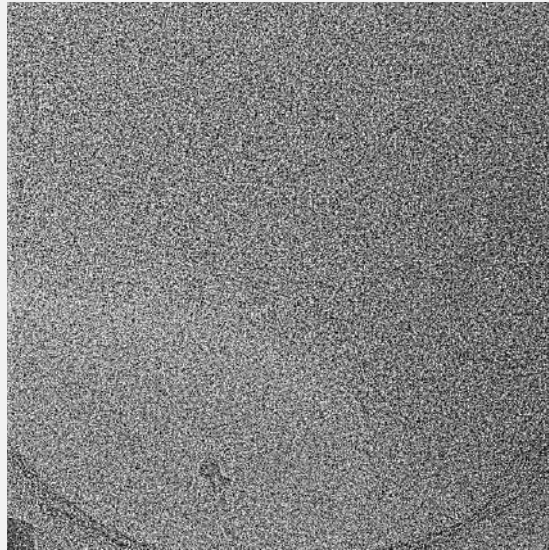
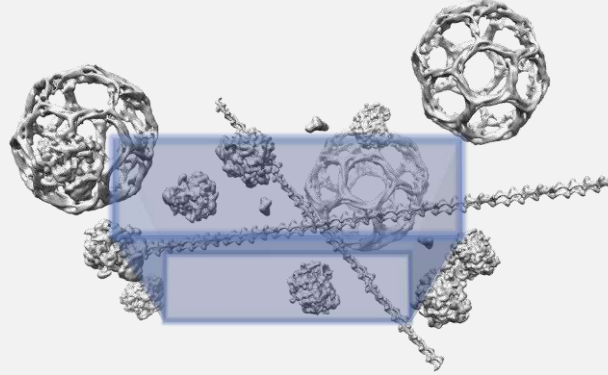
● Weighted back-projection

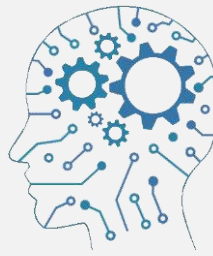
alignment thickness = z  Volume to be re-projected

● Re-projection \rightarrow correlation

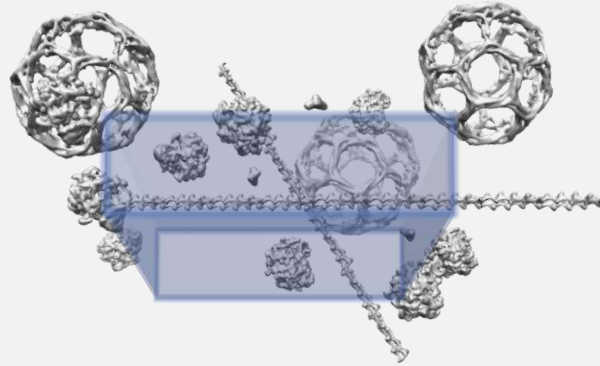


Protomo alignment

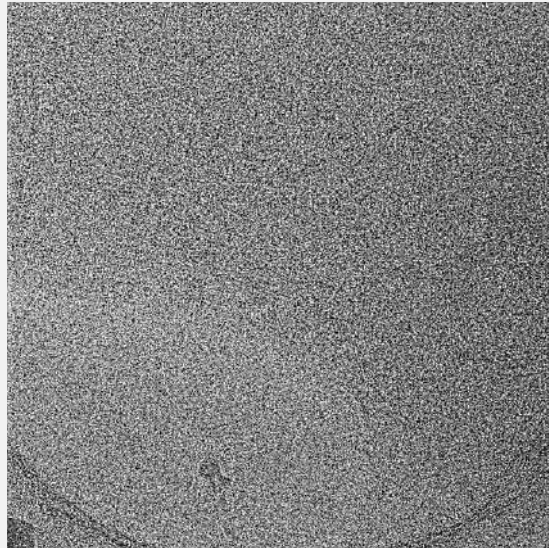


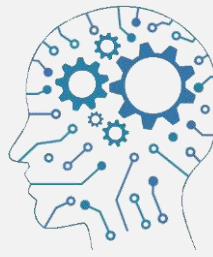


Protomo alignment

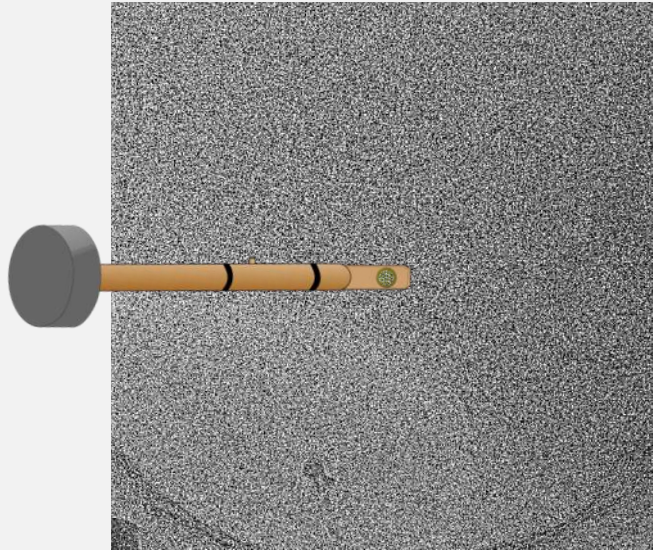
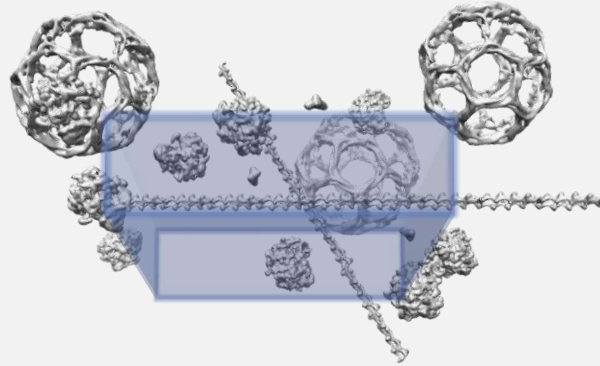


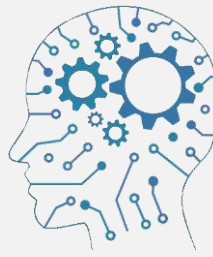
Refine orientations
of objects



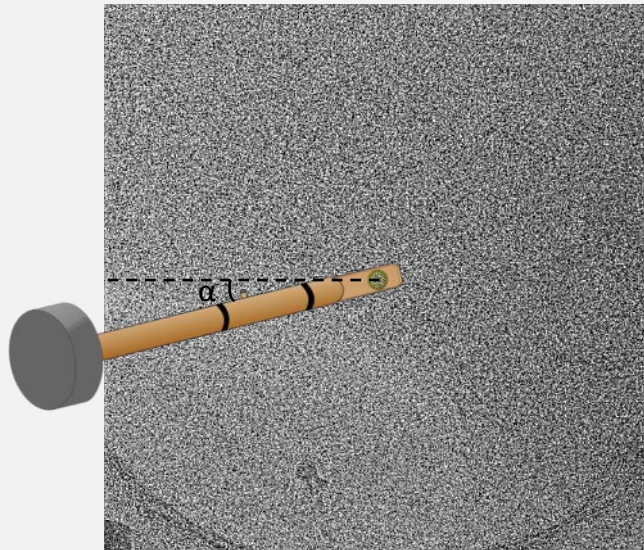
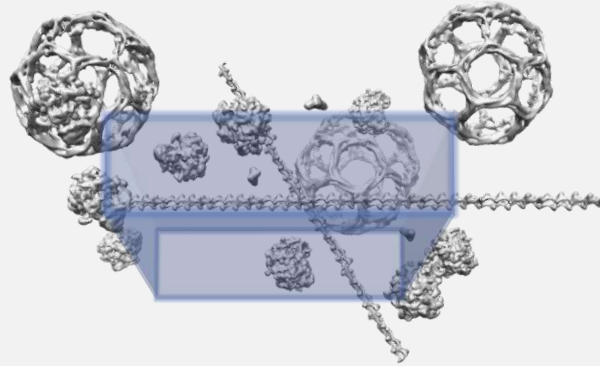


Protomo alignment

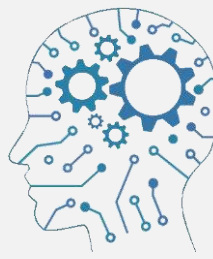




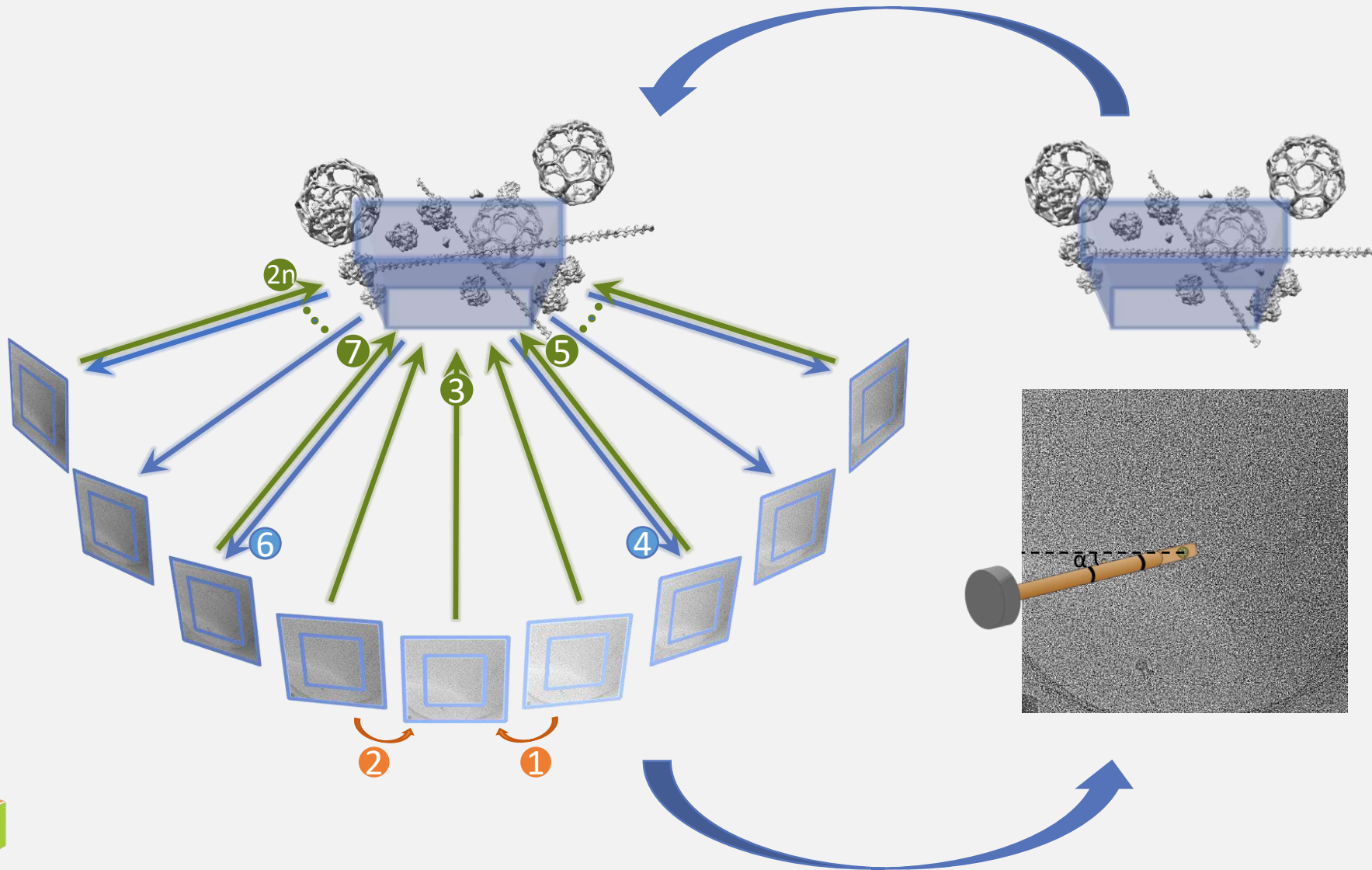
Protomo alignment



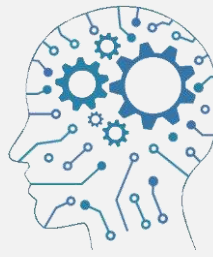
Refine tilt azimuth



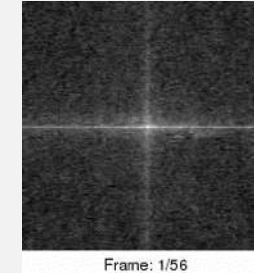
Appion-Protomo refinement



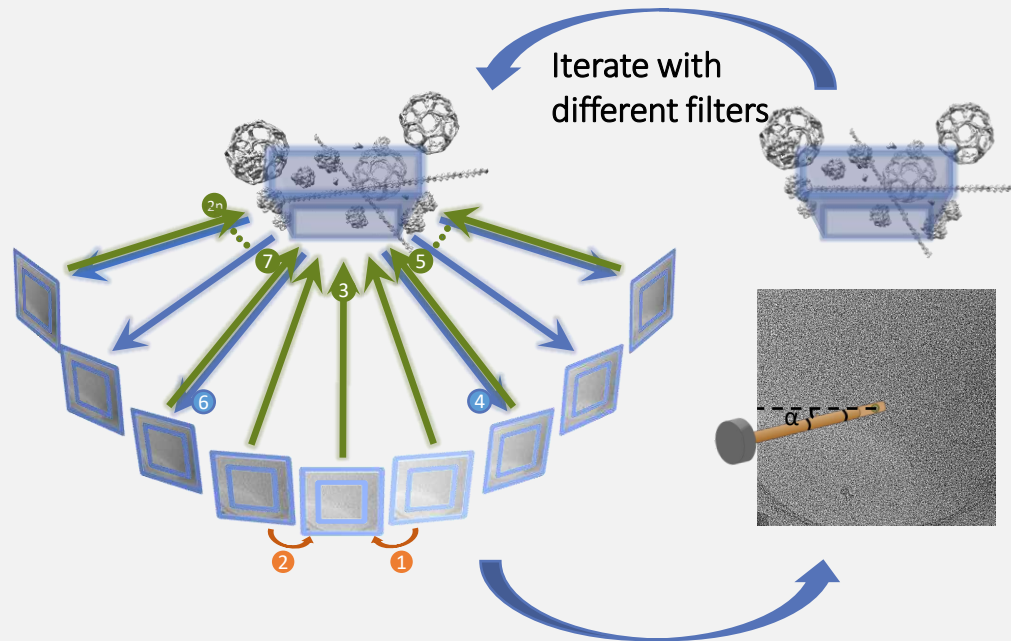
Iterate with
different filters



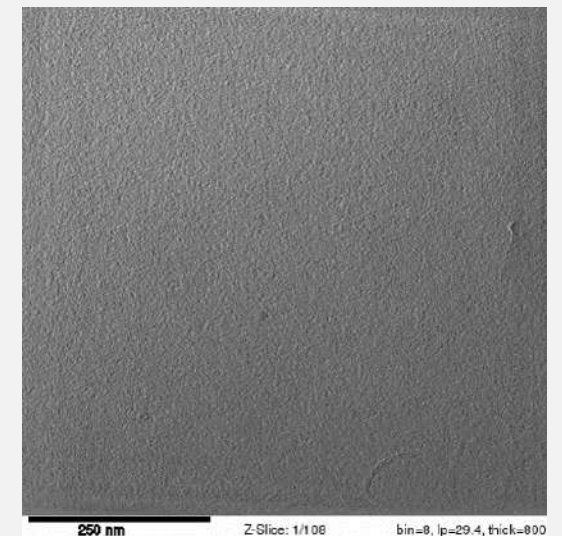
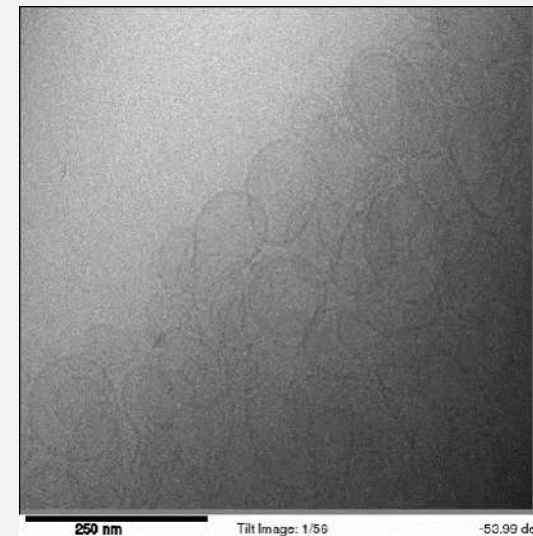
Standard nearest-neighbor alignment



Why is this important?



After Protomo refinement



Questions?

Next: Appion-Protomo hands-on

